

Effects of Blends of Phytobiotic Additives of *Nigella sativa* L., *Trigonella foenum-graecum* L. and *Curcuma longa* on Serum Biochemical Profile of Broiler Chicks

¹Yonatan Kassu, ²Berhan Tamir, ³Etalem Tesfaye, ⁴Tesfaye Sisay and ⁵Yoseph Cherinet

¹Department of Animal and Range Science, Wolaita Sodo University, P.O. Box.138, Wolaita Sodo, Ethiopia

²Department of Animal Production Studies,

College of Veterinary Medicine, Addis Ababa University, P.O. Box: 34, Debre Zeit, Ethiopia

³Debre Zeit Agricultural Research Center, P.O. Box: 32, Debre Zeit, Ethiopia

⁴Institute of Biotechnology, Department of Health Biotechnology,
Addis Ababa University P.O. Box: 30604, Addis Ababa, Ethiopia

⁵Department of Biomedical Sciences, College of Veterinary Medicine,
Addis Ababa University, P.O. Box: 34, Debre Zeit, Ethiopia

Abstract: A study was conducted to compare the effectiveness of blends of different phytobiotic additives on biochemical indices. A total of 270-day-old unsexed broiler chicks (Cobb 500) were randomly allocated to six treatments with three replicates of 15 chicks each reared for 49 days. The experimental diets were: basal diet (T0), positive control basal diet+oxytetracycline at 0.035% (T1), basal diet + a blend of black cumin seed and fenugreek (T2), basal diet + a blend of black cumin seed and turmeric powder (T3), basal diet + a blend of fenugreek seed and turmeric powder (T4) and basal diet +a blend of black cumin seed + fenugreek seed + turmeric powder (T5). Serum total protein was significantly ($P<0.05$) higher in birds fed T3 diets as compared to T0 and T1 diet fed birds. While, birds fed in T5, T4 and T3 test diets had showed an enhanced ($P<0.05$) hypoglycemic and hypocholesterolemic effects than the birds in T1 and T0 diets. The value in the concentration of the liver enzymes Aspartate aminotransferase (AST) and Alanine aminotransferase (ALT) were not showed a significant difference ($P>0.05$) among treatments. However, Alkaline phosphatase (ALP) concentration was lowered ($P<0.05$) in birds fed diets of T5, T1, T4 and T2. Therefore, blends of phytobiotics showed improved hypoglycemic, hypocholesterolemic and hepatoprotective ability making them capable of substituting synthetic antibiotics and imparting positive effects on the health of broilers. Therefore, it is recommended that the three phytobiotics should be used in combined forms in order to utilize their beneficial biochemical links on serum biochemical profile of broiler chicks.

Key words: Antibiotics • Biochemical • Broiler • Blends • Phytobiotic

INTRODUCTION

Serum biochemical profiling has been used in several species of domestic livestock and birds to monitor health and to detect subclinical diseases. Biochemical status is a reflection of many factors such as sex, age, breed, diet, management and stress [1]. The routine uses of sub-therapeutic levels of antibiotics often referred to as antibiotic growth promoters (AGP) in poultry feed have been a common practice for more than 50 years to prevent potential diseases as well as to robust gut health, increase meat yield and improve feed efficiency [2]. The

aggravated utilization of antibiotics has led to the increased drug resistance of pathogens and the accumulation of antibiotic residues in animal products and the environment [3, 4]. This situation requires most countries in the world to restrict or limit the use of antibiotics in poultry feeds and launch "antibiotic-free" labeled feeds [5]. A practical approach to overcome this gap is the use of medicinal plants. Medicinal plants also known as phyto-genic/phytobiotic additives are often very complex mixtures composed of bioactive components, which include several groups of plant products depending on the origin and purpose of

