

Some Biochemical and Immunogenetic Investigations on Buffaloes Suffering from Chronic Endometritis

Samy I.A. Shalaby, Wahid M. Ahmed and Magdy M. Zaabal

Department of Animal Reproduction and A.I.,
Veterinary Research Division, National Research Centre, Giza, Egypt

Abstract: Endometritis is among the main cause of failure of conception in buffaloes in many developing countries including Egypt. In this study the prevalence of the affection and blood changes in affected buffalo-cow were investigated as compared with normal fertile partners. A total number of 385 buffalo-cows was investigated. Blood samples were collected and plasma was analyzed for some biochemical and genetic components. Results indicated that 5.97% of examined buffalo-cows suffer from chronic endometritis. Affected animals revealed high cortisol ($p < 0.01$), A1P, LDH and AST activities ($P < 0.01$), iron ($p < 0.01$) and creatinine ($P < 0.01$) and low progesterone ($P < 0.01$), total lipid ($P < 0.05$), zinc ($P < 0.05$) and copper ($p < 0.05$) concentration in their blood as compared with normal fertile animal. Total proteins and urea values were not significantly changed in buffalo-cows suffering from endometritis. Electrophoretic pattern of plasma protein in buffalo-cows with chronic endometritis was characterized by significant changes in proalbumin ($P < 0.05$), post albumin ($P < 0.05$) and transferrin ($P < 0.01$) values compared to normal cyclic buffaloes. Distribution of serum protein genotypes and estimation of their gene frequencies indicated that the most predominant genetic alleles in buffalo-cows with endometritis are Al^S (0.714) and PtF^B (0.643) while in healthy animals, Al^F (0.684), Pal^A (0.833), $F\alpha_2^A$ (0.684) and Tf^D (0.763) alleles predominated. It was concluded that chronic endometritis is usually associated with disturbed blood constituents such as steroids, total lipids, enzymes, trace elements and protein electrophoresis.

Key words: Buffalo • Endometritis • Biochemical parameters • Genotypes

INTRODUCTION

Buffalo has an important role in the agricultural economy of several developing countries, whereas, such animals significantly contribute to the total milk, meat and work production. Despite having several merits such as utilization of poor-quality roughage, adaptation on harsh environments and resistance to several bovine tropical diseases, the buffalo has relatively poor reproductive efficiency irrespective of their location throughout the world [1].

During peripartum period, animals undergo pronounced physiological changes that suppress both the cellular and humoral defense mechanisms of the hosts thus increasing their susceptibility to various uterine infections. Further, the physiological barriers of reproductive tract are weakened at parturition and

the commensals take upper hand in creating infection more so when the hygienic surroundings are not at their best [2].

A large proportion of failure of conception in farm animals could be attributed to endometritis [1]. Such affection is the most common ailment under field/farm conditions in buffaloes causing decreased fertility resulting in high economic losses [3, 4].

In Egypt, endometritis is an important cause of reproductive failure among buffaloes. It represents 87.3-93.5% from infectious causes of infertility. The incidence was recorded as 4.5- 27% in abattoir surveys and 2.4-27% in living buffaloes [4]. Many etiological factors have been incriminated as a cause of endometritis among which, hereditary, nutritional, managemental, hormonal, disturbance, specific and nonspecific infectious factors [5].

Several authors [4-9] indicated that endometritis induced biochemical alterations in the blood of affected farm animals.

Reproductive performance may be changed as a result of changes in frequency of different genotypes. An increase in proportion of homozygotic loci is typically associated with a decrease in performance and vice versa for heterozygosity [10]. Moreover, correlation between immunogenetics and liability for infection and/or resistance has been recorded [11-13]. There are many studies performed to identify genes coding for productive, reproductive and economically significant traits by analysis of the whole-genome sequence of swamp and river buffaloes [14-16].

The present study was carried out to build a composite diagnostic approach for endometritis in buffaloes at an early stage through investigating some biochemical and immunogenetic aspects in the blood buffaloes suffering from chronic endometritis as compared to normal fertile partners.

