# Incidence of *Listeria* Species in Farmed Tropical Fish in Khuzestan, Iran

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Abstract: The incidence of *Listeria* spp. was determined in freshly-harvested, farmed fish (grass carp and common carp) in Khozestan, South-West Iran. One hundred fish samples were purchased at the fish markets, in the warm and cold seasons. Fish samples? intestines and gills were examined for *Listeria* spp. 18 samples (18%) were positive for *Listeria* spp. isolation. In the warm season, 2 samples of Grass carp fishes were positive for *Listeria monocytogenes* and 3 and 4 samples of both grass carp and common carp fishes were positive for *L. grayi* and *L. innocua* respectively. In the cold season, again 2 samples of grass carp fishes were positive for *L. monocytogenes* and 7 samples for *L. innocua*. no common carp samples were contaminated with *Listeria* spp. This frequency of contamination of fresh farmed fish with *L. monocytogenes* in Khuzestan is a concern and the handling and consumption of these fish, either raw or undercooked, may pose a health risk.

Key words: Incidence · Listeria · Tropical · Fish · Iran

## INTRODUCTION

Listeria spp. are widespread in the environment and commonly found in soil, sewage, dust and water. One particular species, Listeria monocytogenes, has been recognized as a food-borne pathogen since 1981. It is known as the causal agent of listeriosis, which is characterized by fever, chills, convulsions, headache, diarrhea and vomiting which are commonly flu-like symptoms [1]. The bacterium can grow in many foods, even at the refrigerator temperature. The organism has been isolated from freshwater and seawater fish and from frozen and processed sea food [2-7].

A number of food-borne outbreaks and sporadic cases of listeriosis has been reported mostly in North America and Europe [9]. Persons most susceptible to the disease are immunocompromised hosts such as pregnant women and the elderly.

L. monocytogenes and other Listeria spp. have been isolated from fishery products on a regular basis in tropical and temperate regions since 1980s. L. monocytogenes is considered as a psychrotrophic pathogen and may thus be less common in tropical water. Ben Embarek [10] reviewed the incidence of Listeria in seafood worldwide and found that the prevalence of L. monocytogenes varies from 4 to 12% in temperate areas.

Some reports [11-13] have suggested the absence of *L. monocytogenes* in tropical fishes, but Ben Embarek [10] emphasized that more research is needed regarding the incidence of *L. monocytogenes* in tropical seafood. Adesiyun [14] noted a 2% incidence of *L. monocytogenes* in seafood samples from Trinidad. In India, the prevalence of *L. monocytogenes* in tropical fresh shellfish and finfish ranges from 12.1 to 17.2% [15]. In Latin America, the prevalence of *L. monocytogenes* in fish and fish products varies from 0 to 50% [16] and, in India, 9% of fresh/raw seafood samples are contaminated with *L. monocytogenes* [17]. Thus, it may be concluded that *Listeria* can be an important problem even for fisheries in tropical countries.

There are limited data regarding the prevalence of *L. monocytogenes* in seafood consumed in Iran and no information exists on the incidence of *Listeria* spp. in fresh farmed fish in Khuzestan province. *L. monocytogenes* was detected in 6.6 to 10% of common carp and silver carp samples and in 11.6 to 12.5% of farmed rainbow trout, in Tehran and Gilan province in the temperate north of Iran [18]. Other studies in Iran have reported low prevalence of *L. monocytogenes* in frozen and fresh fish samples ranging from 0 to 2.6% [19-20] but the incidence of the bacterium in traditional smoked salmon in Gilan is high, at 34% [21].

The purpose of the present study was to generate information on the incidence of *Listeria* spp. in the freshwater fish in Khuzestan province in South-West Iran, which has a tropical climate for at least 8 months of the year and is a major centre for the culture and export of freshwater fish.

