

## Efficiency of Serological Tests for Detection of Brucellosis in Ruminant at South Provinces of Egypt

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**Abstract:** A total of 2138 serum samples (715 from cattle, 1323 from sheep and 100 from goats) from different districts in Assuit governorate, was tested for the detection of antibodies against *Brucella* spp. Results obtained by Buffer acidified plate antigen test (BAPAT) and Rose bengal test (RBT) as screening tests indicated a positive reactors percentage of 4.6-5.3, 4.4-7.6 and 10-15% followed by overall brucellosis incidence of 4.5, 5.2 and 5.0 % in case of cattle, sheep and goats, respectively. The *Brucella* positive reactors were subjected to confirmation by Serum agglutination test (SAT) and Rivanol test (Riv. T) in addition to the previous tests. It confirmed that the BAPAT indicated seroreactors of 91.3 % in sheep and 100% in cattle and goats. *Brucella melitensis* biotype 3 was isolated from 39 seropositive animals (9 Cattle, 25 sheep and 5 goats). In this investigation, the highest rate of sensitivity (97.4%) was detected by BAPAT, while the highest rate and specificity (87.6%) was by RIV.t.

**Key words:** Brucellosis • Serology • Buffer acidified plate antigen test (BAPAT) • Rivanol test (Riv. T) • Upper Egypt

### INTRODUCTION

Brucellosis, a disease caused by various species of the genus *Brucella*, has the most wide-spread zoonosis in the world [1]. Cross-transmission of brucellosis can occur between cattle, sheep, goats, camels and other species. Brucellosis is still endemic in countries of the Mediterranean basin, the Middle East and Central Asia. Human infection due to *Brucella* from camels is known to occur mostly through the consumption of unheated milk [2-4]. Brucellosis in sheep and goats, caused by *B. melitensis*, one of the most virulent species of *Brucella*, is responsible for important economic losses in sheep and goats farming. Ruminant brucellosis can cause abortion, weak offspring, infertility, loss of milk production and has been responsible for major economic losses [5]. The interest in brucellosis has increased since *Brucella* species has been identified as a potential biological weapon [6].

For several decades it has been recognized as a significant public health problem in the Middle East and recent reports suggested that its incidence is increasing in both ruminants and humans [7, 8] and that currently applied control measures may not be capable of reducing the levels of infection in ruminants [9].

In Egypt, *Brucella melitensis* biovar 3 is considered to be the predominant species of *Brucella* isolated from humans and animals [8]. Outbreaks in cattle due to *B. melitensis* have become a worldwide emerging problem particularly difficult to control due to the lack of knowledge on the epidemiology in this host species and of an effective vaccine [10].

Diagnosis of *Brucella* spp. infection is mainly based on the detection of antibodies in serum by serological tests. The Rose Bengal test (RBT) [11] and complement fixation test (CFT) [12] are the most accepted tests worldwide for this purpose [13] and the only approved for certification of sheep and goats flocks due to brucellosis

status in EU member states [14]. The RBT, due to its low sensitivity on sheep and goats sera, is suggested to be used only for identification of infected flocks (flock screening test) and not for individual animals [15]. Since CFT is regarded as more sensitive and specific, is used for individual testing of animals in infected flocks as well as a confirmatory test [14, 16, 17].

The Rev.1 vaccine was developed by Elberg and Faunce [18] and has been successfully applied in sheep and goat for the control of ovine and caprine brucellosis. It was recognized that Rev.1 vaccination cause existence of positive reactors in serological tests among vaccinated population which lead to difficulties in distinguishing between infected and vaccinated animals by conventional serological tests [19]. Due to these difficulties, the study of the epidemiological situation of the disease is a key element of a successful control program. The gold standard that confirms the presence of the disease is isolation, identification and biotyping of the bacterial agent [20]. Eradication of brucellosis requires accurate diagnosis of the disease among the infected animal population.

## MATERIALS AND METHODS

**Animals:** All animals tested were Egyptian native breeds from farms with a known history of brucellosis according to the directorate of veterinary medicine, Assiut). The samples were taken from slaughtered animals under strict hygienic conditions, kept on ice and sent to our laboratory as soon as possible. The animals all tested positive to at least one of standard tube agglutination test (SAT) and Rose Bengal plate test (RBPT) [12]. Positive samples to the SAT were those with titres  $>1/40$  (50%) according to the European technique [12].

