

## Stress Enhances the Synthesis of the Therapeutically Important Secondary Metabolites in Cyanobacteria

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**Abstract:** This review is an extension of my original study "Characterization of Nitrogen Stress Induced Alterations in Cyanobacterial Photosynthesis." It was clear from the literature that other secondary metabolites were produced during stress. Secondary metabolites are generally defined as compounds that are no longer needed for a primary metabolism of an organism. While most of the metabolites can be categorized as primary or secondary, there is a certain overlap. Some are essential for primary metabolism, however, are only synthesized by species and thus secondary metabolites. Under nitrogen stress conditions, the number of phycobiliproteins increased considerably, accompanied by an increase of their antioxidant activities. Moisturizers are synthesized from the secondary metabolites of extremophilic cyanobacteria. Cyanobacteria protective methods for counteracting harmful ultraviolet radiation from the sun are now widely discussed. Several clinical studies have found a high efficacy secondary metabolite in cancer treatment. Low external pH triggers heat shock proteins which have medicinal principles including antimicrobial and antioxidative action. In HIV patients, insulin sensitivity increases more when Spirulina is used as a nutritional supplement instead of soya. Cyanobacteria provide a rich wealth of chemicals for the discovery of lead compounds and new medicines that are safe yet promising. Pharmaceutical companies should conduct clinical trials on several bioactive molecules obtained from cyanobacteria exhibiting a broad spectrum of activities, such as antitumor, antibacterial and antiviral effects and protease inhibition. Recently, also a growing interest has been shown in the development of bioactive compounds for trade or medical applications.

**Key words:** Chromosomal Damage • Anti-Carcinogenic • Anti-Microbial • Conventional Moisturizers • Phenylketonuria • And Hyperlipidemia

### INTRODUCTION

Cyanobacteria are considered a rich source of secondary metabolites that could be used in the pharmacological field as biotechnology. Recently also a growing interest has been shown in the development of bioactive compounds for trade or medical applications. Intracellular and extracellular metabolites are known to produce by cyanobacteria which have potential biological activities such as antibacterial, anti-fungal, antiviral, anti-inflammatory, anti-pathological, herbicide and immunosuppressive effects [1-3]. The therapeutic benefit of Spirulina has been recorded in several studies. It also offers improved immune responses and decreases serum glucose and lipid levels to heavy and medicinal drugs, including its use for hyperlipidemias, obesity, HIV, diabetes, obesity and hypertension [4-7].

Spirulina platensis is the nature's richest source of vitamin B12 with high levels of amino acid (62%), antiviral, anti-cancer, anti-diabetic, antioxidant, anti-inflammatory and anti-metastatic function. Such features make *S. platensis* extract a potential biomedical pharmaceutical. Spirulina has been well established in industry. It has a wide range of protein sources, polyunsaturated fatty acids ( $\gamma$ -linoleic acid [GLA]), antioxidants (phycocyanin and carotenoids) and vitamins as health food [8].

Thousands of medical, pharmaceutical and nutritional articles on the importance of different cyanobacteria are available, particularly *S. platensis*.

This paper is an extension of my original study "Characterization of Nitrogen Stress Induced Alterations in Cyanobacterial Photosynthesis." The analysis of work in *S. platensis* was noteworthy for alterations in photosynthesis under nitrogen pressure (40ug) [9]. It was clear from the results

Table 1: List of secondary metabolites accumulated during stress conditions in Spirulina

SN	Stress condition	Chemical group	Name of secondary metabolites	Protecting mechanism	Reference
1.	UV stress	UV-absorbing compounds	Mycosporine-like amino acids and Scytonemin (Scy)	Photoprotection	[17]
2.	Carbon stress	Polyketides,	Indolactam ring composed L-valine, L-tryptophan and methionine.	Restoring the carbohydrate levels in organisms.	[18]
3.	Oxidative stress causing gene mutations.	Peptides	Aeruginosamide Patellamide	Self-protecting of genetic materials.	[19]
4.	Heavy metals & free radicles & Reactive oxygen spices	Isoprenoids & Terpenoids	Diphosphate and Dimethylallyl triphosphate (DMADP) precursors & phycocyanin.	For protecting the self-genetic materials & lipids	[20]
5.	Ammonia stress	Nitrogen-containing compounds	Phenylalanine ammonia-lyase	To protect self-cytoskeleton assembly, immune stress, oxidative stress and signal transduction of apoptosis	[21]
6.	Iron stress	Phytohormones, iron-chelators Hepatotoxin, neurotoxins	Cytokinin and gibberellin-like compounds, schizokinen, anachelin and synechobactins, etc. Saxitoxin, anatoxines, microcystins and nodularins.	To improve their growth. and heavy metal poisonings Which are poisonous compounds and exerts a toxic effect on another animal.	[22]
7.	Temperature stress	Carotenoids, Unsaturated fatty acids	Antifreeze proteins, astaxanthin	To protect membranous Rigidity.	[23]
8.	Salt & PH stress	Secondary carotenoids, Heat shock Proteins.		To preserve homeostatic changes.	[24]

that other secondary metabolites were produced during stress. What are they? What is the meaning of secondary stress metabolites? Could we treat different cancers using their special medicinal properties?