### MATERIALS AND METHODS

**Samples:** During Nuvember 2008 to October 2009, one hundred fish samples, 50 common carp (*Cyprinus carpio*) and 50 grass carp (*Ctenopharyngodon idella*) were purchased at the local fish markets of Ahvaz City, Khuzestan, Iran. Fish samples were collected from the local fish farms and transported on ice to the fishmongers within 24 h of capture. All samples were placed in sterile bags, on ice, for transport to the laboratory and processed within 2 h of collection.

**Isolation and Identification of** *Listeria* **spp:** Samples were analyzed for Listeria spp. using the enrichment, selection and isolation protocol recommended by the US Food and Drug Administration [22]. For each sample, 5g of intestine and 2-3 gills were removed aseptically and placed separately in 20 ml of Listeria Enrichment Broth (LEB, Merck, Germany) containing potassium thiocyanate (37.5 g/L, Merck). In a class III safety cabinet, this mixture was homogenized and then incubated at 35° C. On the same day and after 24h and 48h, 100 iL samples from the enrichment culture were plated onto Listeria selective agar (Oxford formulation, Himedia, India) and incubated at 37° C for 48h. The plates were examined for typical Listeria colonies (translucent greenish yellow colonies) and at least 3 suspected colonies were sub-cultured on Brain Heart Infusion Agar (BHIA, Hispanlab) and incubated at 37° C for 48h. All the isolated colonies were characterized by Gram staining, motility at 25° C and 37° C, catalase and

oxidase tests and acid production from xylose and mannitol. For further confirmation; other biochemical reactions, â-hemolytic activity on 5% sheep blood agar (Merck) and CAMP test were performed according to Bergey's Manual of Systematic Bacteriology [23].

Serotyping of the Confirmed Listeria Isolates: Serotyping by slide agglutination was determined using Bacto-Listeria-O antisera types 1 and 4 (Difco, USA). Complete serotyping of confirmed isolates was performed in the Department of Microbiology, Faculty of Veterinary Medicine, University of Tehran, using "in house" production reagents as described elsewhere [24].

# RESULTS AND DISCUSION

Our study showed that 18 out of the 100 (18%) tested samples from grass carp and common carp were positive for Listeria spp (Table 1). In the cold season (From November to March), 2 isolates from grass carp fishes were L. monocytogenes and 7 isolates were identified as L. innocua. In the warm season (From April to October), 2 samples of grass carp fishes were L. monocytogenes and 3 and 4 samples of both grass carp and common carp fishes were positive for L. grayi and L. innocua respectively. L. innocua was detected in 11% of samples and was the predominant isolate among the Listeria spp. This agrees with the findings of Jalali and Abedi [20], Parihar et al. [17] and Dhanashree et al. [25]. Since both L. monocytogenes and L. innocua share the ecological niches, the isolation of both bacteria is not surprising and isolation of L. innocua in fishery products is considered as an indicator of possible contamination with L. monocytogenes [26]. L. grayi was found in 3 samples (3%) only in the warm season and there are no previous reports of isolation of this strain in Iranian seafood. Failure to isolate L. grayi in the cold season needs more

Fish	Number of samples	Number and (%) of samples positive for		
		L. monocytogenes	L. innocua	L. grayi
		Cold session		
Grass carp	25	2(8)ª	7(28)	-
Common carp	25	-	-	-
Warm session				
Grass carp	25	2(8) <sup>b</sup>	2(8)	1(4)
Common carp	25	-	2(8)	2(8)
Total	100	4(4)	11(11)	3(3)

a Both strains were serotype 1/2 a and were isolated from gut and gills

b One strain was serotype 1/2 a and one was serotype 4b and were isolated from gills

investigation. The location of Listeria spp. and *L. monocytogenes* in different parts of fish differed with statistical significance in samples. Up to 75% of the *L. monocytogenes* and 83.3% of Listeria spp. positive samples were gill samples. Also, It can be stated that only grass carp fishes were contaminated by *L. monocytogens*.