MATERIALS AND METHODS

Animals: A total number of 385 buffalo-cows (at least 6 months after last calving) was investigated at Veterinary Clinics and small private farms nearby great Cairo, Egypt. Owner complains, case history and general health status were recorded. Gynaecological examination was performed by rectal palpation aided by ultrasonography with an endorectal array (8.6 M Hertz; Pia Medical Flacse Saote, Netherland) at least twice for two successive weeks and the reproductive status and/or disorders were recorded. Cases revealing chronic endometritis were registered taking in consideration the criteria of reported by Robert [3] such as exudation, congestion, erosion of lining epithelium and enlargement.

Samples: Blood samples on EDTA were collected from buffalo-cows (normal cyclic at luteal phase and those revealing picture of chronic endometritis). Plasma was separated by centrifugation at 3000 r.p.m. for 15 minutes and kept at -20°C until biochemical analysis.

Hormonal Analysis: Plasma progesterone and cortisol levels were assayed using ELIZA micro wells technique, kits from Novotec, Germany [17] and ELIZA reader (Anthos Zenyth 200rt).

Some Other Biochemical Parameters: Glucose, cholesterol, total lipids, triglycerides, total protein, urea, creatinine, calcium, inorganic phosphorus, magnesium,

alkaline phosphatase (AIP), lactate dehydrogenase (LDH) and transaminase (AST, ALT) were calorimetrically determined [18]. Proteins were electrophoretically analyzed using polyacrylamide gel [19].

Immunogenetic Investigation: Gene distribution frequency was determined [20] using the following formula: $P^2 + 2pq + q^2 = 1$ where p and q are the homozygotic genotypes (AA) and (BB) respectively. In the present study 6 blood protein loci were analyzed by electrophoresis, Pre-albumin (Pr) – Albumin (Alb)-post-albumin (Pal)- α -globulin ($F\alpha_2$) -transferrin (Tf) and post-transferrin (Ptf).

Statistical Analysis: Means were calculated for buffalo-cows suffering from endometritis and normal cyclic ones at the luteal phase. Student (t) test and χ^2 were used for studying the differences between the above mentioned groups [21].

RESULTS

Examinations of 385 buffalo cows revealed that 23 animals suffer from chronic endometritis with an incidence of 5.97%.

Progesterone level was low ($P < 0.01$) while, cortisol level was high ($P < 0.01$) in affected animals as compared with normal animals (Table 1).

Energy metabolites and cholesterol revealed little changes in buffalo-cows suffering from endometritis except for total lipids which was significantly decreased ($P < 0.05$) compared to normal (Table 1).

Narrow C/P ratio ($P < 0.05$) with high iron ($P < 0.01$) and low zinc ($P < 0.05$) and copper ($P < 0.05$) were the main characteristic alterations in mineral profiles due to chronic endometritis in buffalo-cows (Table 1).

Total protein and urea values were not significantly changed due to endometritis (Table 1) While, creatinine is high ($P < 0.01$) in affected animals (Table 1). On the other hand, electrophoretic analysis indicated that low albumin ($P < 0.05$), postalbumin ($P < 0.05$) and post transferrin ($P < 0.01$) values were obvious in the plasma of buffalo-cows suffered from chronic endometritis compared to normal cyclic one (Table 1).

Distribution of serum protein genotypes and estimation of their gene frequencies indicated that the most predominant genetic alleles in buffalo-cows with endometritis are Al^S (0.714) and Pt^B (0.643) while in healthy animals, Al^F (0.684), Pal^A (0.833), $F\alpha_2^A$ (0.684) and Tf^D (0.763) alleles predominated (Table 2).

Table 1: Some Biochemical paramters in buffalo-cows suffering from chronic endometritis (Mean \pm S.E.)