Secondary metabolites are generally defined as compounds that are no longer needed for a primary metabolism of an organism. Those are also listed as natural products. These secondary metabolites are usually specific to some organisms and do not occur in each environment.

While most of the metabolites can be categorized as primary or secondary, there is a certain overlap. Some are essential for primary metabolism; however, secondary metabolites are only synthesized by specific species.

Depending on the stress environment, different suites of secondary metabolites can be generated with a high level of structural variations across the different compound groups. A range of secondary metabolites for the development of healthier food products are available from some algal species, namely *Chlorella*, *Hematococcal*, *Dunaliella* and *Spirulina* [10, 11]. However, algae produce secondary alkaloids, flavonoids, glycosides, terpenoids or phenazines [12]. Algae are a product of algae. These thus have a wide range of roles, such as cell defense, chemosensory, photoprotection and antioxidant, defense against predators and grazers. Such products can be used as nutraceuticals, cosmeceuticals and medicines in industrial biotechnology.

Current review focused on different secondary metabolites produced by cyanobacteria when exposed to stress environment.

Secondary metabolites are produced by cyanobacteria often by providing protection and helping to survive as a reaction to the biotic or abiotic pressure of the surrounding environment [13, 14]. Cyanobacteria are a rich source of secondary metabolites because they survive in a variety of environments. Non-ribosomal peptide (NRPs) are widely used as secondary metabolites for cyanobacteria, which are generated using non-ribosomal peptide synthetases. Polyketides which are biosynthesized with polyketide synthase (PKS) from acetyl-CoA are another big group of secondary metabolites. Alkaloids are natural compounds containing nitrogen, commonly poisonous, such as the paralytic poison of shellfish, saxitoxins contained in several cyanobacteria [15, 16]. This class also contains ribosomal peptides and isoprenoids (carotenoids). Table 1 summarizes the list of accumulated secondary metabolites under various stress conditions.

UV radiation exposure can contribute to cyanobacteria's oxidative stress [25]. Cyanobacteria produce secondary UV-absorptive screening biomolecules, such as Mycosporin-like amino acids (MAAs) and scytonemine (Scy) during oxidative stress [17]. which may be capable of free radicles detoxifying. Cyanobacteria protective methods for counteracting

harmful UV radiation from the sun are now widely discussed. Several clinical studies have found a high efficacy secondary metabolite in cancer treatment [26].

Under carbon famine or stress conditions, by synthesizing the secondary metabolites, such as indolactam ring composed of L-valine, L-tryptophan and methionine, the cyanobacteria can be adapted to a new environment. Recent clinical studies have shown that many species of staphylococcus, streptococcus and clostridium microorganisms are effectively killed by these secondary metabolites [27].

Aerobic metabolism byproducts and potent agents causing oxidative damage are reactive oxygen species (ROS). Cyanobacteria are often subjected to external changes in their natural habitat, such as dramatic variations in light intensity, the ability to perceive ROS and rapidly cause antioxidant survival [28].

The generation of ROS, including superoxide, hydrogen peroxide and radical hydroxyl radicals, also causes oxidative stress by causing nuclear acid, protein and lipid damage. Cyanobacteria generate enzyme catalyzed and non-enzyme catalyzed antioxidants to neutralize the toxic effect of ROS [28].

The reduction in the concentration of nitrogen from 200 ug to 40 ug in our experiment led to alterations in the photosynthesis; in electron transport chain [9] and a changed energy transfer between photosynthesis pigments [29]. The rise in nitrogen pressure cannot affect carotenoids-chlorophyll absorption ratio, but the ratio has dropped, suggesting that pigment protein phycobilisome are major targets [29]. In additional experiments to cultivate cyanobacterial (*Spirulina platensis* and *Nostoc muscorum*) under nitrogen stress conditions, the number of phycobiliproteins increased considerably, accompanied by an increase of their antioxidant activities.

The organism has the synthesis of Phenylalanine Ammonia Lyase (PAL) secondary metabolite, which can be removed by using phenolic composites and can be used for the treatment of phenylketonuria [30].

Different secondary metabolites including flavodoxin, saxitoxins, anatoxins, Microcystis, nodularin & phytohormones, iron-chelators to regulate the variety of metabolisms and growth rates, oxidative stress and others trigger deficiency of iron. The researchers have found that these secondary metabolites can have anti-microbial action and heavy metal poisoning antidote by generating their toxic effect on microorganisms and neutralizing heavy metals. According to previously published research studies [31].