**Sample Collection:** Between May 2009 to May 2010, a total number of 2136 blood samples was collected from lymph nodes (retropharyngeal, prescapular, prefemoral, internal iliac and supramammary) and spleen tissues from carcasses of all serologically positive animals (715 from cattle, 1323 from sheep and 100 from goats) in districts (Al-Badari, Assuit, El-Fath, Abnoub, Manflut, Dyrut and El-Qusia) among Assuit province. Blood samples were allowed to clot and the sera were separated by centrifugation and stored at  $-20^{\circ}\text{C}$  until performing serological tests.

**Serological Examination:** All sera were screened for antibodies against *Brucella* by BAPAT and RBPT as screening tests. All positive serum samples were further retested by SAT and RIV.t as qualitative confirmatory tests described by Alton *et al.* [12].

**Bacteriological Examination:** All obtained tissues were cultured on *Brucella* agar selective media (Oxoid), *Brucella* spp. were identified and biotyped as the methods of Alton *et al.* [12]. This part of study hasn't been published.

## RESULTS AND DISCUSSION

Results obtained by BAPA and RBT as screening tests revealed a positive reactor percentage ranges of 4.6-5.3, 4.4-7.6 and 10-15% followed by overall brucellosis incidence of 4.5, 5.2 and 5.0 % from cattle, sheep and goats at Assuit province, respectively (Table 1). These percentages were similar to that obtained in cattle as 4.8% [21], 4.89% and on sheep as 4.8% [22].

Our percentages were higher than those previously obtained in, cattle as 1.9% [23], sheep as, 1.5% [24-26] and 2.16% [27], 2.31% [28] and in goats as, 4.70 [28], 5.8% [22] and 5.85% [24-26]. Moreover, our percentages were lower than those obtained in, cattle as 6.1% [29], 6.6% [30], 7.1-10 % [31] and 30.6-50 % [32], sheep as, 21.20% [27] and 10.4% [30] and in goats as 14.5% [27].

It is note worthy that no single test can identify all infected animals at all stages of the disease [33, 34] and therefore a combination of serological tests (BAPAT, RBPT, TAT, RIV.T) should be included to reduce the number of both false negative and false positive serological reactions.

It is clearly evident that most of the serological tests used were liable to radical change in their incidence, the great number of false positive detected by BAPA and RBT in the first examination was due to the activity of specific and non-specific antibodies [12].

The results indicated the BAPAT (Table 2), among all tests, gave seroreactors of 91.3 in sheep and 100% in cattle and goats as reported [29], followed by 96.9, 82.6 and 100% using RBPT as reported by Shalaby [35] among examined cows and buffaloes.

Interestingly, SAT and Riv.T. indicated seroreactors of 90.6 and 68.75% in cattle, 65.13 and 73.9% in sheep and 100% in goats, respectively (Table 2). Therefore, the above mentioned results indicated the importance of

Table 1: Results of serological diagnosis of brucellosis by BAPAT and RBPT among animals in Assuit province

Location	Cattle			Sheep			Goats		
	No. of animal sampled	Positive	%	No. of animals sampled	Positive	%	No. of animals sampled	Positive	%
Al-Badari	30	-	-	793	60	7.6	15	-	-
Assuit	170	8	4.7	40	-	-	10	-	-
Abnoub	100	5	5.3	90	-	-	15	-	-
El-Fath	200	10	5	140	-	-	10	-	-
El-Qusia	30	-	-	100	5	5	10	-	-
Manflut	110	5	4.6	90	4	4.4	20	3	15
Dyrut	75	4	5.3	70	-	-	20	2	10
Total	715	32	4.5	1323	69	5.2	100	5	5

Table 2: Seroprevalence of brucellosis among reactors animals in Assuit province based on different confirmatory tests

	Serological tests							
	BAPAT		RBPT		SAT		RIV.T	
Animal species/ Total number	+	%	+	%	+	%	+	%
Cattle/32	32	100	31	96.9	29	90.6	22	68.75
Sheep/69	63	91.3	57	82.6	45	65.1	51	73.9
Goat/5	5	100	5	100	5	100	5	100
Total/106	99	93.39	33	31.13	79	74.52	78	73.58

using several procedures to overcome the problem of escaping of some infected animals in diagnosis of brucellosis as emphasized by Necoletti and Muraschi [36]. Therefore, it is of importance to use more than one diagnostic test for the diagnosis of brucellosis.

