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Incidence of the Most Common Toxigenic *Aspergillus* Species in Broiler Feeds in Kermanshah Province, West of Iran

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Abstract: The objectives of this study were to investigate the occurrence and identification of *Aspergillus* species in broiler feeds in Kermanshah province, west of Iran. From April 2008 to March 2009, a total of 50 samples of broiler feeds were collected from different broiler farms located in Kermanshah province. All samples were aseptically transported to the laboratory, homogenized, quartered to obtain a 1 kg laboratory sample and were stored at 4°C for fungal analyses. Ten grams of each feed sample were homogenized in 90 ml sterile physiological saline for 30 minutes to obtain a concentration of 10^{-1} (dilution 1). This mixture was then serially diluted to 10^{-2} (dilution 2), 10^{-3} (dilution 3) and 10^{-4} (dilution 4). From each dilution, 25 µL of mixture was deeply inoculated on dichloran rose-bengal-chloranphenicol agar (DRBC) and incubated at 30°C for 15 days. During incubation period, gross and microscopic features of fungal colonies were studied. Out of 50 feed samples (40%) in dilution 4, were contaminated to *Aspergillus* species. The most prevalent species was *Aspergillus flavus* followed by *Aspergillus niger* and *Aspergillus fumigatus*. These results showed that a potential exists for the production of mycotoxins by the *Aspergillus* species. They suggest an association of mycotoxicosis with poultry feeds in western parts of Iran.

Key words: Aspergillus • Poultry feed • Broiler farms • Kermanshah province

INTRODUCTION

Mold and mycotoxin contamination of feed and feed ingredients occurs worldwide and because of the ubiquitous nature of these micro-organisms they cannot be totally eliminated from feeds and ingredients [1]. The presence of mold and mycotoxins in poultry feeds result from the raw material used in their production. Mold and mycotoxins contamination of the raw materials occur during the pre-harvest and/or the post-harvest periods. During these periods, temperature and humidity, as well as processing and handling of animal feed play an important role in the growth of fungi and mycotoxins contamination [2,3]. In general, the mixed feeds of poultry constitute corn and soybean as major ingredients, which represent an excellent substrate for growth and reproduction of numerous fungi, under favorable conditions such as high moisture and increased temperature [4]. When long-term physiological and environmental conditions for fungal growth are provided, mycotoxins are produced, which can not be removed from the feed completely [4]. Fungal contamination is undesirable because of the potential for mycotoxin production [5]. Fungal toxins can be stored in meat, milk and egg and finally transferred to human beings [6].

Most species of *Aspergillus* and *Penicillium* are able to grow on a wide range of organic substrates. They are essentially saprophytic and are particularly associated with stored moldy plant products [7, 8]. *Aspergillus* genera is the most important toxigenic fungi [9, 10].

Corresponding Author: Yousef Azarakhsh, Department of Pathobiology, Faculty of Veterinary Medicine, Razi University, Kermanshah, Iran. E-mail: arashkk_microbiologist@yahoo.com. Poultry are highly susceptible to mycotoxicoses and mycosis [10-12]. Aspergillosis is an increasingly common ubiquitous fungal infection of birds and occasionally other animals including man. *Aspergillus fumigatus* is the most commonly isolated species from the cases of aspergillosis, followed by *Aspergillus flavus* and *Aspergillus niger* [13].

At present, aflatoxins are considered to be one of the most toxic, carcinogenic compounds produced by several members of the *Aspergillus flavus* in foods and feeds [14, 15]. Therefore, throughout the world great attention is paid to investigation on *Aspergillus* species and elaborating means for controlling them [17]. Data on the microbiota and mycotoxins from poultry feeds in Iran are scarce. For this reason, our aim in this study was to isolate and identify the *Aspergillus* species found as contamination in broiler feeds.

MATERIALS AND METHODS

From April 2008 to March 2009, a total of 50 feed samples were taken randomly from commercial broiler farms in Kermanshah province. All samples were aseptically transported to the laboratory, homogenized, quartered to obtain a 1 kg laboratory sample and were stored at 4°C for fungal analyses [16]. Ten grams of each feed sample were homogenized in 90 ml sterile physiological saline for 30 minutes and serial dilutions of 10^{-1} (Dilution 1), 10^{-2} (dilution 2), 10^{-3} (dilution 3) and 10^{-4} (dilution 4) were made. Then, 25 µL of each dilution was deep point inoculated on dichloran rose-bengalchloranphenicol agar [17] and incubated at 30°C for 15 days. Fungal colonies were selected for identification, according to the methods proposed for the genus [18]. The distinct colonies were picked, subcultured for purification and characterized using standard techniques [19]. Wet mount smears and slide cultured colonies were stained with lactophenol cotton blue. Taxonomic identification of the fungi was made based on macroscopic and microscopic features in accordance with appropriate keys [17]. Identification of Aspergillus species were made as per Raper and Fennell [20]. Statistical analysis of data was performed using SPSS software (Version 16) with 95 percent accuracy. A P-value less than 0.05 was considered significant.