Conventional moisturizing products, such as urea, glycerin and propylene glycol, have high absorptiveness of moisture, but their capacity for preservation of humidity is insufficient [32]. Extremophilic cyanobacteria species have defensive mechanisms that allow them to survive inhospitable environments like the hyper-arid desert and salt. One such mechanism is the production, in form of protective sheaths, slims, or capsules, of copious external polysaccharide layers. Through synthesizing both internal and external polysaccharides, cyanobacteria such as *Chroococcidiopsis* and *Nostoc* species may live with minimal water. Such metabolites can produce hydrating agents [33]. Additional studies have shown in *Nostoc* community, extra polymeric substances (EPS) have high moisture absorption and retention compared to chitosan (hygroscopic agent) and urea (humectant). They noticed that the moisture absorption level of EPS (10.1%) was much higher than the moisture content chitosan (6.3%) and urea (5.8%), despite exposure to 43% of relative moisture for 24hours [34].

Various cyanobacteria may tolerate extremely low temperatures. Ambient temperature transitions can cause a change in diaphragm stiffness and metabolic reaction rates or lower at suboptimal temperatures. Temperature-induced stress reactions can be used to produce useful metabolites, including unsaturated fatty acids, anti-freeze proteins, astaxanthanes or antioxidants, such as antioxidants, anticancer, anti-microbial impact, etc [35].

At different pH, cyanobacteria live for the maintenance of optimal metabolic rates and development. Under low pH than neutral pH, photosynthesis rates can be significantly higher to compensate for higher breathing rates. Increased membranes fatty acid concentration and acid-tolerant wall cell proteins are pathways often associated with low pH adaptation. Low external pH is shown to trigger heat shock protein, as is the case with some other types of stress. Studies have shown that their medicinal principles including antimicrobial and antioxidative action have been demonstrated in the previously published study [36].

The moisture retention capacities of cyanobacteria shall be related to EPS. The genes associated with EPS production are affected by several environmental factors. In many species of cyanobacteria, EPS production rates increase due to excess photo assimilated carbon nutrient deficiency (nitrogen and phosphor) and excretion is a flood mechanism to prevent harm to the photosynthesis devices [37].

Table 2: List of various biomedically interesting secondary metabolites.

S. no	Name of Secondary metabolite	Clinical use	Reference
1	Mycosporine-like amino acids	Sunscreen agent	[39]
2	Spirulan	Antiviral	[40]
3	$\gamma$ -linolenic acid	Precursor to prostaglandins	[41]
4	Phycocyanin	Cosmetic colorants	[42]
5	Apratoxins	Anticancer	[43]
6	Nostodione	Antifungal	[44]
7	Carotenoids	Antioxidant	[45]
8	Anatoxin-a	Anti-Inflammatory	[46]
9	Fischerellin	Antifungal	[46]
10	Scytonemin	Anti-inflammatory, Anti-proliferation	[47]

Table 3: Various Bioactive Metabolites of Cyanobacteria used in Cancer Treatment

Cancer	Active Metabolite	Cyanobacteria	Ref
Lung Cancer	Grassystatin A-B	Lyngbya confervoides	49
	Hectochlorin	L. majuscula	50
	Hermitamides A-B	L. majuscula	51
	Lyngbyastatin 1	L. majuscula	52
	Lyngbyastatin 4	L. confervoides	53
	Lyngbyastatin 5-7	Lyngbya spp.	54
	Lyngbyastatin 8-10	Lyngbya semiplena	55
	Malyngamide 2	L. sordida	56
	Malyngamide C, J and K	L. majuscula	57
	Malyngolide dimmer	L. majuscula	57
	Palmyramide A	L. majuscula	58
	Veraguamides A-G	S. cf. hydnoides	59
	Wewakazole	L. sordida	60
	Wewakpeptins	L. semiplena	59
Colon Cancer	Lagunamide C	L. majuscula	61
	Largazole	Symploca sp.	62
	Belamide A	Symploca sp.	63
	Apratoxins F and G	L. bouilloni	62
	Apratoxins B-C	Lyngbya sp.	62
	Borophycin	N. linckia and N. spongiaeforme	62
	Apratoxin A	Lyngbya majuscula	62
	Ankaraholide A	Geitlerinema	64
	Nostocyclopeptides A1 and A2	Nostoc sp.	65
	Obyanamide	L. confervoides	65
Tasiamide	Symploca sp.	66	
Cervical Cancer	Apratoxin A	Lyngbya majuscula	62
	Caylobolide B	Phormidium spp.	67
	Cryptophycin-1	N. linckia	68
Breast Cancer	Ankaraholide A	Geitlerinema	69
	TZT-1027 analog of dolastatin 10	Symploca sp.	70
	Dolastatin 15	Lyngbya sp.	66
	Cematodin	Lyngbya sp.	66
	synthadotin	Lyngbya sp.	66
	Isomalyngamide A and A-1	L. majuscula	71
	Pitipeptolide C	L. majuscula	72
	Symplostatin 1	S. hydnoides	73
Epidermoid Cancer	Symplostatin 3	Symploca sp.	70
	Tasiamide	Symploca sp.	66
	Tasiamide B	Symploca sp.	74
	Tasipeptins A&B	Symploca sp.	66
	Tiglicamides A&C	L. confervoides	75
	Ulongapeptin	Lyngbya sp.	74
Leukemia	Calothrixin B	Calothrix	76
	Coibamide A	Leptolyngbya sp.	77
	Cryptophycin-1	N. linckia	68