In this investigation the highest rate of agreement was between the result of BAPAT and RBT among tested animals which means that should be supported by other confirmatory serological tests. The present results are nearly similar to those previously obtained [37-39] whereas, these authors noticed that BAPAT was similar in its sensitivity to RBT. But it is much higher sensitive than the SAT and Riv. t. in diagnosis of ovine and caprine brucellosis. However, there are some of disagreement obtained between BAPAT and RIV.T among tested animal, thus for the difference in the mode of action of BAPAT detected only IgG<sub>1</sub> and IgG<sub>2</sub> subclasses of immunoglobulin [40] while in RIV.T test case the Rivanol solution (2-ethoxy-6,9 diamino acridine lactate) added to the serum to promotes the reactivity of the IgG<sub>1</sub>, the most indicative isotype of infection, reduces the reactivity of IgG<sub>2</sub> and precipitates IgM, the most commonly associated with the non-specific reaction, [41].

The recovery of *Brucella* spp. from culturing of lymph nodes and spleen of serologically positive slaughtered animals, according to bacteriological isolation, the isolation incidence reached to 28.1% in cattle and 40.5% in sheep and goats. Identification and biotyping of all the recovered isolates confirmed *Brucella melitensis* biovar 3 was the sole type detected in this investigation (these results haven't published yet).

The distribution of collective samples were illustrated in fig. 1 and brucellosis percentages were 56.6, 11.3, 9.4, 7.5, 5.7 and 4.7% in Al-Badary, Manflut and El-Fath, Assuit, Dyrut and Abnoub and El-Qusia, districts, respectively.

*Brucella* strains were isolated from 24 (28%) out of 86 aborted sheep fetus samples. All *Brucella* strains were identified as *B. melitensis* by biochemical tests and PCR. Of the 36 *B. melitensis* isolates, 3, 32 and 1 were identified as biotype 1, biotype 3 and *B. melitensis* Rev-1 vaccine strain, respectively [42].

Using bacteriological isolation as a gold standard by looking for serological profile of bacteriologically positive animals whereas only when the organism could be isolated and identified that give appositive value and 100% infection but negative bacteriological investigation dose not exclude the presence of brucellosis [43].

Table 3: Evaluation of four serological tests in terms of culture results from different infected animal

	Serological tests			
	BAPAT	RBPT	SAT	RIV.T
Sensitivity* (%)	97.4	94.9	84.2	87.6
Specificity** (%)	60	80	81	85.7

\*Sensitivity = (True positive/True positive + false negative) X 100

\*\*Specificity = (True negative/True negative+ false positive) X 100



Fig. 1: Assuit administrative map showing the district (O) of the collected samples. Districts with (\*) indicate the presence of heavy brucellosis infection of examined animals.

On the other hand, *Brucella* organisms were recovered from serologically negative animals, sensitivity and specificity of different serological tests among different animal species according to bacteriological isolation revealed that the highest rate of sensitivity (97.4%) detected by BAPAT (Table 3) is due to the fact that it detects both IgG and IgM molecules [44]. While the RBPT revealed the high rate of sensitivity (94.9%) more than SAT (84.2%) and Riv.T (87.6%) among tested animal, which was similar to that previously reported [45-48].

SAT appeared to have inferior sensitivity if compared with BAPAT and RBT. This coincided with the results obtained by other investigators [49-52]. While, Riv.T., revealed the highest specificity rate of 85.7% (Table 3) and its sensitivity rate was more than that detected by SAT and lower than by BAPAT or RBT, may be due to

the precipitating activity of the Rivanol solution of the IgM, as recorded by Morgan [53, 54] and so the test only detects IgG<sub>1</sub> and IgG<sub>2</sub> immunoglobulins. These results are in agreement to previously reported results [38, 39, 55, 56]. *B. melitensis* strains isolated in Konya region were found to be from different sources by RAPD-PCR. *B. melitensis* biotype 3 was the most common biotype. *B.* [42].