RESULTS

Mycological survey of 50 samples of broiler feed from western parts of Iran showed the presence of potentially toxigenic and infective *Aspergillus* species



Fig. 1: Aspergillus flavus, isolated from compounded broiler feeds in west of Iran



Fig. 2: Aspergillus niger, isolated from compounded broiler feeds in west of Iran



Fig. 3: Aspergillus fumigatus isolated from compounded broiler feeds in west of Iran

(Table 1, Figures 1-3). Out of 50 feed samples from each dilution, 46 samples (92%) in dilution 1, 38 samples (76%) in dilution 2, 28 samples (56%) in dilution 3 and 20

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Dilution	Aspergillus species							
	A. flavus		A. niger		A. fumigatus		Total	
	Number	%	Number	%	Number	%	Number	%
1	34	73.9ª	10	21.7 ^b	2	4.3°	46	92.0
2	26	68.4ª	10	26.3 ^b	2	5.2°	38	76.0
3	22	78.5ª	4	14.2 ^b	2	7.1 ^b	28	56.0
4	14	70.0ª	4	20.0 ^b	2	10.0 ^b	20	40.0

Table 1: The occurrence of Aspergillus species isolated from 50 broiler feed samples in west of Iran

^{a, b, c} Figures with different superscripts within rows are significantly different (p<0.05)

samples (40%) in dilution 4, were contaminated to *Aspergillus* species. In dilutions 1, 2, 3 and 4, the highest contamination rate belonged to *Aspergillus flavus* (73.9, 68.4, 78.5 and 70%, respectively), followed by *Aspergillus niger* (20, 14.2, 26.3 and 21.7%, respectively) and *Aspergillus fumigatus* (4.3%, 5.2%, 7.1% and 10%). There was a significant difference between the contamination rate to *Aspergillus flavus* and the other isolated *Aspergillus species* (p < 0.05).

DISCUSSION

Mycological survey of 50 samples of broiler feed from western parts of Iran showed the presence of potentially toxigenic and infective Aspergillus species (Table 1). Many researchers have proved that the majority of feeds have species from Aspergillus and Penicillium genera as a predominant flora; Bragulat et at., (1995) alreported high frequency of Aspergillus species in mixed poultry feeds [21]. Magnoli et al. [25] lreported a high frequency of Aspergillus group species found in poultry feeds from Argentina [3]. They reported that Aspergillus flavus and A. parasiticus, which are important aflatoxin producers, were the predominant species isolated. Glenda, et al. [11] reported that Aspergillus and Penicillium species had the highest isolation frequencies followed by Fusarium spp in poultry feeds in Brazil. Many studies have shown that most feeds have species from Aspergillus and Penicillium genera as predominant flora [21, 22].

In the present study, the main contaminating fungus appeared to be *A. flavus*, a potentially toxigenic species for the aflatoxins. Likewise, the species has shown high occurrence frequency in the studies of Labuda and Tancinova [3], Heperkan and Alperden [23], Magnoli *et al.* [24] and Dalcero *et al.* [25] and There was a significant difference between the rate of contamination to *Aspergillus flavus* and other isolated *Aspergillus*

species (*A. niger* and *A. fumigatus*) in 4 dilutions (p < 0.05), similar results obtained from mycological survey on feed ingredients and mixed animal feeds in Ghom province (central parts of Iran) were reported by Khosravi *et al.* [26].

A. flavus was the most prevalent species. This result agrees with Adebajo et al. Dalcero et al. [27, 28]. Magnoli et al. [3] and Accensi et al. [29]. Dutta and Das [4] confirmed the predominance of A. flavus over A. parasiticus in poultry feeds. The majority of these genera representatives such as A. flavus are thermophilic and thermo-resistant and distribute abundantly in tropical to subtropical climates [30]. Lacey and Magan [21] showed that the ideal temperature concerning growth and mycotoxin production ranges 25 to 35°C for A. flavus strains [7, 8]. The average annual range of temperatures in western parts of Iran varies from 21 to 28°C, but is generally more than 24°C indicating favorable condition for A. flavus growth [31, 32].

