

## Morphological Study During Annual Reproductive Cycle of Maleleaf-Nosed Bat, *Hipposiderospeoris* Schneider; Chiroptera

Janbandhu Kishor Sukhadeo

Department of Zoology,  
Institute of Science, R.T. Marg, Nagpur (M.S.) India

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**Abstract:** *Hipposiderospeoris* shows an annual reproductive cycle. The organization of the male reproductive organs shows commencement of spermatogenesis during September-October; Peak spermatogenesis, spermiogenesis and spermiation in December; Regression of testis with storage of sperms in cauda epididymis in February and complete involution of testis and storage of sperms in cauda epididymis in April. The accessory glands in the males remain in an active state for about two months after the regression of the testis. The male accessory glands - the seminal vesicles, prostate and bulbo-urethral glands came to activity in October and remained in an active state until the beginning of April. The active status of the accessory glands and the healthy condition of the epididymal sperms in this bat are physiologically linked together and these appeared to be independent of the factors which maintained spermatogenetic activity of the testis.

**Key words:** Accessory glands • Anatomy • Epididymis • Testis

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### INTRODUCTION

The Order Chiroptera comprises of two Suborders, Megachiroptera and Microchiroptera. Family Rhinolophidae of Microchiroptera comprises two subfamilies- Rhinolophinae and Hipposiderinae. The review on the prolonged survival of spermatozoa in bats observed that reproductive tracts of several species of vespertilionid and rhinolophid bats store sperms in the uterus or oviducts of females and in the caudaepididymides and ductus deferens of males [1].

The study of reproductive cycle in the male noctule *Nyctalusnoctula* observed that spermatogenesis occurs during summer accompanied by a thirteen fold increase in the mean weight of the testes. Spermatozoa were stored in the caudaepididymidis for the duration of winter and no spermatozoa were produced in this period. The lowest weight of caput and caudaepididymidis was recorded in June. The maximum weight of the cauda epididymis was recorded in September, coinciding with maximum sperm content. While the weight of the caput epididymis falls progressively throughout winter and few residual

spermatozoa were observed during November with the weight of the cauda epididymis being maintained throughout winter [2].

Reproduction in the Canyon bat, *Pipistrellushesperus* observed that the male spermatogenetic cycle is initiated in late June and sperms were available for ejaculation in September [3]. The reproductive biology of male leaf-nosed bat, *Macrotuswaterhousii* has shown that the spermatogenic cycle is initiated in June, sperm were available in August [4]. The caudaepididymides of hibernating male *Myotis lucifugus* and *M. velifer* were packed with spermatozoa from September-October and April-May despite involuted testes [5].

While studying the reproductive cycle in the male fruit bat, *Cynopterus sphinx* the active spermatogenesis was observed in October - November and mid-January-April [6]. In *Rhinolophuscapensis*, spermatogenesis occurred between October and May (spring to autumn) and the sperm being released to the cauda epididymis in April and May [7]. While male bat, *Rhinopomahardwickeihardwickei* showed annual

reproductive cycle and the testes were permanently abdominal [8] whereas the seasonal chronological events of the reproductive cycle and changes in structure and function of accessory sex organs of *Taphozous longimanus* at Varanasi [9]. The study is based on the annual variation in the anatomy and histological structure of reproductive system of *Hipposideros speoris* as a mammalian example.

## MATERIALS AND METHODS

The present work is based on the study of 48 males of *Hipposideros speoris* collected with the help of a butterfly net from underground dark places nearby Chandrapur, Maharashtra (India). The collection was made throughout the year from Sept-2008 to Aug-2009. 2 bats were collected once in a fortnight. The animals were anaesthetized with mild clinicleanaesthesia. The reproductive systems from males dissected out, displayed and photographed for anatomical examination.

## RESULTS AND DISCUSSION

*Hipposideros speoris* shows an annual reproductive cycle. In the adults the pubic capsule is large, thick, blackish pink in appearance and loose with external wrinkles (Fig. 1), while in the sub-adults, it is thick, tightly organized and pinkish with greasy appearance. The penis in sub-adult males is long and thin with its tip directed caudally while in the adult males the penis increases in thickness with the tip directed cranially. On the basis of the appearance of the pubic capsule, adults are differentiated from sub-adults.

The organization of the male reproductive organs shows commencement of spermatogenesis during September-October (Fig. 2); Peak spermatogenesis, spermiogenesis and spermiation in December (Fig. 3); Regression of testis with storage of sperms in cauda epididymis in February (Fig. 4) and complete involution of testis and storage of sperms in cauda epididymis in April (Fig. 5). The accessory glands in the males remain in an active state for about two months after the regression of the testis. There is an unbalanced sex-ratio with the females outnumbering the males.

In the present bat, spermatogenesis commenced in September and continued until the second week of January, after which the testes underwent regression and there is complete cessation of spermatogenetic activity.