In conclusion, BAPAT and RBPT serological tests revealed the highest rate of sensitivity that guide us to use these tests as screening tests on animals brucellosis. RIV.T showing the highest rate of specificity that bearing in mind the BAPAT and RBT positive samples should be confirmed by this test.

## REFERENCES

1. Mustafa, M. and P. Nicoletti, 1993. Proceeding of the workshop on guidelines for a regional brucellosis control program for the Middle East, pp: 14-17, Amman, Jordan. FAO, WHO and OIE.
2. FAO/WHO, 1986. Expert committee on brucellosis, Sixth Report. WHO Technical Report series, No. 740. WHO, Geneva.
3. Kiel, F.W. and M.Y. Khan, 1987. Analysis of 506 consecutive positive serological tests for brucellosis in Saudi Arabia. J. Clin. Microbiol., 25: 1384-1387.
4. Madkour, M.M., 1989. Brucellosis. Butterworths, London, pp: 294.
5. Radostits, O.M., C.C. Gay, D.C. Blood and K.W. Hinchcliff, 2000. Veterinary Medicine, 9th ed. ELBS Bailliere Tindall, London, UK, pp: 870-871.
6. Blasco, J.M. and B. Molina-Flores, 2011. Control and Eradication of *Brucella melitensis* Infection in Sheep and Goats. Veterinary Clinics of North America: Food Animal Practice, 27: 95-104.
7. Benkirane, A., 2006. Ovine and caprine brucellosis: World distribution and control/eradication strategies in West Asia/North Africa region. Small Rumin. Res., 62: 19-25.
8. Refai, M., 2002. Incidence and control of brucellosis in the Near East region. Vet. Microbiol., 90: 81-110.