production. The positive effects of medicinal plant or phytogene/ phytobiotic additives can be expected primarily through beneficial effects on the health of individuals by improving the immune system [6]. Some medicinal plants, such as black cumin, turmeric and fenugreek have been found to have an anti-inflammatory, antioxidant, anti-microbial, anti-protozoa, immune-modulatory effects, hypoglycemia, hypocholesterolemia, hepatoprotective, hepatostimulatory, analgesic, antipyretic, antivenin, antiulcer and anti-carcinogenic actions [7-10]. Fenugreek seed extracts have a unique feature of anti-hyperthyroidism and properties of anti-sterility and anti-androgenic effects [11, 12]. Even though phytobiotics provide a potential alternative to the antibiotic usage, more research on their synergetic or interaction effects is needed to understand the best form of utilization in poultry production, since the interaction of two or more agents to produce a combined effect greater than the sum of their individual effects [13]. Thus, the purpose of this study was to evaluate the effectiveness of *Nigella sativa* L, *Trigonella foenum-graecum* L. and *Curcuma longa* L. blend as phytobiotic additives on the biochemical indices of broiler chicken.

MATERIALS AND METHODS

Study Area: The experiment was conducted in the Poultry Farm of College of Veterinary Medicine Addis Ababa University, Bishoftu, Ethiopia. The area is situated at 47 km East of Addis Ababa at an altitude of 1900 m above sea level, latitude of 844' N and longitude of 38° 57' E. The average (25 years) annual rainfall is 851 mm with an average minimum and maximum temperature of 8.9°C and 26°C, respectively. The average relative humidity is 58.6% [14].

Experimental Ration and Treatments: Three medicinal plants, namely *Nigella sativa* L. (Black cumin), *Trigonella foenum graecum* L. (Fenugreek) and *Curicculum longa* (Turmeric) were purchased from the vicinity local markets to incorporate in the diets of broiler chicken as phytobiotic feed additives. The black cumin and fenugreek seed were washed with tap water and sun dried under shade. The dried turmeric rhizome, black cumin and fenugreek seeds were coarsely ground using manual mill and stored in polyethylene bag until required for the formulation of experimental rations.

Six treatment diets of broiler ration containing blends of different phytobiotic additives and control diet was formulated at a lower dosage levels (1%) set for each of

the medicinal plants. The treatments are T0 (negative control diet, which was only basal diet formulated from the feed ingredients of maize, wheat bran, soya bean meal and nuge seed cake), T1 (positive control, the basal diet + Oxytetracycline at 0.035%), T2 (basal diet + a blend of black cumin seed and fenugreek seed), T3 (basal diet + a blends of black cumin seed and turmeric powder), T4 (basal diet + a blend of fenugreek seed and turmeric powder) and T5 (basal diet + a blends of black cumin seed + fenugreek seed + turmeric powder) at equal proportion. The rations were formulated to be nearly isocaloric and isonitrogenous with metabolizable Energy (ME) content of 3000 kcal/kg DM and CP content of 22% during the starter phase of 1-28 days of age and ME content of 3200 kcal/kg DM and CP content of 20% during the finisher phase of 29- 49 days of age [15].

Management of Experimental Birds: A total of 270 unsexed day-old Cobb 500 broiler chicks with average initial body weight of 39.0.4±0.45 g were randomly and equally assigned to the six dietary treatments and with three replications per treatments. Before the commencement of the actual experiment, the experimental pens, watering and feeding trough cleaned and disinfected. The birds were vaccinated against Newcastle disease (Hitchner-B1 strain) at 7-days of age through intraocular and (Lasota strain) a booster dose at 22-days of age and Infectious Bursal Disease (Gumboro) at the age of 14 and 24 days and administered through spring drinking water. The chicks were brooded using 250 watt bulbs with gradual height adjustment as source of heat and light in a deep litter house covered with "teff" straw litter material. Feed was offered after weighed and clean tap water was available all the time throughout the experiment.

