

Diversity and Distribution of Indole Acetic Acid Producing Marine Sponge Associated Bacteria from Gulf of Mannar, Southeast Coast of India

V. Vasanthabharathi and S. Jayalakshmi

Faculty of Marine Sciences, Annamalai University, Tamil Nadu, India

Abstract: Many microbes promote plant growth and many microbial products that stimulate plant growth have been marketed. In coastal agriculture system most of the IAA producing terrestrial or plant symbiotic microbes were used as growth stimulating agent. Those microbes were not withstand high salinity, nutrients etc., so that present study focused on this study. Sponge samples were collected from Gulf of Mannar, Southeast coast of India and identified. About four sponges were collected namely *Callyspongia diffusa*, *Hyattella Cribriiformis*, *Sigmatocia carnosus*, *Spongia officinalis var. ceylonensis*. Sponge associated bacteria were isolated and identified. Then screened for Indole acetic acid production. Among the tested strains maximum indole acetic acid production was observed with *Callyspongia diffusa* associated *P. fluorescens*.

Key words: IAA • Sponges • Gulf of Mannar • Bacteria

INTRODUCTION

Indole-3 acetic acid (IAA) is a phyto hormone which is essential for plant growth and development. The ability to produce the plant hormone indole-3-acetic acid (IAA) is widespread among fungi and bacteria. Some microorganisms which inhabit the aerial or surfaces of plants are capable of IAA synthesis. Such microbes include rhizobia [1].

It has been reported that in some sponge species as much as 40% of animal biomass is attributed to bacteria, which exceeds the bacterial population of seawater by 2 orders of magnitude [2].

There are earlier works reported that marine bacteria have the ability to produce auxins IAA[3], The production of auxins by marine bacteria may offer a chance to use these bacteria as biofertilizers to improve the growth and yield of agricultural crops in coastal saline influenced lands in coastal agricultural system. Strains from marine environment which produce IAA may be used for the production of marine agro friendly biofertilizer production.

There are many studies regarding the synthesis of IAA by many terrestrial bacteria. IAA production by freshwater wetland rhizosphere bacteria [4] but, reports on auxin production by marine sponge associated bacterial strains is very scanty. Hence this study was carried out.

MATERIALS AND METHODS

Collection and Identification of Sponges: Sponges were collected from the Gulf of Mannar, Southeast coast of India (Lat 9°5' N; Long 79°5' E). The sponge sample soon after collection was transferred to a sterile polyethylene bag and transported under frozen condition to the laboratory. It was identified by through microscopic and macroscopic analyses.

Isolation of Bacteria Associated with Marine Sponges: The sponge samples soon after collection was transferred to a sterile polyethylene bag and transported at 4°C to the laboratory for the isolation of associated microbes. On reaching the laboratory, the invertebrate was brought to room temperature and cut aseptically into small pieces (2 × 2 cm) using a sterile scissors. The pieces were freed from adhering particles by vortexing twice for 20 sec. with 2 ml of sterile seawater. The seawater was decanted, which was once again replaced with sterile seawater with continued vortexing between washings. Finally, sample in sterile seawater was homogenized using sterilized mortar and pestle in a Laminar flow chamber. The homogenate was serially diluted up to 10⁻⁶ dilutions and then spread plated on Zobell marine agar plates. The plates were incubated at room temperature for 24-48 hrs. Based on the colony morphology strains were collected and stored [5].

Identification of Sponge Associated Bacteria: All associated bacterial strains which were selected based on morphology were identified biochemically. Morphological characters identified up to the species level by following Bergey's manual of determinative bacteriology [6].

Screening for Indole Acetic Acid (IAA) Producing Potential Strain: Screening was done by using Luria broth supplemented with L-tryptophan. A loop full of strains were inoculated in 10 ml of Luria broth supplemented with L-tryptophan containing tubes and incubated for 72 hrs at 30°C. Then, the cultures were centrifuged at 10,000 x g for 10 min. and the supernatant was collected.

One ml of supernatant was allowed to react with 2 ml of Salkowsky reagent (1 ml of 0.5 M FeCl₃ in 50 ml of 35% HClO₄) at 30°C for 30 min., Pink colour development indicated the presence of IAA [7].