**Secondary Metabolites in Cancer Treatment:** As per GLOBECAN 2018 estimates, cancer is the second leading cause of death worldwide and estimated to be responsible for 9.6 million deaths, about 1 in 6 cancer deaths worldwide [38]. Many cancer therapies are available but are limited due to some serious side effects and the development of drug-resistant cancer cells. Their goal is to encourage the use in conjunction with established Golden Standard Medicines of the use of natural anticancer products. Cyanobacteria make a variety of biomedically important, therapeutically useful bioactive compounds.

One of the most important cancer and other disease treatments available nowadays is chemotherapy with limited effectiveness due to some serious side effects and the development of medical-resistant cancer cells. Cyanobacteria provide a rich wealth of chemicals for the discovery of lead compounds and new medicines that are safe yet promising. Natural products are up to 80 % of the most recent antibacterial and cancer medicines approved between 1983 and 1994 [48]. Table 3 summarizes various anti-carcinogenic biologically active metabolites produced by cyanobacteria.

*S. subsalsa* extract was tested by the MTT viability test to T47D human cell line. The highest power in decreasing the viability of T47D cells has been found at 1, fractions 40, 60, 80 and 90. [78]. Considering that *Spirulina* strain from the Baltic compounds is produced, *S. subsalsa* not only have high activity with T47D cells, but they seem also to operate selectively because they have no strong inhibitory effect against the enzymes tested. This shows the potential of cyanobacteria as an origin of significant cytotoxic agents [78].

**HIV Infection:** Patients with HIV may experience sugar irregularities due to infections and antiretroviral medications. A randomized trial to determine the effect of *Spirulina platensis* versus soybean was conducted in 33 insulin-resistant HIV-infected patients as food supplements on HIV / HAART-associated resistance to insulin (IR). This pilot study reveals that in HIV patients, insulin sensitivity increases more when spirulina is used as a nutritional supplement instead of soya [79].

**Chromosomal Damage:** *Spirulina* has shown that the chromosome aberrations caused by CCl<sub>4</sub> in bone marrow cells are ameliorating, which confirms its defense from chromosome damage [80].

## CONCLUSIONS

Cyanobacteria are a rich source of secondary metabolites due to their ability to thrive under a variety of environments. Depending on the stress environment different sequences of secondary metabolites can be produced with a high structural variation across the various compound groups. The properties of cyanobacterial extracts are antiviral, anticancer, hypocholesterolemic, anti-diabetic, antioxidant, non-inflammatory and anti-metastatic, potential for biomedical applications. The review demonstrates that the cyanobacteria have the potential for expanded utilization in drug discovery. Hence, pharmaceutical companies should conduct clinical trials on several bioactive molecules obtained from cyanobacteria exhibiting a broad spectrum of activities.

## REFERENCES

1. Alejandro, M.S.M., A.D. Rodriguez R.G.S. Berlinck and M.T. Hamann, 2009. Marine pharmacology in 2005-6: Marine compounds with anthelmintic, antibacterial, anticoagulant, antifungal, anti-inflammatory, antimalarial, antiprotozoal, antituberculosis and antiviral activities; affecting the cardiovascular, immune and nervous systems and other miscellaneous mechanisms of action. *Biochimica et Biophysica Acta*, 1790: 283-308.
2. Orio, C., 1983. *Spirulina*, the edible microorganism. *Microbiological Reviews*, 47(4): 551-578.
3. Nuhu, A.A., 2013. *Spirulina* (Arthrospira): An Important Source of Nutritional and Medicinal Compounds. *Journal of Marine Biology*, pp: 1-8.
4. Kaori, K., Z. Piao, S. Suzuki, T. Fujimori, Tajiri, K. Nagai, T. Iyoda, A. Yamada, T. Hayakawa, M. Ishiwara, S. Horaguchi, A. Belay, T. Tanaka K. Takano and M. Hangy, 2015. *Spirulina*-Templated Metal Microcoils with Controlled Helical Structures for THz Electromagnetic Responses. *Scientific Reports*, 4: 4919.
5. Wang, Y.F., Y. Tie P.I. Boross J. Tozser A.K. Ghosh R.W. Harrison and I.T. Weber, 2007. potent new antiviral compound shows similar inhibition and structural interactions with drug-resistant mutants and wild type HIV-1 Protease. *Journal of Medicinal Chemistry*, 50(18): 4509-15.