Measurements: The experiment was lasted for 7 weeks. Blood was collected at the end of the experimental period from two birds in each replication picked up randomly. About 2-3 ml blood samples were collected from the wing vein. The blood was transferred into non-heparinized tubes and later the coagulated blood samples were centrifuged for 15 minutes at 4000 rpm and the clear serum was separated and stored in a deep freezer at -20°C pending for biochemical analysis. Serum total protein and albumin were determined according to Doumas and Witt and Trendelenburg [16, 17]. Globulin concentration was calculated as the difference between total protein and albumin. Albumin/ Globulin g/100 ml (A/G ratio) was also calculated. Total cholesterol was determined according to Watson [18]. The estimation of alanine amino transferase

(ALT) and aspartate amino transferase (AST) followed the proposed optimized formulation of the International Federation of Clinical Chemistry (IFCC) [19], while measurement of alkaline phosphatase (ALP) was as per the German Society for Clinical Chemistry [20]. Sample analysis was conducted using the instrument HumaStar 80 automated chemistry analyzer (HUMAN Gesellschaft fuer Biochemical und Diagnostic GmbH, Germany).

Statistical Analysis: The collected data were subjected to one-way analysis of variance using the general linear models (GLM) procedures of SAS statistical package version 9.3 (SAS, 2010). Duncan's test was used to detect the differences among treatment means [21]. The model used was:

$$Y_i = \mu + \alpha_i + \varepsilon_i$$

where: Y_i = the dependent variables for the i th, μ = overall mean effect, α_i = the i th treatment effect, ε_i = the random error variation.

RESULTS

Serum Protein Metabolites: The analyses of biochemical profile due to feed supplementation of blends of medicinal plants of *Nigella sativa* L., *Trigonella foenum-graecum* L. and *Curcullum longa* L. as phytobiotics feed additives in broilers are illustrated in Table 1. The serum protein and globulin levels were significantly different ($P < 0.05$) among the treatments and was higher for phytobiotics additive blends of T3; of course the value was similar ($P > 0.05$) with T5 and T2. The serum globulin was also higher ($P < 0.05$) in T3 as compared to the rest of the treatments. However, there were no significant differences ($P > 0.05$) revealed on albumen and A/G ratio on birds fed the different treatment diets.

Serum Carbohydrate Metabolites: The influence of blends of phytobiotics feed supplementation on serum glucose and total cholesterol (Table 1) showed that there were significant differences ($P < 0.05$) among the treatments. The serum glucose and total cholesterol was significantly ($P < 0.05$) reduced in birds supplemented with blends of phytobiotic additives of T4 and T5 than the control T0 and T1. However, there was no significant difference ($P > 0.05$) observed among T0, T1 and T2 birds in serum glucose. Similarly, phytobiotic additives in T4 and T5 fed bird had lower ($P < 0.05$) serum total cholesterol as compared to the control T0 and T1.

Hepatotoxicity Indicators: The effect of supplementation of blended phytobiotic additives of *Nigella sativa* L., *Trigonella foenum-graecum* L. and *Curcullum longa* L. on liver enzyme hepatotoxicity (AST, ALT and ALP) is presented in Table 1. There were significant difference ($P < 0.05$) among the treatments on liver enzyme, ALP concentration. However, there were no significant differences ($P > 0.05$) were revealed on AST and ALT concentration in birds fed different blends of phytobiotics additives. While, the activity of AST observed in the range of 131.57-155.33 with no significant ($P > 0.05$) differences among treatments. While, the activity of ALT tended to decrease from T0 to T3, T1, T5 and T4, respectively. Serum ALP activities were reduced significantly ($P < 0.05$) by supplementing blends of phytobiotic feed additive diets of T1, T4 and T5 than the control diets. The minimum serum ALP activity was recorded from treatment T5 than T0 and T2 diet fed birds ($P < 0.05$).