The serotypes of fish-associated L. monocytogenes have been reported by few authors. Adesiyun [14] found that all the strains isolated from seafood were serotype 4b whereas the tropical isolates of L. monocytogenes belonged to serotypes 1 and 4 [27]. Basti et al. [19] noted that L. monocytogenes isolated from fresh, smoked and salted Iranian fish belong to either serovar 1/2a or 1/2b. Our results showed that the isolates of L. monocytogenes from farmed tropical fish belonged to serotypes 1/2a and 4b. The serotypes of *L. monocytogenes* more commonly associated with sporadic cases and outbreaks of listeriosis are 4b, 1/2a and 1/2 b, accounting for more than 90% of all cases reported [28]. All our isolates of L. monocytogenes were 1/2a and 4b; this underscores of food items such as farmed tropical fish to cause listeriosis. The increased number of listeriosis cases over the last few years and the fact that the majority of strains were identified as virulent in our study, indicate that the present HACCP-based strategy should be readjusted to take this in to consideration.

Khuzestan province is one of the major regions in Iran for culture and export of freshwater fish. Most of the fish farms are situated near Karone, the biggest fresh water river in Iran and use the river as water intake. Isolation of *Listeria* spp. from the gut and gills of fishes indicates that the fishes are contaminated via the river or by the animal feces used for fertilizing the farm ponds. The river also could be contaminated with *L. monocytogenes* from farm sewage or other sources.

In conclusion, isolation of *L. monocytogenes* in tropical fish in this study indicates that there is a risk of acquiring listeriosis through this product in Iran. Although the bacterium will be killed by sufficient cooking, it may pose a health risk in the kitchen or restaurants by cross-contamination of cooked food or other kinds of ready-to-eat food. Using a risk analysis approach, research should be carried out to assess the prevalence of *L. monocytogenes* in other kind of fresh water and seawater fishes, also in frozen, smoked and lightly preserved fish product in Khozestan province.

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### REFERENCES

- Marth, E.H., 1988. Disease characteristics of Listeria monocytogenes. Food Technology, 42: 165-168.
- Alves, V.F., E.C. De Martinis, M.T. Destro, B.F. Vogel and L. Gram, 2005. Anti listeria activity of a *Carnobacterium piscicola* isolated from Brazilian smoked fish (surubim [Pseudoplatystoma sp.]) and its activity against a persistent strain of *Listeria monocytogenes* isolated from surubim. Journal of Food Protection, 68: 2068-2077.
- Bou-M'Handi, N. and A. El Marrakchi, 2003. Incidence of *Listeria* spp. in marine environment in Morocco. Sciences Des Aliments, 23: 513-523.
- Colburn, K., C.A. Kaysner, C. Abyta and M.M. Wekell, 1990. *Listeria* species in California Coast esturine environment. Appl. Environ. Microbiol., 56: 2007-2011.
- Hartemink, R. and F. Georgsson, 1991. Incidence of Listeria species in seafood and seafood salads. Intl. J. Food Microbiol., 12: 189-195.
- Rorvik, L.M., 2000. Listeria monocytogenes in the smoked salmon industry. Intl. J. Food Microbiol., 62: 183-190.
- Rorvik, L.M., B. Aase, T. Alvestad and D.A. Caugant, 2000. Mulecular epidemiological survey of Listeria monocytogenes in seafoods and seafood-processing plants. Appl. Environ. Microbiol., 66: 4779-4784.
- FAO, 1999. Fisheries Report No. 604. Expert consultation on the trade impact of *Listeria* in fish products. Amherst. MA. USA.
- Laciar, A.L. and O.N.P. De Centorbi, 2002. Listeria species in seafood; Isolation and characterization of Listeria spp. from seafood in San Luis, Argentina. Intl. J. Food Microbiol., 19: 645-651.
- 10. Ben Embarek, P.K., 1994. Presence, detection and growth of *Listeria monocytogenes* in seafoods: a review. Intl. J. Food Microbiol., 23: 17-34.