6. Karkos, P.D., S.C. Leong C.D. Karkos N. Sivaji and D.A. Assimakopoulos, 2011. Spirulina in clinical practice; evidence-based human applications. Evidence Based Complementary and Alternate Medicine, (2).
7. Esquenazi, E., C. Coates, L. Simmons, D. Gonzalez, W.H Gerwick, P.C. Dorrestein and C. Coates, 2008. Visualizing the spatial distribution of secondary metabolites produced by marine cyanobacteria and sponges via MALDI-TOF imaging. Molecular of Biosystems, 4: 562-570.
8. Spolaore, P., C. Joannis-Cassan, E. Duran and A. Isambert, 2006. Commercial applications of microalgae. Journal of Bioscience and Bioengineering, 101: 87-96.
9. Peter, P., A. Phaninatha Sarma K. Shamim Basha and S.D.S. Murthy, 2010. Nitrogen Chlorosis Induced Alterations in the Photosynthetic Electron Transport Activities of the Cyanobacterium, Spirulina platensis. African Journal of Basic & Applied Sciences, 2(3-4): 60-63.
10. Zepka, L.Q., E. Jacob-Lopes R. Goldbeck L.A. Souza-Soares and M.I. Queiroz, 2010. Nutritional evaluation of single-cell protein produced by Aphanothece microscopic Nägeli. Bioresource Technology, 101: 7107-7111.
11. Kuhad, R.C., A. Singh K.K. Tripathi and R.K. Saxena., 1997. Microorganisms an alternative source of protein. Nutrition Reviews, 55: 65-67.
12. Pelizer, L.H., E.D.G. Danesi, C.O. Rangel, C.E.N. Sassano, J.C.M. Carvalho S. Sato and I.O. Moraes, 2003. Influence of inoculum and concentration in Spirulina platensis cultivation. Journal of Food Engineering, 56: 371-375.
13. Romay, Ch., R. González, N. Ledón, D. Ramirez and V. Rimbau, 2003. C-Phycocyanin A Biliprotein With Antioxidant, Anti-Inflammatory and Neuroprotective Effects. Current Protein Peptide Science, 4(3): 207-16.
14. Ambrosi, M.A., C.O. Reinehr, T.E. Bertolin, J.A.V. Costa and L.M. Colla, 2008. Propriedades de saúde de Spirulina spp. Revista de Ciências Farmacêuticas Básica e Aplicada, 29(2): 109-117.
15. Rastogi, R.P. and R.P. Sinha, 2009. Biotechnological and industrial significance of cyanobacterial secondary metabolites. Biotechnology Advances, 27: 521-539.
16. Bethan, K. and C. Llewellyn, 2018. Secondary Metabolites in Cyanobacteria. Secondary Metabolites - Sources and Applications.
17. Rastogi, P.R. and D. Madamwar, 2015. UV-Induced Oxidative Stress in Cyanobacteria: How Life is able to Survive? Biochemistry & Analytical Biochemistry, 4: 173.
18. Chen, C.Y., K.L. Yeh, R. Aisyah, D.J. Lee and J.S. Chang, 2010. Cultivation photobioreactor design and harvesting of microalgae for biodiesel production: a critical review. Bioresource Technology, 102(1): 71-81.
19. Eiji, S., H. Ohkawa, K. Moriya, T. Matsubara, Y. Nagaike, I. Iwasaki, S. Fujiwara, M. Tsuzuki and Y. Nakamura, 2010. Carbohydrate Metabolism in Mutants of the Cyanobacterium Synechococcus elongates PCC 7942 Defective in Glycogen Synthesis. Applied and Environmental Biology, 76(10): 3153-3159.
20. Corinne, C. and F. Chauvat, 2015 Responses to oxidative and heavy metal stresses in cyanobacteria: recent advances. International Journal of Molecular Sciences, 16(1): 871-886.
21. Antonia, H., A.M. Muro-Pastor and E. Flores, 2001 Nitrogen control in cyanobacteria. Journal of Bacteriology, 183(2): 411-425.
22. Mathew, B.S., D.A. Contreras, R.G. Treble and H.G. Wegeera, 2012. Two distinct pathways for iron acquisition by iron-limited cyanobacterial cells: Evidence from experiments using siderophores and synthetic chelators. Botany, 90(3): 181-190.
23. Amitav, B., 2019. High-Temperature Stress and Metabolism of Secondary Metabolites in Plants. Effect of High Temperature on Crop Productivity and Metabolism of Macro Molecules. Academic Press, pp: 391-484.
24. Tünay, K. and R. Erenler, 2018. Effect of Salt and pH Stress of Bioactive Metabolite Production in Geitlerinema carotinosum. Turkish Journal of Agriculture - Food Science and Technology, 6(9): 1274-1278.
25. Kamer, I., R. Sarig, Y. Zaltsman, H. Niv, G. Oberkovitz, L. Regev, G. Haimovich, Y. Lerenthal, R.C. Marcellus and A. Gross, 2005. Proapoptotic BID is an ATM effector in the DNA-damage response, Cell, 122: 593-603.
26. Rastogi, R.P., R.P. Sinha, S.H. Moh, T.K. Lee, S. Kottuparambil, Y.J. Kim, J.S. Rhee, E.M. Choi, M.T. Brown, D.P. Häder and T. Han, 2014. Ultraviolet radiation and cyanobacteria. J Photochemistry Photobiology. B, Biology, 141: 154-169.