9. Hegazy, Y.M., A.L. Ridler and F.J. Guitian, 2009. Assessment and simulation of the implementation of brucellosis control program in an endemic area of the Middle East. *Epidemiol. Infect.*, 137: 1436-1448.
10. Álvarez, J., J.L. Sáez, N. García, C. Serrat, M. Pérez-Sancho, S. González, M.J. Ortega, G. Josep, L. Carbajo, F. Garrido, J. Goyache and L. Domínguez, 2011. Management of an outbreak of brucellosis due to *B. melitensis* in dairy cattle in Spain. *Res. Veterinary Sci.*, 90(2): 208-211.
11. Davies, G., 1971. The Rose Bengal Test. *Vet. Rec.*, 88: 447-449. European Council Directive, 1964. 64/432/EEC. On health problems affecting intra-community trade in bovine animals and swine.
12. Alton, G.G., L.M. Jones, R.D. Angus and J.M. Verger, 1988. Techniques for the brucellosis laboratory, pp: 17-62. Institutional de la Recherche Agronomique, Paris.
13. Garin-Bastuji, B. and J.M. Blasco, 1997. Caprine and ovine brucellosis (excluding *Brucella ovis* infection). In: Manual of Standards for Diagnostic Tests and Vaccines, 3rd ed. Office International des Epizooties, Paris, France, pp: 350-362.
14. European Council Directive, 1991. 91/68/EEC. On animal health conditions governing intra-community trade in ovine and caprine animals.
15. Blasco, J.M., B. Garin-Bastuji, C.M. Marin, G. Gerbier, J. Fanlo, M.P.J. Bagues and C. Cau, 1994. Efficacy of different Rose Bengal and complement fixation antigens for the diagnosis of *Brucella melitensis* infection in sheep and goats. *Vet. Rec.*, 134: 415-420.
16. Nicoletti, P., 1969. Further evaluation of serologic test procedures used to diagnose brucellosis. *Am. J. Vet. Res.*, 30: 1811-1816.
17. MacMillan, A.P., 1990. Conventional serological tests. In: K. Nielsen, J.R. Duncan, (Eds.), Animal Brucellosis. CRC Press, Boca Raton, Florida, pp: 153-197.
18. Elberg, S.S. and K.J. Faunce, 1957. Immunization against *Brucella* infection. VI. Immunity conferred on goat by a nondependent mutant from a streptomycin-dependent mutant strain of *Brucella melitensis*. *J. Bacteriol.*, 73: 211-217.
19. Diaz, R., P. Garatea, L.M. Jones and I. Moriyon, 1979. Radial immunodiffusion test with a *Brucella* polysaccharide antigen for differentiating infected from vaccinated cattle. *J. Clin. Microbiol.*, 10: 37-41.
20. Cunningham, B., 1977. A difficult disease called brucellosis. An international symposium, Taxes A and M. univer.press, collage station, USA., pp: 11-20.
21. Alton, G.G., 1963. A report to the government of United Arab Republic on the control of brucellosis. 1<sup>st</sup> Ed. FAO and WHO, Geneva, Switzerland.
22. Samaha H., T.R. Mohamed, R.M. Khoudair and H.M. Ashour, 2009. Sero-diagnosis of Brucellosis in Cattle and Humans in Egypt. *Immunobiol.*, 214: 223-226.
23. Fayed, A.A., S.A. Karmy, H.I. Yousef and M.M. Ayoub, 1982. Serological study on brucellosis in Aswan governorate. *Vet. Med. J.*, 30: 491-497.
24. Hamada, S., M. El-Hidik, I. Sherif, H. El sawah and M. Yousef, 1963. Serological investigations on brucellosis in cattle, buffaloes and camels. *J. Arab. Vet. Med. Ass.*, 23: 173-178.
25. El-Gibaly, S.M., 1969. Studies on brucellosis in dairy animals in UAR. Ph. D., Faculty of Vet Med., Cairo University.
26. El-Gibaly, S.M., F.M. Goda, S.M. Nada and E.M. Sayour, 1977. Preliminary studies on Epidemiology of brucellosis as a zoonotic disease in an infected area in Egypt. First Arab Biologists Congress, Alex. Egypt, pp: 26-30.
27. Kaoud, H.A., M.M. Zaki, A.R. El-Dahshan and S.A. Nasr, 2010. Epidemiology of brucellosis among farm animals. *Nature and Science*, 8: 190-197.
28. Nada, S.M.M., 1982. Further studies on caprine and ovine brucellosis microorganisms. Ph.D. Thesis, Faculty of Vet. Med., Cairo Univ.
29. Ghobashy, H.M.M., I.A. Samaha, A.M. Montaser, M.K. El-Kholi and S.M. El-Gibaly, 2009. Sero-surveillance on Brucellosis among farm animals in some governorates in Egypt. *Egypt. J. Appl. Sci.*, 24.
30. Yahia, S.I., 1961. Trials for the application of rapid tests in the detection of brucellosis using the stained antigens pro. 2<sup>nd</sup> Arab, Ann. Vet cong. held in Arep., pp: 117-125.
31. Montasser, A.M., M.E., Hamdy, E.M. El-Bayoumy and R.M. Khoudeir, 2001. Bacteriological profile of *Brucella* isolated from cattle in Egypt. *Proc. 6<sup>th</sup> Sci. Cong. Society for cattle diseases*, Assuit, pp: 136-170.
32. Fahmy, S.K. and M.F. Bendary, 1970. Incidence of brucellosis and efficiency of strain 19 vaccine among Frisian herd in A.R.E. *J. Arab. Vet. Med. An.*, 31: 235-242.
33. Morgan, W.J.B., 1971. Some recent advances in the diagnosis of brucellosis. *Irish Vet. J.*, 25: 214-221.
34. Cordes, D.O. and M.E. Carter, 1979. Persistence of *Brucella abortus* infection in six herds of cattle under brucellosis eradication. *New Zealand Veterinary Journal*, 27: 255-259.