Parameter (mg/dL)	Normal cyclic (luteal)	Endometritis
Progesterone (ng/ml)	4.21 \pm 0.52	1.87 \pm 0.17**
Cortisol (μ g/dL)	1.31 \pm 0.23	3.11 \pm 0.19**
Glucose(mg/dL)	67.55 \pm 3.08	71.28 \pm 4.54
Cholesterol(mg/dL)	201.65 \pm 4.58	183.78 \pm 9.90
Total lipids(mg/dL)	416.80 \pm 13.26	367.73 \pm 19.36*
Triglycerides(mg/dL)	81.20 \pm 1.84	81.85 \pm 2.27
AIP U/L)	12.99 \pm 0.39	21.62 \pm 1.84**
LDH U/L)	45.63 \pm 3.12	56.80 \pm 2.26**
AST U/L)	31.30 \pm 2.84	51.14 \pm 2.42**
ALT U/L)	12.70 \pm 0.11	12.51 \pm 0.10**
Calcium (mg/dL)	10.54 \pm 0.34	9.93 \pm 0.24
Inorg.phosphorous	5.08 \pm 0.26	5.24 \pm 0.92
Calcium: phosphorous	2.08 \pm 0.30	1.21 \pm 0.19*
Mg (mg/dL)	3.54 \pm 0.04	3.54 \pm 0.20
Iron(μ g/dL)	217.15 \pm 5.63	250.14 \pm 11.73**
Zinc (μ g/dL)	108.75 \pm 4.16	86.57 \pm 8.24*
Copper (μ g/dL)	90.63 \pm 5.57	70.86 \pm 8.24*
Total proteins	6.52 \pm 0.13	6.35 \pm 0.20
Urea (mg/dL)	33.80 \pm 1.56	35.45 \pm 2.70
Creatinine (mg/dL)	2.00 \pm 0.80	2.35 \pm 0.03**
Electrophoresis (%)		
Prelabumin	8.55 \pm 1.89	6.96 \pm 0.33
Albumin	18.34 \pm 1.41	13.46 \pm 1.71*
Postalbumin	5.82 \pm 0.36	3.42 \pm 0.68**
A-globulin	6.03 \pm 1.02	4.87 \pm 1.82
Transferrin	16.33 \pm 1.02	23.78 \pm 6.33
Posttransferrin	12.28 \pm 1.47	5.83 \pm 0.84**

*p<0.05 **p<0.01

Table 2: Distribution of plasma protein genotype in buffalo-cows suffering from endometritis (Mean \pm S.E.)

Protein		Normal cyclic (luteal)	Endometritis
Pre-albumin	AA	7 (5.200)	2 (1.750)
	AB	6(9.500)	3 (3.500)
	BB	6 (4.300)	2 (1.750)
	Gene frequency	Pr ^A =0.526 Pr ^B =0.474	Pr ^A =0.500 Pr ^B =0.500
	X ²	2.600	0.130
Albumin	FF	7 (8.900)	3 (3.500)
	FS	12 (8.200)	4 (4.900)
	SS	- (1.900)	- (0.600)
	Gene frequency	Alb ^F =0.684 Alb ^S =0.316	Alb ^F =0.714 Alb ^S =0.286
	X ²	4.000*	1.400
Post-albumin	AA	11 (10.400)	4 (3.500)
	AB	3 (4.200)	2 (2.900)
	BB	1 (0.400)	1 (0.600)
	Gene frequency	Pal ^A =0.833 Pal ^B =0.167	Pal ^A = 0.714 Pal ^B =0.286
	X ²	1.200	0.640
A-globlin	AA	4 (1.900)	2 (0.900)
	AB	4 (8.200)	1 (3.200)
	BB	11 (8.900)	4 (2.900)
	Gene frequency	F α_2 A =0.316 F α_2 B =0.684	F α_2 A = 0.357 F α_2 B =0.643
	X ²	4.900*	3.22**
Transferrin	DD	12 (11.000)	2 (2.900)
	DE	5 (6.900)	3 (3.200)
	EE	2 (1.100)	2 (0.900)
	Gene frequency	Tf ^D =0.763 Tf ^E =0.237	Tf ^D =0.643 Tf ^E =0.357
	X ²	1.300	1.600
Postransderrin	AA	7 (5.800)	2 (0.900)
	AB	7 (9.400)	1 (3.200)
	BB	5 (3.800)	4 (2.9000)
	Gene frequency	Ptf ^A = 0.553 Ptf ^B = 0.447	Ptf ^A = 0.357 Ptf ^B = 0.643
	X ²	1.200	3.200**

*p > 0.05 **p > 0.01

DISCUSSION

Endometritis is among main causes of low reproductive potentials in buffaloes and causes great financial losses represented by the cost of therapy, failure of conception and repeat breeding [1, 22]. Since the endometrium is central to the control of reproductive events such as luteolysis, maternal recognition of pregnancy, early embryo development and placentation in the ruminants, it is logical to assume that its inflammation would cause infertility in the ruminants [22]. In fact, postpartum (PP) endometritis which occurs during the first three to six weeks of calving adversely affects the fertility indices in cattle. The condition not only affect the uterus but also the ovary [23] and may be due to various factors viz. dystocia, retained foetal membranes (RFM), caesarean section, contamination of uterus at the time of artificial insemination or natural insemination, induced parturition, stillbirth, twins, ovarian inactivity, cystic ovaries, unhygienic calving environment and metabolic disorder associated with parturient metabolic disorders [24].

