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Molecular Characterization of *Echinococcus granulosus* Cysts Isolated from Some Animals in Egypt

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Abstract: Hydatidosis is one of the most important parasitic zoonoses and remains a public health and economic problem all over the world. The hydatid cysts (HC) were collected from freshly slaughtered 270 (180 camels, 90 cattle) at Cairo abattoir and from 120 inspected donkeys at Giza zoo, Egypt. The incidence of infection of HC was 18.9%, 3.3% and 14.2% among examined camels, cattle and donkeys respectively, 5.7%, 66.7% and 90.2% had in HC liver respectively, while the infection rate was 94.3%, 33.3% and 9.8% in the lung of examined camels, cattle and donkeys respectively. The rate of fertile cysts was found to be 42 (79.24%) in camel and 15 (29.4%) in donkeys. while, all cysts collected from the inspected cattle were found calcified. Germinal memberanes were used for DNA extraction followed by PCR amplification. The PCR yielded similar amplified DNA band of the same molecular size marker at 1115 bp in different isolates of Hydatid. No band variation of ITS 1 gene could be detected by PCR-RFLP by using two restriction enzymes. Amplification product of ITSI after digestion with MSP1 showed at 661 bp, while those restricted with RSA1 enzyme appeared at 745 bp.

Key words: Hydatid cyst · Molecular · Antigens · PCR · Camel · Cattle · Donkey · Egypt

INTRODUCTION

Hydatidosis is a global animal and human health problem of increasing economic and public health importance [1]. It is a helminthic cyclo-zoonosic disease caused by the larval stage (Metacestode) of the canid tapeworm *Echinococcus* that require at least one other vertebrate host for completion of its life cycle. The disease is endemic in many parts of the world [2]. It is one of the major zoonotic parasitic diseases in the Middle East and Arab North Africa from Morocco to Egypt [3]. The disease has great public health importance and economic impact in countries where livestock industry is an important segment of the agricultural sector and when livestock production is based mainly on extensive grazing system [4].

Regarding its molecular characterization *E. granulosus* poses a high degree of genetic diversity based on genome pattern, morphology and host specificity have allowed the differentiation of at

least Ten different genotypes (G1-G10) among which G4 (Horse strain) have been formerly characterized [5-8] and Camel strain (G6) have been formally in Eastern Africa [9] North Africa [10] and Tunisia [11].

The present study was conducted on slaughtered camel, cattle and scarified donkeys aiming to determine the incidence of hydatid infection. PCR was used for amplification of DNA extracted from fertile HC for identification of ITS1 gene of camel and donkeys followed by further identification by PCR-RFLP using two digestive enzymes *MSP1* and *RSA1*.

MATERIALS AND METHODS

This study was assessed and approved by Faculty of Veterinary Medicine, Cairo University Ethics Committee and therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki.

Corresponding Author: O.A. Mahdy, Department of Parasitology, Faculty of Veterinary Medicine, Cairo Univ. El-Giza, Egypt. **Sampling:** The HC were collected from freshly slaughter animals 270 (180 camels, 90 cattle) at Cairo abattoirs and from inspected 120 donkeys slaughtered at the zoo of Giza Zoo, Egypt. Intact HC, isolated from the infected animals, were put separately in the polythene bags containing ice and brought to Veterinary Medicine of Cairo University for further processing. Examination of all internal organs was also done by using palpation and incision for the detection of HC according to the technique recommended by Gracy [12].

Microscopic Identification of HC: The suspected infected organs were collected from slaughtered and scarified animals for routine microscopic examination according to [13]. Cyst fluid was obtained from pulmonary and hepatic cysts for demonstration of protoscolices and hooklets. Protoscolices were isolated from the fertile cysts and then washed three times by phosphate buffer saline (PBS), pH 7.2 and preserved in 70% alcohol (v/v) for isolation of DNA [14].

DNA Extraction: DNA was extracted from germinal layer of fertile HC using Genei Ultrapure TM Mammalian Genomic DNA Purification Tissue Kit (Bangalore Genei). According to manufacturer's protocols. The germinal layer of fertile HC DNA extrats were stored at-20 °C until used in molecular characterization work.