27. Michalek-Wagner, K., 2001. Seasonal and sex-specific variations in levels of photo-protecting mycosporine-like amino acids. *Marine Biology*, 139: 651-660.
28. Latifi, A., M. Ruiz and C.C. Zhang, 2009. Oxidative stress in cyanobacteria. *FEMS Microbiol Reviews*, 33(2): 258-78.
29. Peter P., A. Phaninatha Sarma, M.D. Azeem ul Hasan and S.D.S. Murthy, 2010. Studies on the Impact of Nitrogen Starvation on the Photosynthetic Pigments Through Spectral Properties of the Cyanobacterium, *Spirulina platensis*: Identification of Target Phycobiliprotein under Nitrogen Chlorosis. *Botany Research International*, 3(1): 30-34.
30. Jian-Qiang, K., 2015. Phenylalanine ammonia-lyase, a key component used for phenylpropanoid production by metabolic engineering. *RCS Advances*, 5: 62587-62603.
31. Gaurav, S., M. Kumar, M.A. Ali and N.D. Jasuja, 2014. Effect of Carbon Content, Salinity and pH on *Spirulina platensis* for Phycocyanin, Allophycocyanin and Phycoerythrin Accumulation. *Journal of Microbial & Biochemical Technology*, 6: 4.
32. Zhao, L., F. Fan, P. Wang and X. Jiang, 2013. Culture medium optimization of a new bacterial extracellular polysaccharide with excellent moisture retention activity. *Applied Microbiology and Biotechnology*, 97: 2841-2850.
33. Warren Rhodes, K.A., C.P. McKay, L.N. Boyle, M.R. Wing, E.M. Kiekebusch, D.A. Cowan, F. Stomeo, S.B. Pointing, K.F. Kaseke, F. Eckardt, J.R. Henschel, A. Anisfeld, M. Seely and K.L. Rhodes, 2013. Physical ecology of hypolithic communities in the central Namib Desert: the role of fog, rain, rock habitat and light. *Journal of Geophysical Research: Biogeosciences*, 118: 1451-1460.
34. Li, H., J. Xu Y. Liu, S. Ai, F. Qin, Z. Li, H. Zhang and Z. Huang, 2011. Antioxidant and moisture-retention activities of the polysaccharide from *Nostoc commune*. *Carbohydrate Polymers*, 83: 1821-1827.
35. Abed, R.M., S. Doritos and K. Sudesh, 2009. Applications of cyanobacteria in biotechnology. *Journal of Applied Microbiology*, 106: 1-12.
36. Goldman, J.A.L., M.L. Bender and M.M.M. Francois, 2017. The effect of pH and pCO<sub>2</sub> on photosynthesis and respiration in the diatom *thalassiosira weissflogii*. *Photosynthesis Research*, 132(1): 83-93.
37. Chakraborty, T. and R. Pal, 2014. An overview of cyanobacterial exopolysaccharides: features, composition and effects of stress exposure. *International Journal of Life Sciences*, 8(4): 1-9.
38. Bray, F., F. Ferlay, I. Soerjomataram, R.L. Siegel, L.A. Torre and A. Jemal, 2018. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. a Cancer Journal For Clinicians*, 68(6): 394-424.
39. Rastogi, R.P., R.P. Richa, Sinha, S.P. Singh and D.P. Häder, 2010. Photoprotective compounds from marine organisms. *Journal of Industrial Microbial Biotechnology*, 37: 537-558.
40. Burja, A.M., S. Dhamwichukorn and P.C. Wright, 2003. Cyanobacterial postgenomic research and systems biology. *Trends in Biotechnology*, 21(11): 504-511.
41. Chaiklahan, R., N. Chirasuwan V. Loha and B. Bunnag, 2008. Lipid and Fatty acids extraction from the cyanobacterium *Spirulina*. *Science Asia*, 34: 299.
42. Nyok-sean, L., M. Matsui and A.A. Abdullah, 2015. Cyanobacteria: Photoautotrophic microbial factories for the sustainable synthesis of industrial products. *Journal of Biomedicine and Biotechnology*, 2015(2).
43. Vijayakumar, S. and M. Menakha, 2015. Pharmaceutical applications of cyanobacteria-A review. *Journal of Acute Medicine*, 5: 15-23.
44. Burja, A.M., B. Banaigs E. Abou-Mansour J. Grant Burgess and P.C. Wright, 2001. Marine cyanobacteria A prolific source of natural products. *Tetrahedron*, 57: 9347-9377.
45. Spolaore, P., C. Joannis-Cassan E. Duran and A. Isambert, 2006. Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*, 101: 87-96.
46. Abed, R.M.M., S. Dobretsov and K. Sudesh, 2009. Applications of cyanobacteria in biotechnology. *Journal of Applied Microbiology*, 106: 1-12.
47. Rastogi, R.P., R.R. Sonani and D. Madamwar, 2015. Cyanobacterial sunscreen scytonemin: Role in photoprotection and biomedical research. *Applied Biochemistry and Biotechnology*, 176(6): 1551-63.
48. Vijayakumar, S. and M. Menakha, 2015. Pharmaceutical applications of cyanobacteria. A review. *Journal of Acute Medicine*, 5(1): 15-23.
49. Clare, J.J., S.N. Tate M. Nobbs and M.A. Romanos, 2005. Voltage-gated sodium channels as therapeutic targets. *Drug Discovery Today*, 5(11): 506-520.