DISCUSSION

Blends of phytobiotic feed additives effects on biochemical profile defined by black cumin + turmeric mixture as phytobiotic additives resulted in increased serum protein and globulin level than the antibiotics supplemented birds. The results were in agreement with El-Khalek and El-Naggar [22] who reported that addition of black cumin seeds, ginger, thyme and oregano oil mixtures were improved plasma total protein and globulin. Similarly, Yattoo *et al.* [23] also reported that the total protein was increased in the black cumin and fenugreek mixture supplemented birds. In inimitable study El-Bahr and Saad, [24] concluded that dietary supplementation of black cumin seed and turmeric mixture improved the serum protein of fish. As serum protein depends on the availability of dietary protein, these mean that the proteins of the black cumin + turmeric mixture diets were more available to the birds to support the nutritional requirements. The increase in total protein and globulin might be the synergetic effect of the active compounds in black cumin (thymoquinone and thymohydroquinone) and turmeric (curcuminoids and curcumin) which promote protein deposition, maintained a stable colloid osmotic pressure and improve the transportation of metabolic protein in birds [25]. In contrary to the current findings Toghyani *et al.* El-Bahr and Al-Azraqi and Fallah and Mirzaei [26, 27, 25] indicated that plasma total protein was not changed in broiler chicks fed black cumin seeds and turmeric powder.

Table 1: Serum biochemical profiles of broilers fed blends of black cumin, fenugreek seed and turmeric powder

Parameters	Treatments					SEM	P value	
	T0	T1	T2	T3	T4			T5
Total protein (g/dl)	2.50b	2.67b	3.33ab	4.50a	2.83b	3.5ab	0.19	0.02
Albumen (g/dl)	1.49	1.50	2.0	1.67	1.50	1.83	0.09	0.59
Globulin (g/dl)	1.0b	1.17b	1.33b	2.83a	1.33b	1.67b	0.17	0.02
A/G ratio	0.75	0.92	0.80	1.75	1.08	1.13	0.12	0.16
Glucose (mg/dl)	203.67a	180.0ab	185.0ab	161.67bc	135.33c	153.17bc	6.49	0.02
Cholesterol (mg/dl)	153.67a	151.67a	158.50a	136.17ab	114.33b	110.17b	12.37	0.03
AST (IU/L)	131.57	155.53	153.02	153.23	134.52	132.73	8.97	0.94
ALT (IU/L)	13.93	11.89	14.66	13.79	7.88	10.60	1.15	0.56
ALP (IU/L)	1451.77a	1014.13bc	1362.74ab	1078.35abc	1040.52bc	876.11c	57.88	0.02

*a-c Means in a row with different superscripts differ significantly ($P < 0.05$); T0= Control basal diet, T1= Antibiotics, T2= Black cumin + fenugreek, T3= Basal + Black cumin + Turmeric, T4= Fenugreek + Turmeric, T5= Black cumin + Fenugreek + Turmeric. A/G= Albumen to globulin ratio, AST= Aspartate aminotransferase, ALT= Alanine aminotransferase, ALP= Alkaline phosphatase

The blood glucose level was reduced in blend of fenugreek seed and turmeric powder better than the antibiotics supplemented birds. However, phytobiotic additives (black cumin + fenugreek + turmeric) mixtures were equivalent to the synthetic antibiotics (oxytetracycline). This is in agreement with Sameer *et al.* [28] who report that low doses of vanadate (0.2 mg/ml) in combination with fenugreek powder was found to be comparable and effective in correcting altered carbohydrate metabolism in rats to the synthetic vanadate (0.6 mg/ml) drug. Comparable to these findings, Al-Kassie *et al.* [29] reported that turmeric was effective in lowering blood glucose level than black cumin seed and their mixtures in broilers. Abou-Elkhair *et al.* [30] reported that supplementation of mixtures of (0.5% of black pepper and turmeric) and (0.5% black pepper, turmeric and 2% coriander seeds) had significantly lowered the blood glucose in broiler birds. Similarly to the present study, El-Bahr and Al-Azraqi [27] reported that the glucose level of diabetic rats (253.3%) has been reduced to 28.6% when rats have been treated with black cumin and turmeric mixture. In agreement with this study, El-Bahr and Saad [24] reported that black cumin + turmeric mixture (5gm/kg diet) were improved blood glucose and other biochemistry profile of Mugil cephalus fish.