In the present study approximately 6% of the examined buffalo-cows suffered from endometritis. This incidence lies within ranges given by Vale *et al.* [25] and Youssein *et al.* [26] as 4.5-7.5%. However, high incidence (11.9-14.6%) was given by El-Hariri *et al.* [27]. Variations in the reported incidence was related to milk production, managemental system and endocrine dysfunction [3, 28].

Progesterone level was lower while, cortisol level was higher in the affected animal compared to values of normal luteal phase. In this respect, Ismail *et al.* [29] concluded that uterine disorder disturbed steroidogenesis. The condition is mainly due to replacement of luteal cells with fibroblast in pathologically persisted corpora lutea [5] as well as the efficiency of the affected endometrium to produce prostaglandin $F\alpha_2$ [30]. Moreover, the type of microorganism could not be denied [7]. On the other hand, the high cortisol level agree with Zrally *et al.* [31] and Ezzo *et al.* [32] who related the condition to the stressful circumstances of the affected animals as the high cortisol level minimize the inflammatory reaction and suppresses the allergic response to various antigens.

Buffalo-cows suffering from chronic endometritis had low plasma level of energy metabolites and cholesterol, especially total lipids. Majeed *et al.* [33] found that buffaloes with metritis showed increased serum glucose level. Ash-turker *et al.* [34] revealed low total cholesterol

level in infertile buffalo, while Salem *et al.* [35] and Yassein *et al.* [36] reported high cholesterol and total lipid in repeat breeder bovine, respectively. These conflicting results were mainly related to differences in the managerial systems of the affected animals.

Concerning AIP, AST and LDH higher activity in affected buffalo-cows, many authors [9, 24, 37] indicated that the higher enzymatic activity in infertile compared to fertile animals mainly reflected the hypermetabolic rate and stress in the affected animals [9]. Moreover, it was recorded that increased enzymatic activity in the uterine flushing could be diagnostic criteria in the assessment of the intensity of endometritis [24].

Changed C/P ratio with high iron and low zinc and copper concentrations were evident in the plasma of the affected buffalo-cows. However, balanced C/P ratio is essential for animal pituitary function and fertility [8]. Concerning iron, similar results were reported by Hahn *et al.* [37] who found high iron concentration, while Moustafa *et al.* [38] found low concentration in repeat breeder bovines. On the other hand, Hahn *et al.* [37] and Moustafa *et al.* [38] reported low copper concentration, while El-Taweel [39] found high value and Nada *et al.* [5] found no significant changes in plasma copper concentration in buffalo-cows with endometritis. Zinc concentration was low in repeat breeder [33] and did not changed in buffalo-cows with endometritis [7]. However, Ahmed *et al.* [8] concluded that mineral homeostasis is essential for normal fertility in farm animals. No significant changes could be detected in total plasma proteins in buffalo-cows with endometritis. However, creatinine value was high, while albumin and α -globulin values were low with high transferrin. Similar results were previously reported [34-37].

In the present study, the distribution of genotypes and gene frequencies of studied loci indicated that the albumin genotype SS ($Alb^S=0.684$), globulin genotype BB ($F\alpha_2^B=0.643$) and post-transferrin genotype BB ($Ptf^B=0.643$) were mostly associated with endometritis. and this finding agree with data obtained by Osman [40] who reported association of homozygote genotypes with endometritis in Egyptian buffaloes. On the other hand, the current results revealed more predominance of genetic alleles in were Alb^F (0.684), Pal^A (0.833) and Tf^D (0.763) in normal cyclic buffalo-cows and this result is similar to those previously recorded by Zheng *et al.* [41], Shalaby *et al.* [42] and Elkhadrawy *et al.* [43]. The possible correlation between some genetic markers and animal resistance could be attributed to the amplifying synthesis of $TNF-\alpha$ and $IL-1\beta$ genes that contribute to