Among the *Aspergillus* species isolated from broiler feeds, *A. niger* was the second most prevalent species. This is in agreement with Osho *et al.* [29] who reported that out of the 50 samples collected from various commercial poultry farms located in southwest Nigeria, *A. niger* was one of the common fungi found in the feeds [10]. *Rhizopus* spp. had the highest frequency of occurrence (44%), *Fusarium* spp. 42%, *A. flavus* 40%, occurrence and *A. niger*, (38%).

The occurrence of *Aspergillus* species in broilers feed is particularly important because there are known as the most toxigenic among the fungi. Most studies indicate that there is no correlation between the presence of a toxin and the producing fungus in the same substrate, but the presence of toxigenic fungi in feeds may be an indicative of their potentiality to produce mycotoxins. When the storage conditions are not appropriate and the toxigenic fungus is present, this may be able to produce a mycotoxin [33]. The results of this study showed that the broiler feeds in Kermanshah province were highly (92% in dilution 1 and 40% in dilution 4) contaminated with *Aspergillus* species which are the most common toxigenic fungi found in feeds. This study warrant the need for analyzing the samples for *Aspergillus* mycotoxins, especially aflatoxins and also to design effective management strategies to prevent contamination of poultry feed to *Aspergillus species* and *Aflatoxin*. The study highlights a potential risk of poultry feeds getting contaminated with hazardous toxic compound and potentially infective *A. fumigatus*, thus making it for further analysis and continual monitoring and evaluation of feeds.

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REFERENCES

- 1. Accensi, F., M.L. Abarca and F.J. Cabanes, 2004. Occurrence of Aspergillus species in mixed feeds and component raw materials and their ability to produce ochratoxin A. Food Microbiol., 21: 623-627.
- Adebajo, L.O., A.A. Idowu and O.O. Adesanya, 1994. Mycobiota and mycotoxins production in Nigeria corn and corn-based snacks. Mycopathologia, 126: 183-192.
- Badripour, H., 2010. Country pasture profiles. http://www.fao.org/ag/AGP/AGPC/doc/Counprof/Ir an/Iran.htm#main.
- 4. Bankole, S.A. and K.A. Kpodo, 2005. Mycotoxin contamination in food systems in West and Central Africa. In Reducing the impact of Mycotoxin in Tropical Agriculture with emphasis on health and trade in Africa, Accra, Ghana, pp: 13-16.
- Baron, E.J.O., L.R. Peterson and S.M. Finegold, 1994. Bailey and Scott's Diagnostic Microbiology. 9th. edition. Mosby, Baltimore.
- Bragulat, M.R., M.L. Abarca, O. Castella and J. Cabanes, 1995. Mycological survey on mixed poultry feeds and mixed rabbit feeds. J. Sci. of Food and Agric., 67: 215-220.
- Cotty, J.P., 1997. Aflatoxin-producing potential communities of *Aspergillus* section *flavi* from cotton producing areas in the United States. Mycology Res., 101: 698-704.

- Dalcero, A., C. Magnoli, S. Chiacchiera, G. Palacios and M. Reynoso, 1997. Mycobiota and incidence of aflatoxin B1, zearalenone and deoxinyvalenol in poultry feeds in Argentina. Mycopathologia, 137: 179-184.
- Dalcero, A., C. Magnoli, M. Luna G. Ancasi, M.M. Reynoso, S. Chiachiera, R. Miazzo and G. Palacio, 1998. Mycoflora and naturally occurring mycotoxins in poultry feeds in Argentina. Mycopathologia, 141: 37-43.
- Dutta, T.K. and P. Das, 2000. Isolation of aflatoxigenic strains of *Aspergillus* and detection of aflatoxin B1 from feeds in India. Mycopathologia, 151: 29-33.
- 11. Glenda, R., J. Oliveira, M. Ribeiro, M.E. Fraga, L.R. Cavaglieri, G.M. Direito, K.M. Keller, A.M. Dalcero and C.A. Rosa. 2006. Mycobiota in poultry feeds and natural occurrence of aflatoxins, fumonisins and zearalenone in the Rio de Janeiro State, Brazil, Mycopathologia, 162: 355-362.
- Gourama, H. and L. Bullerman, 1995. Aspergillus flavus and A. parasiticus: Aflatoxigenic fungi of concern in foods and feeds: A review. J. Food Protection, 58: 1395-1404.
- Heperkan, D. and I. Alperden, 1988. Mycological survey of chicken feed and some feed ingredients in Turkey. J. Food Protection, 51: 807-810.
- Joffe, A.Z., 1983. *Fusarium* as field, stored and soil fungi under semiarid conditions in Israel. In: Y. Ueno (ed.), trichothecenes-chemical, biological and toxicological aspects. Oxford, pp: 95-110.
- Khosravi, A.R., M. Dakhili and H. Shokri, 2008. A mycological survey on feed ingredients and mixed animal feeds in Ghom province, Iran, Pakistan J. Nutrition, 7: 31-34.
- King, A.D., A.D. Hocking and J.I. Pitt, 1979. Dichloran-rose bengal medium for enumeration of molds from foods. Applied Environmental Microbiol., 37: 959-964.
- 17. Klich, M.A., 2002. Identification of Common Aspergillus Species. Utrecht, Netherlands, CBS.
- Kubena, L.F., R.B. Harvey and W.E. Huff, 1993. Efficacy of a hydrated sodium calcium aluminosilicate to reduce the toxicity of aflatoxin. Poultry Sci., 72: 51-59.
- Labuda, R. and D. Tancinova, 2006. Fungi recovered in poultry feed mixture. Annals of Agricultural and Environmental Medicine, 13: 193-200.