50. Davies-Coleman, M.T., T.M. Dzeha, A.G. Christopher So. Hess, L.K. Pannell, D.T. Hendricks and C.E. Arendse, 2003. Isolation of homodolastatin 16, a new cyclic depsipeptide from a Kenyan collection of *Lyngbya majuscula*. *Journal of Natural Products*, 66(5): 712-715.
51. Graf, K.M., M.G. Tabor M.L. Brown and M. Paige, 2009. Synthesis of (S)-jamaicamide C carboxylic acid. *Organic Letters*, 11(23): 5382-5385.
52. Dale, J.A., D.L. Dull and H.S. Mosher, 1969. a-Methoxy-a-trifluoromethylphenylacetic acid, a versatile reagent for the determination of enantiomeric composition of alcohols and amines. *Journal of Organic Chemistry*, 34(9): 2543-2549.
53. Ainslie, R.D., J.J. Barchi, Jr M. Kuniyoshi, R.E. Moore and J.S. Mynderse, 1985. Structure of malyngamide C. *Journal of Organic Chemistry*, 50: 2859-2862.
54. Taiki, U., M. Sueda, T. Kamura, T. Kawahara, X. Han, T. Okino and F. Matsuda, 2012. Synthesis and biological activity of kalkitoxin and its analogues. *Journal of Organic Chemistry*, 77: 357-370.
55. Golakoti, T., W.Y. Yoshida, S. Chaganty and R.E. Moore, 2001. Isolation and structure determination of nostocyclopeptides A1 and A2 from the terrestrial cyanobacterium *Nostoc* sp. ATCC53789. *Journal of Natural Products*, 64: 54-59.
56. Shi, S.R., R.J. Cote and C.R. Taylor, 1999. Standardization and further development of antigen retrieval immunohistochemistry: strategies and future goals. *Journal Histotechnology*, 22: 177-192.
57. Bhatnagar, I. and S.K. Kim, 2010. Immense essence of excellence: marine microbial bioactive compounds. *Marine Drugs*, 8: 2673-2701.
58. Cruz-Rivera, E. and V.J. Paul, 2007. Chemical deterrence of a cyanobacterial metabolite against generalized and specialized grazers. *Journal of Chemical Ecology*, 33: 213-217.
59. Nogle, L.M., B.L. Marquez and W.H. Gerwick, 2003. Wewakazole, a novel cyclic dodecapeptide from a Papua New Guinea *Lyngbya majuscula*. *Organic Letters*, 5: 3-6.
60. Han, B., D. Goeger, C.S. Maier and W.H. Gerwick, 2005. The wewakpeptides, cyclic depsipeptides from a Papua New Guinea collection of the marine cyanobacterium *Lyngbya semiplena*. *Journal of Organic Chemistry*, 70: 3133-3139.
61. Carter, D.C., R.E. Moore, J.S. Mynderse, W.P. Niemczura and J.S. Todd, 1984. Structure of majusculamide C, a cyclic depsipeptide from *Lyngbya majuscula*. *Journal of Organic Chemistry*, 49: 236-241.
62. Leusch, H., R.E. Moore, V.J. Paul, S.L. Mooberry and T.H. Corbett, 2001. Isolation of dolastatin 10 from the marine cyanobacterium *Symploca* sp. VP642 and total stereochemistry and biological evaluation of its analogue symprostatin 1. *Journal of Natural Products*, 64: 907-910.
63. MacMillan, J.B., T.F. Molinski and A. Caylobolide, 2002. A unique 36-membered macrolactone from a Bahamian *Lyngbya majuscula*. *Organic Letters*, 4: 1535-1538.
64. Hong, J. and H. Luesch, 2012. Largazole: From discovery to broad-spectrum therapy. *Natural Product Reports*, 29: 449-456.
65. Burja, A.M., B. Banaigs, E. Abou-Mansour, J.G. Burgess and P.C. Wright, 2001. Marine cyanobacteria: a prolific source of natural products. *Tetrahedron*, 57: 9347-9377.
66. Kwan, J.C., K. Taori V.J. Paul and H. Luesch, 2009. Lyngbyastatins 8-10, elastase inhibitors with cyclic depsipeptide scaffolds isolated from the marine cyanobacterium *Lyngbya semiplena*. *Marine Drugs*, 7: 528-538.
67. Andrianasolo, E.H., H. Gross, D. Goeger, M. Musafija-Girt, K. McPhail, R.M. Leal S.L. Mooberry and W.H. Gerwick, 2005. Isolation of swinholide A and related glycosylated derivatives from two field collections of marine cyanobacteria. *Organic Letters*, 7(7): 1375-1378.
68. Mozzachiodi, R., R. Scuri, M. Roberto and M. Brunelli, 2001. Caulerpenyne, a toxin from the seaweed *Caulerpa taxifolia*, depresses after hyperpolarization in invertebrate neurons. *Neuroscience*, 107: 519-526.
69. Hong, J. and H. Luesch, 2012. Largazole: From discovery to broad-spectrum therapy. *Natural Product Reports*, 29: 449-456.
70. McPhail, K.L., J. Correa, R.G. Linington, J. Gonzalez E. Ortega-Barria, T.L. Capson and W.H. Gerwick, 2007. Antimalarial linear lipopeptides from a Panamanian strain of the marine cyanobacterium *Lyngbya majuscula*. *Journal of Natural Products*, 70: 984-988.
71. Doi, T., Y. Numajiri A. Munakata and T. Takahashi, 2006. Total synthesis of apratoxin A. *Organic Letters*, 8: 531-534.
72. Linington, R.G., D.J. Edwards, C.F. Shuman, K.L. McPhail, T. Matainaho and W.H. Gerwick, 2008. Symplocamide A, a potent cytotoxin and chymotrypsin inhibitor from the marine cyanobacterium *Symploca* sp. *Journal of Natural Products*, 71(1): 22-27.