protective immune response and exert anti-microbial action. Moreover, Yi *et al.* [12] recorded a significant association between CYP19 polymorphism and endometritis. Also, Anurag *et al.* [13] found a polymorphism in the promotor region of TNF- α of water buffalo (*Bubalus bubalis*) which is associated with disease resistant. Molecular techniques have been used recently in the diagnosis of endometritis in different livestock. Several researches have focused on the transcriptional profile of TLRs and proinflammatory cytokines in peripheral blood or tissues in cows and mares and there was a significant correlation between the expression of these genes and the presence of endometritis [44]. TLR2 and TLR4 are members of the TLR family which is highly expressed in the endometrial cells of the uterus and they play a key link between endometritis and immune system response, therefore they can be considered as detection markers for endometritis [45]. Toll-like receptors play an essential role in innate immunity through recognition of pathogen-associated molecular patterns (PAMPs) and activation of proper innate and adaptive immune responses against pathogens [46, 47].

In conclusion, chronic endometritis is usually associated with disturbed blood constituents such as steroids, total lipids, enzymes, trace elements and protein electrophoresis. For future strategy to increase productivity and to improve reproductive efficiency of Egyptian buffalo, it is very important to identify a specific genotypes associated with resistance and susceptibility to occurrence of endometritis which help in genetic selection in breeding program.

REFERENCES

- Gahlot, S.C., S. Kumar, A. Kumaresan, S. Vairamuthu, K.K. Saraf, L. Sreela, R.K. Baithalu, S.S. Lathwal and T.K. Mohanty, 2017. Biochemical analysis of uterine fluid for identification of indicators for subclinical endometritis in the water buffalo (*Bubalus bubalis*). *Reprod Domes. Anim.* Assessed on line at DOI: 10.1111/rda.13051
- Bhadaniya, R., M.C. Prasad, H.H. Savsani, V.A. Kalaria, D.T. Fefar, B.S. Mathpati and B.B. Javia, 2016. Pro-inflammatory cytokine expression studies of subclinical and clinical endometritis in endometrial tissues of buffaloes. *Tropical Animal Health and Production* on line: <https://doi.org/10.1007/s11250-019-01802-8>.
- Roberts, S.J., 1986. *Veterinary Obstetrics and Genital Disease*. 3rd ed. Wood Stock.
- Hemeida, N.A., 1988. Reproductive problems in buffaloes in the world *proc. 2nd World Buff. Cong.* New Delhi, India, pp: 197-205.
- Nada, A.R., W.M. Ahmed and A.S. Abdoon, 1993. Studies on endometritis in buffalo cows-proteome level in relation to infection. *Egypt. J. Comp. Path. Clin. Path.*, 6: 231-238.
- El-Bagdady, Y.R., 1984. Studies on some serum organic constituents during diffealo cows. *J. Egypt. Vet. Med. Assoc.*, 44: 17.
- Ahmed, Y.F., 1987. Hormone profile in relation to some reproductive changes in the buffalo cow. Ph.D. Vet. (Pathology), Cairo University.
- Ahmed, W.M., E.M. Ismail, M.R. Shalash and N.A. Hemeida, 1993. Minereals profiles during pregnancy and peri partum of native cow. *Anim. Sci. Abroad*, 20: 27-28.
- Ahmed, W.M. and S.I.A. Shalaby, 1993. Some enzymatic activities in sera and uterine homogenates of buffaloes and cows during some reproductive phases and disorders. *Indian J. Anim. Sci.*, 63: 1163-1139.
- Bradfors, G.E., J.L. Spearow and J.P. Hanrahan, 1991. Genetic variation an improvement in reproduction. In "Reproduction in Domestic Animals". 4th ed. Academic Press, pp: 605-636.
- Zaabal, M.M., A.M. Hamam, A.S. Mohsen and N.N. Bucataru, 1994. Relationship between immunogenetic structures and resisittance to some infectious disease in cattle. *Egypt. J. Immun.*, 1: 113-117.
- Yi, K., L.Y. Yang, Z. Lan and M. Rong, 2016. The association between CYP19 polymorphism and endometrosis risk: a system review and meta analysis. *European Journal of Obstetrics and Gynecology and Reproductive Biology*, 199: 42-48.
- Anurag, S., S.K. Mishra, S. Lavokumar, V.S. Karan, K. Namit, S. Monika, M. Mukes, S.K. Niranjan, K. Avnish and D.S. Kataria, 2019. Detection of polymorphism in the region of TNF-alpha gene of water buffalo (*Bubalus bubalis*) and its association with disease resistance. *Indian J. Animal Research*, 53(12): 1572-1576.
- Sun, T., J. Shen, A. Achilli, N. Chen, Q. Chen, R. Dang, Z. Zheng, H. Zhang, X. Zhang and S. Wang, 2020. Genomic analyses reveal distinct genetic architectures and selective pressures in buffaloes. *Giga Science*, 9(2): 1-12.
- Machado, V.S. and R.C. Bicalho, 2015. The infectious disease epidemiologic triangle of bovine uterine diseases. *Anim. Reprod*, 12(3): 450-464.