Polymerase Chain Reaction (PCR) Assay: Amplification of ITS1 gene was done by using of primers described by Bowles and McManus [15]. The primer was designed as forward 5' GTC GTA ACA AGG TTT CCG TA'3 and reverse 5'TCT AGA TGC GTT CGA A (G/A) TGT CGA TG'3, (Jena, Bioscience, Germany). A100-bp DNA was used as molecular size marker. The amplification

Table 1: Incidence of HC infection in examined animal

reaction was carried in 25ìl volume containing 500mM Kcl, 10 mM Tris-Hcl (PH9.0), 1% Triton x-100, 4 mM Mgcl, 100uM dNTPs each, 15-20ng of ITS1 primer, 25ng of DNA and 1.5 units of Tag DNA polymerase. For data analysis PCR assay was performed in thermal cycler (Teche TC-512UK); Initial denaturation at 95°C for 6 min., 30 cycles of 94°C for 45 sec., 55°C for 60 sec. and 72°C for 90 second. Then followed by final extension at 72°C for 1 min.

PCR-Mediated RFLP: PCR product were digested with MSP1 and RSA1 (10u) using buffer recommended by the manufacture (Jena Bioscience, Germany). Restriction fragments were separated by gel electrophoresis through 2%TBE agarose gel. PCR products were analyzed after electrophoresis in 1.5% (W/v) agarose gel and visualized in ethidium bromide (15).

RESULTS

The data demonstrated in Table 1 cleared that the total incidence of infection by HC was 18.9%, 3.3% and 14.2% among examined camels, cattle and donkeys respectively. The cyst was diagnosed in 5.7 %, 66.7% and 90.2% of the examined liver respectively, while, it was 94.3%, 33.3% and 9.8% in the examined lung of camels, cattle and donkeys respectively, Table 2. Moreover, the rate of fertile cysts was found to be 42 (79.24%) in camel and 15(29.4%) in donkeys. while, all Microscopically, the higher incidence of viable motile protoscoleces (60.4%) was found in HC of camel origin, then (23.5%) in HC of donkey origin. The highest incidence of non motile protoscoleces (18.9%) were found in that of camel origin while it was 5.9% in that of donkey origin, Table 4.

Animals	No. Ex.	No. Inf.	%	No. cysts	Mean and average No. Cyst/ animals
Camels	180	34	18.9	53	(1-3)1.55
Cattle	90	3	3.3	6	(1-2)1
Donkeys	120	17	14.2	51	(2-3)3
Total	390	54	13.8	110	

Table 2: Incidence of HC distributions in different site of infection in the examined animals

		Site of infection	Site of infection					
		Lung		Liver				
Infected animals	No. cysts	 No.	%	 No.	%			
Camels	53	50	94.3	3	5.7			
Cattle	6	2	33.3	4	66.7			
Cattle Donkeys	51	5	9.8	46	90.2			
Total	110	57	51.8	53	48.2			

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Animals	No. Ex.	No Inf.	%	No. Cyst	Sterile HC		Fertile HC		Calcified HC	
					No	%	No.	%	No.	%
Camels	180	34	18.9	53	3	5.7	42	79.24	8	15.1
Cattle	90	3	3.3	6	0	0	0	0	6	100
Donkey	120	17	14.2	51	17	33.3	15	29.4	19	37.3
Total	390	54	13.8	110	20	18.2	57	51.8	33	30.0

Table 3: Incidence of sterile and fertile HC in the examined animals

Table 4: Incidence of HC according to viability of Protoscolces

Animals	No. Cyst	No. fertile cyst No.	Fertile cysts with				
			Motile Protosco	blices	Non-motile Protoscolices		
			 No.	%	 No.	%	
Camels	53	42	32	60.4	10	18.9	
Cattle	6	0	0	0	0	0	
Donkeys	51	15	12	23.5	3	5.9	
Total	110	57	44	40	13	11.9	

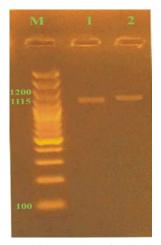


Fig. 1: Agarose gel electrophoresis of the PCR-derived amplicons of of HC germinal layer of *E. granulosus* ITS1 gene, separated on 2% agarose gel and stained with ethidium bromide Lanes: (M) 100 bp DNA ladder (consists of repeats of 100 bp fragment size, Fermintas), lane 1Camel DNA and 2 Donkey DNA containing 1115 bp.