Estimation of IAA: The cultures were centrifuged at 13000 x g for 10 min. and the supernatant was collected. The presence of IAA was measured in a spectrophotometer by adding 2 ml of Salkowsky reagent to 1 ml of supernatant, incubated for 30 min. The optical density was read at 530 nm. The recorded OD values were plotted in a standard curve prepared from commercially available IAA (8) and their concentration was calculated.

RESULTS AND DISCUSSIONS

Density of Bacteria Associated with Sponges: The sponges viz., *Callyspongia diffusa*, *Hyattella Cribriiformis*, *Sigmatocia carnosia*, *Spongia officinalis var. ceylonensis* were analysed for associated bacterial population. In *Callyspongia diffusa* bacterial density was in the range of 7.68x10³ CFU/g to 1.1 x 10⁷ CFU/g, whereas in the other three species i.e., *Hyattella Cribriiformis*, *Sigmatocia carnosia*, *Spongia officinalis var. ceylonensis* respectively 3.13 x10³CFU/g to 1.6x 10⁷CFU/g, 6.77 x10³CFU/g to 1.5 x 10⁷CFU/g, 2.69 x10³CFU/g to 1.4 x 10⁷CFU/g were the bacterial density found. Likewise, about 10 marine bacterial strains were isolated from the marine sponge *Callyspongia diffusa* by Kalirajan *et al.* [9]. Krishnan *et al.*[10] got 6.5 x 10⁶cfu/ gm⁻² total heterotrophic bacteria associated with the sponge, *Stylissa* sp. Boobathy *et al.* [11] extracted the bioactive protein from *P.aeruginosa*, *E.coli*, *V.cholera*, *V.heamolyticus* which was associated with *Callyspongia diffusa*.

Ravikumar *et al.* [12] isolated Cynobacterial symbionts from the sponge *Sigmatocia carnosia*.

In the present study most of the bacterial strains which were associated with sponges were indole acetic acid producers. *Callyspongia diffusa* associated bacterial strains were produced indole acetic acid. Among the strains *P.fluorescens* (0.41 µg/ml) followed by *A. hydrophila* (0.22 µg/ml), *P. putida* (0.21 µg/ml), *B. subtilis* (0.13 µg/ml) *B.licheniformis* (0.12 µg/ml), *A.faecalis*(0.11 µg/ml), *P. aeruginosa* (0.11 µg/ml) (Fig.1). In *Hyattella Cribriiformis* associated bacteria maximum IAA produced with *B. subtilis* (0.26 µg/ml) followed by *P. putida* (0.21 µg/ml), *B.megaterium*(0.2 µg/ml), *C.kutcheri*(0.13 µg/ml) *B. cereus*, *L. plantarum* (0.1 µg/ml) (Fig.2).