16. Osman, N.M. A.A. Abou Mossallam, F.R. El-Seedy and E.R. Mahfouz, 2018. Single Nucleotide Polymorphisms in TLR4 Gene and Endometritis Resistance in River Buffalo (*Bubalus bubalis*). Jordan Journal of Biological Sciences, 11(5): 577-583.
17. Wisdom, G.B., 1976. Competitive immunoenzymatic colorimetric method for quantitative determination of progesterone in serum or plasma. Clin. Chem., 22(8): 1243-1255.
18. Wilding, P. and J.H. Kennedy, 1977. Manual of Routnr Method in Clinical Chemistry for use in Intermediate Laboratories, WHO.
19. Carlstrom, A. and B.G. Johnson, 1983. Electrophoresis immunofixation. Scand. J. Immunology, 17: 23-30.
20. Merkoreva, E.K., 1977. Genetic Base of Selction in Faom Animals. Moscow, Koloc, pp: 121.
21. Snedecor, G.W. and W.G. Cochran, 1976. Statistical Methods, 6 Univ. Press. USA.
22. Kharayat, N.S., C. Sharma, G.R. Kumar, D. Bisht, G. Chaudhary, S.K. Singh, G.K. Das, A.K. Garg, H.K. and N. Krishnaswamy, 2019. Differential expression of endometrial toll-like receptors (TLRs) and antimicrobial peptides (AMPs) in the buffalo (*Bubalus bubalis*) with endometritis Veterinary Research Communications 2019. <https://doi.org/10.1007/s11259-019-09761-z>
23. Behera, B.K., C.G. Sharma, S.K. Singh, H. Kumar, R.K. Chaudhari, A.S. Mahla, G.K. Das and N. Krishnaswamy, 2016. Relationship between endometritis and oxidative stress in the follicular fluid and luteal function in the buffalo. Reprod Domes. Anim. Reprod Domes. Anim. DOI: 10.1111/rda.
24. Sharma, P. and S. Srivastava, 2018. Comparative efficacy of ceftiofur sodium, gentamicin sulfate and *Zingiber officinale* On physical characteristics of cervical mucus in endometritic buffaloes Haryana Vet., 57(2): 226-228
25. Vale, W.O. Ohashi, J. Sousa and H. Biberis, 1988. Clinical reproductive problem of buffalo in latin America. 2nd World Buff. Cong. India.
26. Yessein, S., H. Shawki, M.M. Bashandy, S. Essawy and Ibtihal Abdallah, 1995. Clinicopathological studies in female infertile buffaloes. Buffalo J., 11: 83-89.
27. El-Hariri, M.N.E., M.A. Omar and M.R. Shalash, 1980. Some patterns of sexual disorders in the buffalo cow. Proceedings of 9th international Congress on Animals Reproduction and A.L. Madrid-Spain, IV: 754-758.
28. Sheldon, M., P.C.C. Molinari, Th.J.R. Ormsby and J.J. Bromfield, 2020. Preventing postpartum uterine disease in dairy cattle depends on avoiding, tolerating and resisting pathogenic bacteria. Theriogenology, doi: .2020.01.01716/j.theriogenology.2020.01.0172020;
29. Ismail, M., R. Soliman and M. Hatem, 1985. Correlation between the presence of pathogenic microorganisms and the level of lysozme in cervical mucus of repeat breeding buffaloes and cows. J. Egypt. Vet. Med. Ass., 45: 255-264.
30. Jubb, K.V.F., P.C. Kenedy and N. Palmer, 1985. Pathology of Domestic Animals. The female genital system. 3rd Ed. 3: 305-407. Acad. Press, Inc. Harcourt Brace Jovanovich, Publishers.
31. Zrally, Z., P. Kalab, J. Conderle, V. Kummer and J. Roszyk, 1989. Plasma progesterone, estrdiol, 11- hydroxyl corticsteoids concentration in cows with normal purperium and cow with puerperal disorder. Veterinaria Medicina, 34: 515-523.
32. Ezzo, O.H., W.M. Ahmed and A.R. Nada, 1993. Effect of some reproductive phases and disorders on plasma cortisol levels in buffaloes. Indian J. Anim. Sci., 63: 729-730.
33. Majeed, M.A., J. Iqbal and M.N. Chaudhry, 1990. Blood chemistry of clinical metritis in Nili-Ravi buffaloes of tow age groups and at two stages of lactation. Pakistan Vet. J., 10: 55-59.
34. Ashturkar, R.W., V.D. Aher and A.P. Bhokre, 1995. Studies on infertility problems in non-descript buffalo and cows. Indian Vet. J., 72: 1050-1052.
35. Salem, F.S., N.M. Moustafa, A. El-Taweel, R.H. Youssef and M.Z. Abdel-Aziz, 1994. Studies on some blood biochemical constituents in normal and abnormal cycling buffaloes. I. Peotein, lipids and transaminases. Proceedings, 4th World Buffalo Congress, Sao Paulo, Brazil, 27-30 June, 638-640.
36. Yessein, S., H. Shawki, M.M. Bashandy S. Essawy and Ibtihal Abdallah, 1995. Clinicopathological studies in female infertile buffaloes. Buffalo J., 11: 83-89.
37. Hahn, V., F. Zdelar, I. Harapin, A. Alegro, L. Bedrica, N. Dasorric and D. Viduc, 1989. Iron and copper content of the blood serum of cows with reproductive disorders. Veterinarski Glasnik, 43: 739-744.