However, the cauda epididymis is full of healthy spermatozoa until about the first week of April [2 and 5] after which there is a sudden fall in their population and most underwent degenerative changes.

The male accessory glands - the seminal vesicles, prostate and bulbo-urethral glands came to activity in October and remained in an active state until the beginning of April. The active status of the accessory glands and the healthy condition of the epididymal sperms in this bat are physiologically linked together and these appeared to be independent of the factors which maintained spermatogenetic activity of the testis. Alternately the factors which triggered the onset of sexual activity in the males of *H. speoris*, continue to operate on the accessory glands even after the cessation of spermatogenetic activity in the testes (Figs. 1-5). *H. speoris* developed the mechanism to overcome the disadvantages of low fecundity in the females, to ensure that all adult females do conceive during the breeding season. Similar observations were reported in *H. speoris* [10].

The testes are ellipsoidal in shape and post-abdominal in position. Both in the adult as well sub-adult males, the testis are enclosed in a thick muscular sac - the pubic capsule [11, 12]. The epididymis closely abuts the testis on its dorso-medial aspect and is recognized into distinct caput and cauda regions and a thin inconspicuous corpus epididymis. The ductus deferens which emerges from the cauda epididymis is short in length and it enters the abdominal cavity along with the components of the spermatic cord. In the abdominal cavity, the ductus deferens leaves the other components of the spermatic cord (internal spermatic nerve, spermatic artery and spermatic vein), passes mesially and loops dorsal to the ureter and joins the ampullae of Henle of the respective side. The male accessory reproductive organs comprise of a paired ampullae of Henle, a prostate, paired Cowper's glands and urethral glands. The accessory reproductive organs are all distinctly cyclic varying in gross size, weight, histology and secretory activity [12, 13].

### Abbreviations:

T: Testis; e: Epididymis; Pb.c.: Pubic Capsule; U: Urinary bladder; am.H: Ampulla of Henle; Cg: Cowper's gland; P: Penis; Vd: Vas deferens; Cpe: Caput epididymis; Cde: Cauda epididymis; Pr: Prostate; Sperm storage in epididymides (arrow).

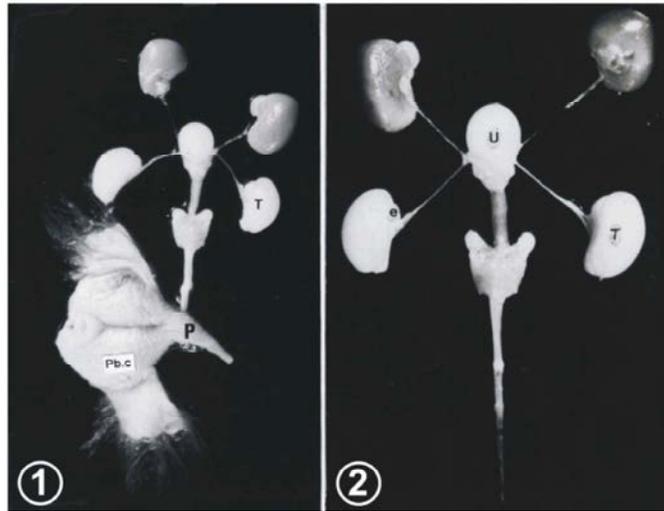


Fig. 1: Photograph of the ventral view of male reproductive system of *H. speoris* during commencement of breeding season with the intact public capsule

Fig. 2: Photograph of the ventral view of male reproductive system of *H. speoris* during commencement of breeding season after the removal of the pubic capsule Note the large size of the testis as compared to the epididymis and the large penis