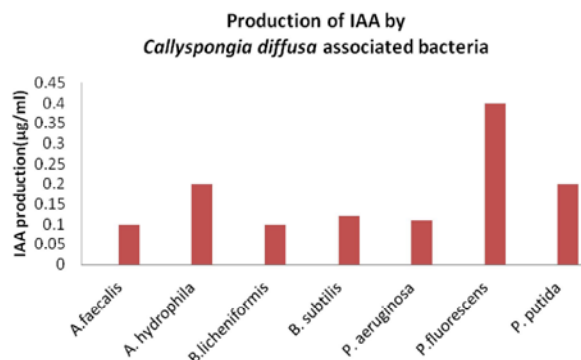


Fig. 1: IAA by *Callyspongia diffusa* associated bacteria

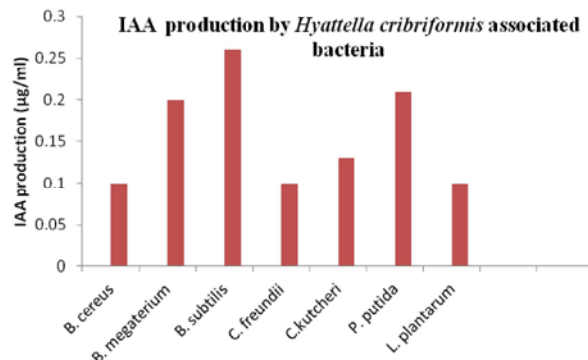


Fig. 2: IAA by *Hyattella cribriiformis* associated bacteria

In *Sigmatocia carnosia* maximum IAA production was observed with *P. putida*(0.28 µg/ml), *B. pumilis* (0.24 µg/ml), *B. megaterium*, *S. marcescens* (0.23 µg/ml), *B. cereus*, *P.aeruginosa*(0.21 µg/ml), *L. divergens* (0.14 µg/ml), *B.brivis*(0.12 µg/ml). (Fig.3).Maximum IAA production was observed with *B. subtilis* (0.3 µg/ml) *B. megaterium*, *C.glutamicum*(0.28 µg/ml), *E.coli* (0.27 µg/ml) minimum was observed with (*B. brevis* 0.21 µg/ml) in *Spongia officinalis var. ceylonensis* associated bacteria (Fig.4).

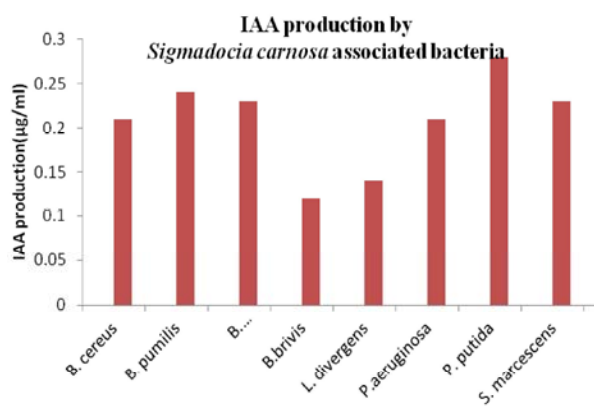


Fig. 3: IAA production by *Sigmoidocia carnosa* associated bacteria

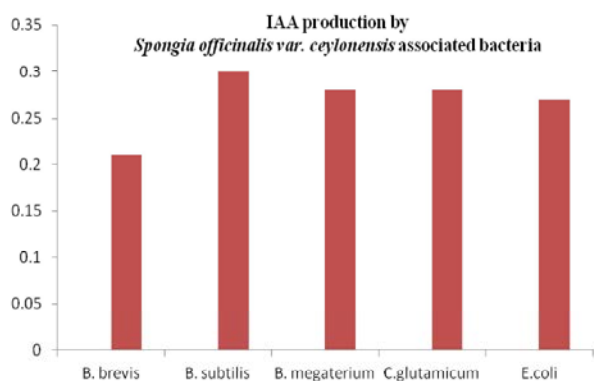


Fig. 4: IAA production by *Spongia officinalis var. ceylonensis* associated bacteria

Among the associated bacteria *P. fluorescens* from *Callyspongia diffusa* is a novel Iaa producer. Sergeeva *et al.* [13] stated that 83% of the symbiotic isolates tested positive production of auxin-like compounds compared to 38% of the free-living ones, Prabha Devi *et al.* [14] collected the sponge *Halichondria* sp., from the Gujarat coast of the Indo Pacific region and they isolated the *Bacillus* sp. SAB1, extracted indole (1), 3-phenylpropionic acid.

Jayaprakashvel *et al.* [15] studied the IAA production from Halotolerant bacteria associated with rhizospheres of *Sueveda* sp. along the Kelambakkam salterns, Tamil Nadu. The plant growth-promoting rhizobacterial strain *Pseudomonas putida* Rs-198 was isolated from salinized soils from Xinjiang Province produce the IAA [16].

The present study thus proved the potential of the marine sponge associated bacteria has potent to produce indole acetic acid. In future it can be specially used in coastal agricultural system.