- Lacey, J., 1991. Natural occurrence of mycotoxins in growing and conserved crops. In: J.E. Smith and R.S. Henderson, (eds), Mycotoxins and Animal Foods. CRC Press, pp: 363-390.
- Lacey, J. and N. Magan, 1991. Fungi in cereal grains, their occurrence and water and temperature relationships. In: J. Chelkowski, (ed), Cereal Grain, Mycotoxins, Fungi and Quality in Drying and Storage. Elsevier, Amsterdam, pp: 77-118.
- Lambuda, R., D. Tamcinova and K. Hudec, 2003. Identification and enumeration of *Fusarium* species in poultry feed mixtures from Slovakia. Annals of Agricultural and Environmental Medicine, 10: 61-66.
- Lebars-Bailly, S., J.D. Bailly and H. Brugere, 1999. Accidents due to cheese molds manufacturing. Revue de Médecine Vétérinaire, 150: 413-430.
- Lozada, A.F., 1995. Isolation and identification of mycotoxigenic fungi in selected foods and feeds. Food Additives and Contaminants, 3: 509-514.
- Magnoli, C., A. Dalcero, S.M. Chiacchiera, R. Miazzo and M. Sa' enz, 1998. Enumeration and identification of *Aspergillus* group and *Penicillium* species in poultry feeds in Argentina. Mycopathologia, 142: 27-32.
- Maria, L., M. Marina and B. Fernando, 2003. Fungal flora and mycotoxins detection in commercial pet food, Lisboa, Portugal, 98: 179-183.
- Nyamongo, J. and M. Okioma, 2005. The aflatoxin outbreaks in Kenya in 2004 and 2005: a case study. In: Reducing impact of Mycotoxins in Tropical Agriculture with emphasis on Health and Trade in Africa, Accra, Ghana, pp: 13-16.

- Opera, M.N. and I.C. Okoli, 2005. Strategies for reduction of mycotoxin contaminations in Animal productions panacea for the problems in Southeastern Nigeria. In: Reducing impact of Mycotoxins in Tropical Agriculture with emphasis on Health and Trade in Africa. Accra, Ghana, 66: 13-16.
- Osho, I.B., T.A.M. Awoniyi and A.I. Adebayo, 2007. Mycological investigation of compounded poultry feeds used in poultry farms in southwest Nigeria, African J. Biotechnol., 6: 1833-1836.
- Pattron, D.D., 2006. *Aspergillus*, Health implication and recommendations for public health food safety. Internet J. Food Safety, 8: 19-23.
- Pitt, J.I. and A.D. Hocking, 1997. Fungi and Food Spoilage, 2nd ed. Blackie Academic and Professional, London.
- 32. Pitt, J.I., 2000. Toxigenic fungi: which are important? Medical Mycol., 138: 17-22.
- Raper, K.D. and D.I. Fennel, 1965. The genus Aspergillus. Bultimore, The Williams and Wilkins Co., pp: 357-404.
- Rippon, J.W., 1988. Medical Mycology. 2nd ed. W.B. Saunders Company, Chicago, Illinois, USA, pp: 640-641.
- Trenholm, H.L., L.L. Charmley and D. PreLusky, 2000. Mycotoxin binding agents: An Update. Farming Today, 1: 11.