38. Moustafa, N.M., F.S. Salem, R.H. Youssef, R.H. El-Taweel and A. Anwer, 1994. Studied on some blood biochemical constituents in normal and abnormal cycling buffaloes. II. Macro- & micro-elements. Proceeding 4th World Buffalo Cong. Sao Paulo, Brazil, 27-30 June, pp: 635-637.
39. El-Taweell, A., 1985. Comparative study of certain minerals in whole blood, correlation to infertility of buffalo cow. II Role of some micro-elements in reproduction. 1st World Buff.Cong.December, 27-31, Egypt, pp: 455-457.
40. Osman, M.N., 2016. Characterization of some innate immune genes associated with endometritis in river buffaloes. Thesis PhD degree – Beni-Suef university. Faculty of Veterinary Medicine.
42. Zheng, W.M., S.J. Lai and R.X. Shi, 1995. Serum albumin polymorphism of swamp buffalo in China. Chinese J. Anim. Sci., 31(1): 3-6.
43. Shalaby, S.I., W.M. Ahmed, M.M. Zaabal and H.A. Sabra, 1997. Studies on metabolic profile tests and plasma protein genotypes in normal cyclic buffalo-cows as affected by body condition scores. J. Agri. Sci. Mansoura Univ., 22(9): 2821-2832.
44. Elkhadrawy, H.H., M.M. Zaabal, W.M. Ahmed and S.I. Shalaby, 2010. Genetic constitution affecting ovarian activity in Egyptian cattle and buffaloes. Global J. Molecular Science, 5(2): 53-57.
45. Galvão, K.N., 2012. Association between immune function and development of uterine disease in dairy cows. Anim Reprod, 9(3): 318-322.
46. Ganesan, A., M. Sankar, R. Saravanan, G.K. Das, H. Kumar, A. Kumaresan and K. Narayanan, 2013. Mini-Review Article Endometrial Toll - like Receptors and Postpartum Uterine Infection, 1: 37–41.
47. Alfano, F., S. Peletto, M.G. Lucibelli, G. Borriello, G. Urciuolo, M.G. Maniaci, R. Desiato, M. Tarantino, A. Barone, P. Pasquali and P.L.. Acutis, 2014. Identification of single nucleotide polymorphisms in Toll-like receptor candidate genes associated with tuberculosis infection in water buffalo (*Bubalus bubalis*). BMC Genetics, 15(1): 139.