73. Masatoshi, T.K., J.K. Nunnery, N. Engene, E. Esquenazi, T. Byrum, P.C. Dorrestein and W.H. Gerwick, 2010. Palmyramide A, a cyclic depsipeptide from a Palmyra Atoll collection of the marine cyanobacterium *Lyngbya majuscula*. *Journal of Natural Products*, 73: 393-398.
74. Salvador, L.A., J.S. Biggs, V.J. Paul and H. Luesch, 2011. Veraguamides A-G, cyclic hexadepsipeptides from a dolastatin 16-producing cyanobacterium *Symploca cf. hydroides* from Guam. *Journal of Natural Products*, 74(5): 917-927.
75. Horgen, F.D., E.B. Kazmierski, H.E. Westenburg, W.Y. Yoshida and P.J. Scheuer, 2002. Malevamide D: isolation and structure determination of an isodolastatin H analogue from the marine cyanobacterium *Symploca hydroides*. *Journal of Natural Products*, 65: 487-491.
76. Foster, B.J., M. Fortuna, J. Media, R.A. Wiegand and F.A. Valeriote, 1998. Cryptophycin 1 cellular levels and effects in vitro using L1210 cells. *Investigational New Drugs*, 16: 199-204.
77. Palermo, J.A., P.B. Flower and A.M. Seldes, 1992. Chondriamides A and B, new indolic metabolites from the red alga *Chondria* sp. *Tetrahedron Letters*, 33: 3097-3100.
78. Karolina, S., M. Wiglusz and H. Mazur-Marzecab, 2018. Bioactive metabolites produced by *Spirulina subsalsa* from the Baltic Sea. *Oceanologia*, 60(3): 245-255.
79. Azabji-Kenfack, M., L.G. Ekali, S. Eugene, O.E. Arnold, E.D. Sandrine, D.V. Weid, E. Gbaguidi, J. Ngogang and J.C. Mbanya, 2011. The Effect of *Spirulina platensis* versus Soybean on Insulin Resistance in HIV-Infected Patients: A Randomized Pilot Study. *Nutrients*, 3(7): 712-724.
80. Ashgan, A.A.G., A.E.M. Khaled, H.S. El-Sayed, H.M. Aboul-Ela, O.K. Shalaby, A.A. Khaled and L.A. Mohamed, 2018. Optimization of *Spirulina platensis* Biomass and Evaluation of its Protective Effect against Chromosomal Aberrations of Bone Marrow Cells. *Fisheries and Aquaculture Journal*, 10: 260.