The hypoglycemic effect of the phytobiotic mixtures especially fenugreek + turmeric might be due to the synergetic effects of fenugreek and turmeric active ingredients: trigonelline, the carbohydrate galactomannana, pectin from fenugreek and curcumin from turmeric and from both the amino acid (4-hydroxyisoleucine), which increases the gastric mucosal secretion and some flavonoids possess insulin-like properties and thereby can reduce the blood glucose level. The hypoglycemic effect of black cumin

seed mixtures perhaps explained by an insulin-like stimulation of glucose uptake by muscle and adipose tissue [31] or inhibition of intestinal glucose absorption [32].

The reduction of serum cholesterol by supplementation of different blends of phytobiotics (black cumin + fenugreek + turmeric) and (fenugreek + turmeric) were in agreement with El-Khalek and El-Naggar [22] who reported depression in the serum cholesterol level due to the supplementation of black cumin + fenugreek mixtures to the birds. In agreement with our findings, Al-Kassie *et al.* [29] recorded lowering in cholesterol level in the blends of turmeric and cumin at 0.75 and 1%. Black cumin and turmeric mixture restored the cholesterol to normal level in diabetic rats and fish [25- 28]. Similarly, Fallah *et al.* [33] who reported that addition of 1.5% artichoke leaves meal in diet with mentha extract 200 mg/kg in drinking water decrease cholesterol in broilers. On contrary, Abd El-Latif *et al.* [34] who reported rosemary and garlic at 100 and 200 mg/kg increase serum cholesterol in broiler chicken. Whereas, Narimani-Rad *et al.* [35] who reported that pennyroyal, ziziphora and peppermint mixture at 0.5 and 1% and Mohebbifar and Toriki [36] ground pits of palm dates with dried garlic and Myandoab and Hosseini-Mansoub [37] Liquorice root extract at 200 ppm with 1% probiotic (*Lactobacillus acidophilus* and *Lactobacillus casei*) was not have any considerable effect on glucose and total cholesterol levels.

The hypocholesterolemic effect of black cumin seeds may relate to the soluble fibers contents and sterols, especially β -sitosterol which decreases dietary cholesterol absorption and increases bile acid synthesis and degradation [38, 39]. The hypocholesterolemic effect of turmeric related to the altered activity of two effective

enzymes in cholesterol metabolism, HMG-CoA reductase (3-hydroxy 3-methylglutaryl Coenzyme A) and mediated by the stimulation of hepatic cholesterol 7 α -hydroxylase [40, 41]. Whereas, fenugreek might have contributed to the hypocholesterolemic effect by inhibiting the bile acid and cholesterol absorption from intestine, thereby, decreasing cholesterol level in blood [42, 43].

Results of liver function enzymes showed that no significant change in level of AST and ALT compared to the negative control and antibiotics supplemented birds. The current findings were in agreement with El-Khalek and El-Naggar [22] who reported that black cumin and other different medicinal plants mixture do not have toxic effect in liver as indicated AST and ALT activities. Similarly, Abd El-Latif et al. [34] who reported rosemary and garlic at 100 and 200 mg/kg had no significant difference on AST level in broiler chicken. The alkaline phosphate (ALP) activities were significantly lowered in birds supplemented with phytobiotic additives blends (black cumin + fenugreek + turmeric) and (fenugreek + turmeric), which were also comparable to the antibiotics drugs. Reductions of these enzymes are important as their accumulation in the liver are related to toxicity and hepatocellular damage. However, the level was within the standard range forwarded by Campbell [44] for the healthy chicken thus indicating the role of phytobiotic feed additives as hepatoprotective agents.

CONCLUSION

In conclusion, the improvement in some biochemical parameters in chickens fed blends of (black cumin + turmeric), (fenugreek + turmeric) as well as mixtures of the three phytobiotic additives (Black cumin + fenugreek + turmeric) revealed the ability of blends of phytobiotics additive to lower blood glucose and cholesterol levels as well as the hepatoprotective ability in broiler chicken; hence can substitute synthetic antibiotics and can impart positive effects on the health of broilers. The results revealed beneficial biochemical links between the three phytobiotics that justify their combined use. Therefore, it is recommended that the three phytobiotics should be used in combined forms in order to utilize their beneficial biochemical links on serum biochemical profile of broiler chicks.