REFERENCES

1. Jones, J.B., R.E. Summons, B.G. Entsch, C.W. Rolfe, P. Letham and D.S. Letham, 1982. Mass spectrometric identification of indole compounds produced by *Rhizobium* strains. *Phytochem.*, 9: 2029-2036.
2. Friedrich, A.B., I. Fischer, P. Proksch, Hacker and U. Hentschel, 2001. Temporal variation of the microbial community associated with the Mediterranean sponge *Aplysina aerophoba*. *FEMS Microbiol. Ecol.*, 38: 105-113.
3. Jayaprakashvel, M., P. Thiruchitrabalam, R. Muthezhilan, C. Kaarthikeyan, K. Karthik and A. Jaffar Hussain, 2011. (Plant growth promotion potential of halotolerant bacteria isolated from Kelambakkam Salterns, Tamil Nadu). In: National conference on Marine Explorations of the Natural Bioactive Compounds from the Marine Resources" during Feb 10 and 11th 2011 organized by Department of Marine Science, Bharathidasan University.
4. Haida-Alija, L., 2003. (Identification of indole-3-acetic acid producing freshwater wetland rhizosphere bacteria associated with *Juncus effuses*) L. *Can. J. Microbiol.*, 49: 781-787.
5. Vasanthabharathi, V. and S. Jayalakshmi, 2012. Bioactive potential of symbiotic bacteria and fungi from marine sponges. *Af. J. Biotechnol.*, 11(29): 7500-7511.
6. Buchanan, R.E. and N.E. Gibbons, 1974. In: *Bergey's Manual of Bacteriology*. 8th edition. (Eds) Williams and Wilkins, Baltimore, U.S.A. pp: 93-110.
7. Patten, C.L. and B.R. Glick, 1996. Bacterial biosynthesis of indole-3-acetic acid. *Can. J. Microbiol.*, 42: 207-220.
8. Khan, Z. and S.L. Doty, 2009. Characterization of bacterial endophytes of sweet potato plants. *Plant Soil*, 6: 104-107.
9. Kalirajan, A., A. Ranjitsingh and J. Appadorai, 2013. Antioxidant potential and biochemical evaluation of metabolites from the marine bacteria *Virgibacillus* sp. associated with the sponge *Callyspongia diffusa*. *Sci. Biol. Med. Org. Phcog. Net.*, 3: 47-51.
10. Krishnan, P., M. Balasubramaniam, S.D. Roy, K. Sarma, R. Hairun and J. Sunder, 2014. Characterization of the antibacterial activity of bacteria associated with *stylissa* sp, a marine sponge. *Adv. Anim. Vet. Sci.*, 2(1): 20-25.

11. Boobathy, S., T.T. Ajithkumar and K. Kathiresan, 2009. Isolation of symbiotic bacteria and bioactive proteins from the marine sponge, *Callyspongia diffusa*. Ind. J. Biotechnol., 8: 272-275.
12. Ravikumar, S., A. Nural shiefa and A. Kalaiarasi, 2011. Bio inoculation of *Sigmoidocia carnosa* sponge on the growth of sponge associated symbionts. Adv. App. Scie. Rese., 2(2): 422-427.
13. Sergeeva, E., A. Liaimer and B. Bergman, 2002 Evidence for production of the phytohormone indole-3-acetic acid by cyanobacteria. Planta, 215: 229-238.
14. Prabha Devi, S. Wahidullah, C. Rodrigues and Lisette D. Souza, 2010 The Sponge-associated Bacterium *Bacillus licheniformis* SAB1: A Source of Antimicrobial Compounds. Mar. Drugs., 8: 1203-1212.
15. Jayaprakashvel., M., K. Abishamala, C. Mathan Periasamy, J. Satheesh, A. Jaffar Hussain and M.C. Vanith, 2014. Isolation and Characterization of Indole Acetic Acid (IAA) Produced by a Halo Tolerant Marine Bacterium Isolated from Coastal Sand Dune Plants Bio. Biotechnol. Res. Asia, 11(1): 263-269.
16. Peng, Y., H.E. Yanhui, W.U. Zhansheng, L.U. Jianjiang and Chun Li, 2014 Screening and optimization of low-cost medium for *Pseudomonas putida* Rs-198 culture using RSM. Braz. J. Microbiol., 45(4): 1229-1237.