35. Shalaby, N.A., 1996. Immunological and bacteriological studies on *Brucella* infection among cows and buffaloes in Egypt. M.V.Sc. Thesis Fac. Vet Med. Cairo Univ.
36. Nicoletti, P. and T.F. Muraschi, 1966. Bacteriologic evaluation of serologic test procedures for the diagnosis of brucellosis in problem cattle herds. Amer. J. Vet. Res., 27: 689-694.
37. El-Bayoumy, E.M., 1989. Some studies on Brucellosis in sheep and goats, M.V.Sc. Faculty of Vet Med., Cairo University.
38. Hamdy, M.E.R., 1992. Epidemiological studies on *Brucella melitensis* in dairy animals and man. Ph.D. Thesis. Faculty of Veterinary Medicine, Cairo University.
39. Anwar, H.K.H., 1999. Abortion in farm animals in Menoufia governorate. Ph.D. Thesis, Faculty of Veterinary Medicine, Suez Canal University.
40. Wright, P.F. and K.H. Nielsen, 1988. Application of enzyme immunoassay in the veterinary medicine serodiagnosis of bovine brucellosis. In: Ngo, T.T. (editor). Nonisotopic immunoassay. Plenum Publishing Corporation.
41. Mikolon, A.B., I.A. Gardner, S.K. Hietala, J.H. Anda, E.C. Pestana, S.G. Hennager and A.J. Edmondson, 1998. Evaluation of North American Antibody Detection Tests for Diagnosis of Brucellosis in Goats. J. Clin. Microbiol., 36: 1716-1722.
42. Aras, Z. and M. Ateş, 2011. The first report of isolation and molecular characterisation of *Brucella melitensis* Rev-1 vaccine strain from an aborted sheep fetus in Turkey. Small Ruminant Res., 95: 150-159.
43. Robertson, L., D. Farrell and P.M. Highliffe, 1977. The isolation of *Brucella melitensis* from contaminated sources. *Brucella* Vet. J., 133: 193-195.
44. Nelson, J.W., 1989. The interpretation of the titre responses: group discussion, Brucellosis seminar, Cairo, Egypt.
45. Alton, G.G., L.M. Jones and D.E. Pietz, 1975. Laboratory techniques in brucellosis. 2<sup>nd</sup> Edition Monograph, World Health Organization, Series No. 55S, Geneva, Switzerland.
46. Montasser, A.M., M.E. Hamdy, S.I. Ibrahim and H.M. Ghobashy, 2002. The impact of serological antibody titers on diagnosis and control of brucellosis in infected farm animals. J. Egypt. Vet. Med. Ass., 62: 167-177.
47. Akhtar, R., Z.I. Chaudhry, A.R. Shakoori, M.D. Ahmad and A. Aslam, 2010. Comparative efficacy of conventional diagnostic methods and evaluation of polymerase chain reaction for the diagnosis of bovine brucellosis. Veterinary World, 3: 53-56.
48. Al-Farwachi, M.I., B.A. Al-Badrani and T.M. Al-Nima, 2010. Detection of *Brucella* antigen in the aborted ovine fetal stomach contents using a modified ELISA test. Iraqi Journal of Veterinary Sciences, 24: 1-4.
49. El-Gibaly, S.M., 1993. Correlation between serotests and isolation of *Brucella melitensis* in an infected sheep pharm. Proc. 2<sup>nd</sup> Sci. Cong. Society for cattle Disease, Assiut, pp: 194-203.
50. Sayour, A.E., 1995. An approach towards the use of some unconventional serological tests for the diagnosis of brucellosis. M.V.Sc. Thesis, Faculty of Vet. Med., Cairo Univ.
51. Hosein, H.I., F.Z. Dawood and M.N. El-sheery, 2002. Evaluation of the policy of test and slaughter for control of brucellosis in Egypt. 10<sup>th</sup> Sci. Cong. Fac. Vet Med. Assiut Univ.
52. Aggad, H., 2003. Serological studies of animal brucellosis in Algeria. Assiut Vet. Med. J., 49: 121-130.
53. Morgan, W.J.B., 1967. The serological diagnosis of bovine brucellosis. Vet. Rec., 80: 616-621.
54. Pietz, D.E. and W.O. Cowart, 1980. Use of epidemiological data and serological tests in bovine brucellosis. Javma, 177: 1221-1226.
55. Kim, J.M., S.C. Jung, J.M. Park, K.J. Hyun and J.S. Mah, 1988. Properties of *Brucella* spp. Isolated from *Brucella* reactor cattle comparison of seven methods for diagnosis. Res. Rep. Of the Rural Dev. Admins. Vet., 30: 1-6.
56. El-Enbaawy, M., J. El-Jakee, A. Fayed and M. Refai, 1995. Evaluation of competitive ELISA in comparison with other conventional tests for detection of bovine brucellosis in Egypt. J. Egypt. Vet. Med. Ass., 55: 769-780.