ACKNOWLEDGMENT

The authors are grateful to Wolaita Sodo for their financial support. Addis Ababa University is particularly acknowledged for the financial support via the third round

Thematic Research Project. The authors are also highly indebted to all staff members of College of Veterinary Medicine, Addis Ababa University for their collaboration and facilitation.

Declaration of Interest: The authors declare that they have no conflict of interest.

REFERENCES

1. Piotrowska, A., K. Burlikowska and R. Szymeczko, 2011. Changes in blood chemistry in broiler chickens during the fattening period. *Folia Biologica.*, 59(3-4): 183-187.
2. Gaskins, H.R., C.T. Collier and D.B. Anderson, 2002. Antibiotics as growth promotants: mode of action. *Anim. Biotechnol.*, 13: 29-42.
3. Jallailudeen, R.L., M.J. Saleh, A.G. Yaqub, M.B. Amina, W. Yakaka and W. Muhammad, 2015. Antibiotic residues in edible poultry tissues and products in Nigeria: A potential public health hazard. *Int. J. Anim. Vet. Adv.*, 7: 55-61.
4. Toaha, S.M, B.R. Mollah and M.U. Ahammad, 2016. Use of dietary fenugreek (*Trigonella foenum-graecum* L.) seed for the production of safe broiler lean meat. *Res. Agric. Livest. Fish.*, 3(2): 305-314.
5. Cogliani, C., H. Goossens and C. Greko, 2011. Restricting antimicrobial use in food animals: lessons from Europe banning nonessential antibiotic uses in food animals is intended to reduce pools of resistance genes. *Microbe.*, 6: 274-279.
6. Windisch, W., K. Schedle, C. Plitzner and A. Kroismayr, 2008. Use of phytogetic products as feed additives for swine and poultry. *J. Anim. Sci.*, 86: 140-148.
7. Gilani, A.H., Q. Jabeen and M.A.U. Khan, 2004. A Review of medicinal uses and pharmacological activities of *Nigella Sativa*. *Pakistan J. Biol. Sci.*, 7: 441-451.
8. Raskin, I., D.M. Ribnicky, S. Komarnytsky, N. Llic, A. Poulev, N. Borisjuk, A. Brinker, Moreno, D.A. Ripoll, C. Yakoby, N. O'Neal, J.M., Cornwell, T. Pastor and B. Fridlender, 2002. Plants and human health in twenty-first century. *Tren. Biotech.*, 20 (12): 522-531.
9. Jayaprakasha, G.K., B.S. Jena, P.S. Negi and K.K. Sakariah, 2002. Evaluation of antioxidant activities and anti-mutagenicity of turmeric oil: A byproduct from curcumin production. *Z. Natu. Forsch.*, 57: 828-35.