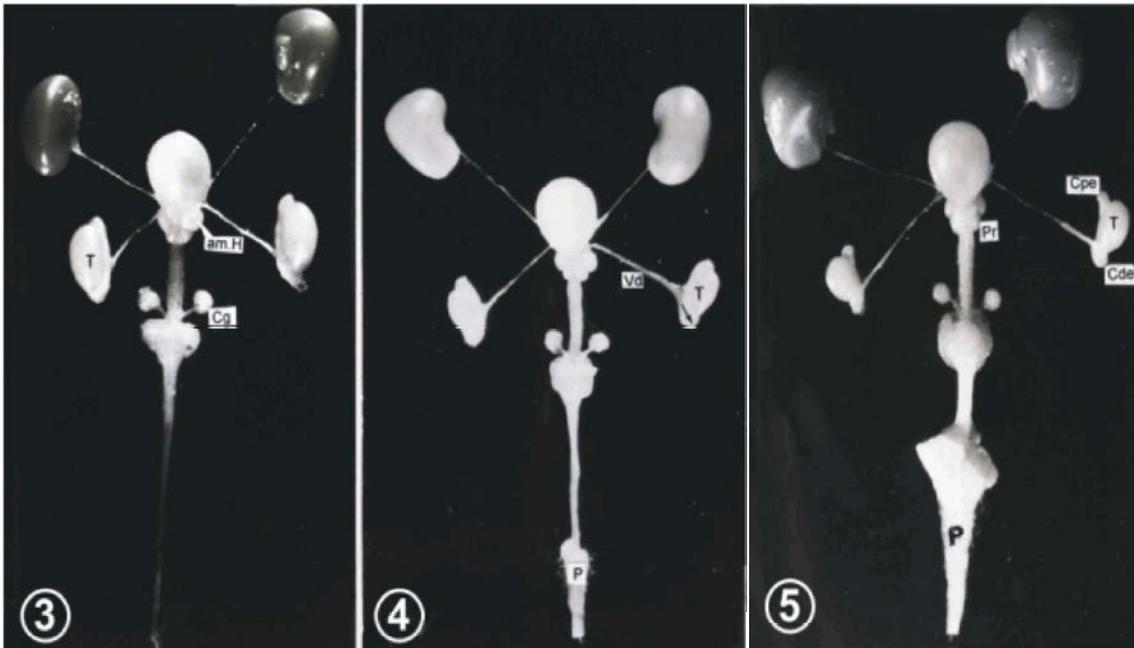


Fig. 3: Photograph of the dorsal view of the male genitalia of *H. speoris* shows peak activity of the testes. Note, the vas deferens joins the ampulla of Henle of the respective side. The duct of the Cowper's gland joins the urethra dorsally

Fig. 4: Photograph of the dorsal view of the testes and accessory reproductive organs of *H. speoris* shows regression of the testes with storage of sperm in the cauda epididymides. Note, the size of the testis is reduced. Epididymides get swollen due to sperm storage

Fig. 5: Photograph of the male genitalia of *H. speoris* shows involuted testes - with storage of spermatozoa in the cauda epididymides. Note, the testis size is further reduced as compared to February and the cauda epididymis is extended beyond the margin of testis as it is packed with sperms

## CONCLUSION

In *H. speoris* spermatogenesis continues from September to second week of January, followed by regression and complete cessation of spermatogenetic activity. The cauda epididymis contain healthy spermatozoa until the first week of April [2 and 5] after which there is a sudden decrease in number and most of the spermatozoa underwent degenerative changes.

## REFERENCES

1. Racey, P.A., 1975. The prolonged survival of spermatozoa in bats. In the Biology of the Male Gamete, pp: 385-416. Eds. J.G. Duckett and P.A. Racey. Academic Press, London.
2. Racey, P.A., 1974. The reproductive cycle in male noctule bats *Nyctalus noctula*. J. Reprod. Fert., 41: 169-182.
3. Krutzsch, P.H., 1975. Reproduction of the canyon bat, *Pipistrellus hesperus*, in South-Western United States. Am. J. Anat., 143: 163-200.
4. Krutzsch, P.H., R.H. Watson and C.D. Lox, 1976. Reproductive biology of the male leaf-nosed bat, *Macrotus waterhousii* in South-western United States, Anat. Rec., 184: 611-636.
5. Krutzsch, P.H., E.G. Crichton and R.B. Nagle, 1982. Studies on prolonged spermatozoa survival in Chiroptera: A Morphological examination of storage and clearance of Intrauterine and caudaepididymal spermatozoa in the bats, *Myotis lucifugus* and *M. velifer*. The American Journal of Anatomy, 165: 421-434.
6. Krishna, A. and C.J. Dominic, 1984. Reproductive cycle in the male fruit bat, *Cynopterus sphinx* (Vahl, 1797). Lynx (Praha), n.s. 22 CS. 19-26.
7. Bernard, R.T.F., 1985. Reproduction in the Cape horseshoe bat (*Rhinolophus capensis*) from South Africa. South African Journal of Zoology, 20: 129-135.
8. Banerjee, S. and K.B. Karim, 1986. Male reproductive cycle in the Indian Mouse-tailed bat, *Rhinopomahardwickei hardwickei*. J. Curr. Biosci., 3(4): 168-175.
9. Singh, U.P., 1997. Reproductive biology of the male sheath-tailed bat, *Taphozous longimanus* (Emballonuridae) from India. Biomed. Environ. Sci., 10(1): 14-26.
10. Gopalakrishna, A. and D. Bhatia, 1980. Storage of spermatozoa in the epididymis of the bat, *Hipposideros speoris* (Schneider). Curr. Sci., 49: 951-953.
11. Brosset, A., 1962. The bats of Central and Western India, Part II. Journal of the Bombay Natural History Society, 59: 583-624.
12. Bhatia, D., 1980. Sex-cycle and associated phenomena in the bat, *Hipposideros speoris* (Schneider) from Maharashtra. Ph.D. thesis, Nagpur University, India.
13. Pal, A.N., 1977. Studies on the male accessory reproductive organs in some Indian bats. Ph.D. thesis, Nagpur University, India.