- 11. Fuchs, R.S. and P.K. Surenran, 1989. Incidence of *Listeria* in tropical fish and fishery products. Lett. Appl. Microbiol., 9: 49-51.
- Karunasagar, I., S.K. Karunasagar and W. Geobel, 1992. Incidence of *Listeria* species in tropical seafood. In 11th International symposium on the problems of listeriosis (Abstract 155). Copenhagen, Denmark.
- Manoj, Y.B., G.M. Rosalind, I. Karunasagar and S.K. Karunasagar, 1991. Listeria in fish and fish handling area, Mangalors, India. Asian Fishery Science, 4: 119-122.
- Adesiyun, A.A., 1993. Prevalence of Listeria spp. Campylobacter spp. Salmonella spp. Yersinia spp. and toxigenic Escherichia coli on meat and seafoods in Trinidad. Intl. J. Food Microbiol., 10: 395-403.
- Jeyasekaran, G., I. Karunasagar and I. Karunasagar, 1996. Incidence of *Listeria spp.* in tropical fish. Intl. J. Food Microbiol., 31: 333-340.
- Destro, M.T., 2000. Incidence and significance of Listeria in fish and fish products from Latin America. Intl. J. Food Microbiol., 62: 191-196.
- Parihar, V.S., S.B. Barbuddhe, M.L. Danielsson-Tham and W. Tham, 2008. Isolation and characterization of *Listeria* species from tropical seafoods. Food Control, 19: 566-569.
- 18. Sarkeshik Khabazi, S.M., 2001. Incidence of Listeria monocytogenes in cultivated and smoked fishes in Tehran and Gilan province, Iran (D.V.M. thesis no: 2783). Department of Food Hygiene, Faculty of Veterinary Medicine. Tehran University, Iran.
- Basti, A., A. Misaghi, T.Z. Salehi and A. Kamkar, 2006. Bacterial pathogens in fresh smoked and salted Iranian fish. Food Control, 17: 183-188.

- Jalali, M. and D. Abedi, 2008. Prevalence of Listeria species in food products in Isfahan, Iran. Intl. J. Food Microbiol., 122: 336-340.
- Nik Pay, A., 2002. Survey of contamination of smoked fishes to Listeria monocytogenes, Staphylococcus aureus in Gilan province, Iran (D.V.M. thesis no: 2260). Department of Food Hygiene, Faculty of Veterinary Medicine. Tehran University, Iran.
- Lovett, J., 1988. Isolation and enumeration of Listeria monocytogenes. Food Technol. Biotechnol., 42: 172-175.
- Seelinger, H.P.R. and D. Jones, 1986.
  Listeria. In P.H.L. Sneath, N.S. Maine, M.E. Sharpe and J.K. Holt, Bergey's Manual of Systematic Bacteriology (pp. 1235-1245). Baltimore, Maryland: Williams and Wilkins.
- Seelinger, H.P.R. and K. Höhne, 1979.
  Serotyping of *Listeria monocytogenes* and related species. In T. Bergan and J.R. Norris, Methods in microbiology (pp. 31-49). London: Academic Press.
- Dhanashree, B., S.K.Otta, I. Karunasagar, V. Goebel and I. Karunasagar, 2003. Incidence of *Listeria spp*. in clinical and food samples in Mangalore, India. Intl. J. Food Microbiol., 20: 447-453.
- 26. Jinneman, K.C., M.M. Wekell and M. Eklund, 1999. Incidence and behaviour of *L. monocytogenes* in fish and seafood products. In E.T. Ryser and E.H. Marth, Listeria, Listeriosis and Food Safety (pp. 631-655) New York: Marcel Dekker.
- Karunasagar, I. and I. Karunasagar, 2000. Listeria in tropical fish and fishery products Intl. J. Food Microbiol., 62: 177-181.
- 28. Farber, J.M. and P.I. Peterking, 1991. *L.monocytogenes*, a food-borne pathogen, Microbiol. Rev., 55: 476-511.