10. Kavirasan, S., G.H. Naik, R. Gangabhairathi, C.V. Anuradha and K.I. Priyadarsini, 2007. *In-vitro* studies on antiradical and antioxidant activities of fenugreek (*Trigonella foenum-graecum*) seeds. Food Chem., 103: 31-37.
11. Kamal, R., R. Yadav and J.D. Sharma, 1993. Efficiency of the steroidal fraction of the fenugreek seed extract on the fertility of male albino rats. Phytother. Res., 7(2):134-138.
12. Tahiliani, P. and A. Kar, 2003. The combined effects of *Trigonella* and *Allium* extracts in the regulation of hyperthyroidism in rats. Phytomedicine, 10(8): 665-668.
13. Van Vuuren, S. and A. Viljoen, 2011. Plant-based antimicrobial studies-methods and approaches to study the interaction between natural products. Planta. Med., 77: 1168.
14. DZARC (Debre Zeit Agricultural Research Center). 2003. Annual Research Report 2002/03, Ethiopian Institute of Agricultural Research, Debre Zeit, Ethiopia.
15. Leeson, S. and J.D. Summers, 2005. Commercial Poultry Nutrition. 3rd Ed, Nottingham University Press, England, pp: 398.
16. Dumas, B., 1971. Colorimetric determination of serum albumin. Clin. Chem. Acta., 31: 400-403.
17. Witt, L. and C. Trendelenburg, 1982. A method for rapid determination of total protein of serum, J. P. Clin. Biochem., pp: 220-235.
18. Watson, D., 1960. A simple method for determination of serum cholesterol. Clin. Chem. Acta., 5: 637-642.
19. Bergmeyer, H.V., M. Horder and R. Rej, 1986. Approved recommendation on IFCC methods for the measurement of catalytic concentration of enzymes. Part II. IFCC method for Aspartate Amino Transferase, J. Clin. Chem. Clin. Biochem., 24: 497.
20. GSCC (German Society for Clinical Chemistry), 1972. Recommendations of the Enzyme Commission. Z. Klin. Chem. Klin. Biochem., 10: 281.
21. Duncan, D.B., 1955. Multiple range and multiple F tests. Biometrics, 11: 1-42.
22. El-Khalek, E.A. and A.S. EL-Naggar, 2016. Growth and physiological response of gimmizah chicks to dietary supplementation with ginger, black seeds, thyme and oregano oil as natural feed additives. Egypt. Poult. Sci., 36(4): 1169-1182.
23. Yattoo, M.A., R.K. Sharma and N. Khan, 2012. Effect of fenugreek and black cumin seeds as feed additives on blood biochemical profile and performance of broilers. Indian J. Anim. Nutri., 29(2): 174-178.
24. El-Bahr, S.M. and T.T. Saad, 2008. Effect of Black cumin seeds (*Nigella sativa*) and/or Turmeric (Curcumin) on hematological, biochemical and immunological parameters of Mugil cephalus fish vaccinated with *Aeromonas hydrophila* bacterin, pp: 365-388. The 13th scientific congress, 25-28, November, Faculty of veterinary medicine, Assuit University.
25. Fallah, R. and E. Mirzaei, 2016. Effect of dietary inclusion of turmeric and thyme powders on performance, blood parameters and immune system of broiler chickens. J. Livest. Sci., 7: 180-186.
26. Toghyani, M., M. Toghyani, A. Gheisari, G. Ghalamkari and M. Mohammadrezaei, 2010. Growth performance, serum biochemistry and blood hematology of broiler chicks fed different levels of black seed (*Nigella sativa*) and peppermint (*Mentha piperita*). Livest. Sci., 129: 173-178.
27. El-Bahr, S.M. and A.A. Al-Azraqi, 2014. Effects of dietary supplementation of turmeric (*Curcuma longa*) and black cumin seed (*Nigella sativa*) in streptozotocin induced diabetic rats. Int. J. Bioche. Res. Rev. 4(46): 481-492.
28. Sameer, M., T. Asia, R.N.K. Bamezaia, F. Seemi, N. Basirb and B. Zaheer, 2003. Lower doses of vanadate in combination with trigonella restore altered carbohydrate metabolism and antioxidant status in alloxan-diabetic rats. Clinica. Chimica. Acta., 342: 105-114.
29. Al-Kassie, G.A.M., A.M. Mohseen and R.A. Abd-Al-Jaleel, 2011. Modification of productive performance and physiological aspects of broilers on the addition of a mixture of cumin and turmeric to the diet. Res. Opin. Anim. Vet. Sci., 1(1): 31-34.
30. Abou-Elkhair, R., H.A. Ahmed and S. Selim, 2014. Effects of black pepper (*Piper nigrum*), turmeric powder (*Curcuma longa*) and coriander seeds (*Coriandrum sativum*) and their combinations as feed additives on growth performance, carcass traits, some blood parameters and humoral immune response of broiler chicken. Asian-Austr. J. Anim. Sci., 27(6): 847-854.
31. Benhaddou-Andaloussi, A., L.C. Martineau, D. Vallerand, Y. Haddad, A. Afshar, A. Settaf, P.S. Haddad, 2010. Multiple molecular targets underlie the antidiabetic effect of *Nigella sativa* seed extract in skeletal muscle, adipocyte and liver cells. Diabet. Obesi. Metabol., 12: 148-157.

32. Meddah, B., R. Ducroc, M. Faouzi, B. Eto, L. Mahraoui, A. Benhaddou-Andaloussi, L.C. Martineau, Y. Cherrah and P.S. Haddad, 2009. *Nigella sativa* inhibits intestinal glucose absorption and improves glucose tolerance in rats. J. Ethnopharmacol., 121: 419-424.
33. Fallah R., A. Kiani and A. Azarfar, 2013. Effect of artichoke leaves meal and mentha extract (*Mentha piperita*) on immune cells and blood biochemical parameters of broilers. Global Veterinaria, 10(1): 99-102.
34. Abd El-Latif, A.S., N.S. Saleh, T.S. Allam and E.W. Ghazy, 2013. The effects of rosemary (*Rosemarinus officinalis*) and garlic (*Allium sativum*) essential oils on performance, hematological, biochemical and immunological parameters of broiler chickens. British J. Poult. Sci., 2(2): 16-24.
35. Narimani-Rad, M., A. Nobakht, H.A. Shahryar and A. Lotf, 2011. Influence of dietary supplementation of medicinal plants mixture (ziziphora, menta pulagum and peppermint) on some serum biochemical and immunological measures in broiler chickens. Middle-East J. Scientific Res., 8(2): 457-459.
36. Mohebbifar, A. and M. Toriki, 2011. Growth performance and humoral immune response of broiler chicks fed diets containing graded levels of ground date pits with a mixture of dried garlic and thyme. Global Veterinaria, 6(4): 389-398.
37. Myandoab, M.P. and N. Hosseini-Mansoub, 2012. Comparative effect of Liquorice root extract medicinal plants and probiotic in diets on performance, carcass traits and serum composition of Japanese quails. Global Veterinaria, 8(1): 39-42.
38. Ali, B.H. and G. Blunden, 2003. Pharmacological and toxicological properties of *Nigella sativa*. Phytother. Res., 21: 299-305.
39. Ali, O.A.A., N. Suthama and L.D. Mahfud, 2014. The effect of feeding black cumin (*Nigella sativa*) and vitamin C on blood lipid profiles and growth performance of broilers. Int. Refe. J. Engin. Sci., 3(4): 28-33.
40. Kim, M. and Y. Kim, 2010. Hypocholesterolemic effects of curcumin via up-regulation of cholesterol 7 α -hydroxylase in rats fed a high fat diet. Nutr. Rese. Prac., 4: 191-195.
41. Daneshyar, M., M. Ghandkanlo, F. Bayeghra, F. Farhangpajhoh and M. Aghaei, 2011. Effects of dietary turmeric supplementation on plasma lipoproteins, meat quality and fatty acid composition in broilers. Agri. Conspe. Scienti., 41(4): 420-428.
42. Meghwal, M. and T.K. Goswami, 2012. A review on the functional properties, nutritional content, medicinal utilization and potential application of Fenugreek. J. Food. Proc. Techol., 3(9): 1-10.
43. Mukhtar, M.A., K.A. Mohamed, O.A. Amal and H. Ahlam, 2013. Response of broiler chicks to different dietary levels of spearmint oil (SPO) as a natural growth promoter. Univ. Bakht Alruda Scienti. J., 6: 175-183.
44. Campbell, T.W., 2012. Clinical chemistry of birds. In: M.A. Thrall, G. Weiser., R.W. Allison. and T.W. Campbell (2nd Eds). Veterinary hematology and clinical chemistry, pp: 582-598. Wiley-Blackwell, A John Wiley & Sons, Inc., Publication. Iowa, USA.