Identification of the genetic characters of hydatid cysts obtained from camel and donkeys after PCR amplification of ITS1 gene showed similar pattern of PCR product, all amplified DNA products have band of the same molecular size at 1115bp on agarose gel (Fig.1). Further more molecular analysis using PCR-RFLP for amplification to the product of ITS1 after digestion with B. Agarose gel electrophoresis of the PCR-derived amplicons of of HC germinal layer of *E. granulosus* ITS1

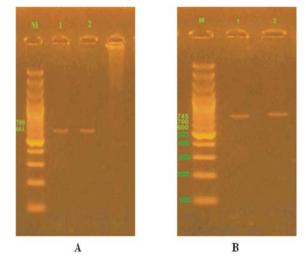


Fig. 2: A Agarose gel electrophoresis of the PCR-derived amplicons of of HC germinal layer of *E.* granulosus ITS1 gene after digestion with MSP1 separated on 2% agarose gel and stained with ethidium bromide Lanes: (M)100 bp DNA ladder(consists of repeats of 100 bp fragment size, Fermintas), lane 1Camel DNA and 2 Donkey DNA containing 661 bp.

gene after digestion with RSP1 separated on 2% agarose gel and stained with ethidium bromide Lanes: (M) 100 bp DNA ladder (consists of repeats of 100 bp fragment size, Fermintas), lane 1Camel DNA and 2 Donkey DNA containing 745 bp. MSP1 showed that all strainssharing in the MW at 661 bp (Fig. 2A), while those restricted with RSA1 enzyme appeared at 745 bp (Fig. 2B).

DISCUSSION

Echinococcus granulosus is medically and economically one of the most important zoonoses. Hydatid cyst develops in the internal organs of human and herbivore intermediate hosts, mainly in the liver and lung [16-17]. In Egypt the prevalence of hydatidosis still a point under investigation, In the present study, the incidence of HC was18.9 % in slaughtered camels from Cairo abattoirs. This result was in agreement with [18-19] who recorded 18.9% and 17.61% in slaughtered camels in Ismailia and Cairo abattoirs, Egypt, This result considered to be lower than that mentioned by [20] who found (7.67%) in camels in Assiut governorate, Egypt. In addition, the presently incidence in examined slaughtered cattle was 3.3%, this result disagreement with [21,22] who recorded (31.8%) and (13.58%) in Hawassa, Ethiopia and South West of Iran. However, [20, 23] who mentioned there is no infection was recorded in the examined cattle and buffaloes. Presently, incidence of donkey HC infection was (14.2%) this finding considered to be higher than that result by [24].

The current study indicated that the rate of infection in camel was higher in Lung (94.3 %) than in liver (5.7 %). An observation in accordance with that noticed in Egypt camel the previously authors found that 100% [20], 94.6% [19] and 63.7%. [25] in examined lung camel.

The majority of infected donkeys (90.2%) in the present study harbored hydatid cysts in their liver. This result was in agreement with [8], who revealed that the majority of infected donkeys (70%) harbored HC in their livers at Beni-Suef, Egypt.

Based on the epidemiology and molecular studies, the fertility of cyst is one of the most important factors in the epidemiology of *E. granulosus*. The fertility of cyst varies depending on the hosts and geographical situations [26]. In the current study, fertility rate of HC in camel and donkeys have been found to be 42 (79.24%) in camel lung and 15 (29.4%) in donkey liver. The high rate of fertile cyst may indicate that the cause of infection in investigated animals might be due to camel and donkeys strain (G6 and G4). As such genotype is commonly recognized as a predominating species of *E. granulosus* in Mediterranean countries [27].

Molecular genetics study has been carried out to identify the genetic characters of HC obtained from the infected camel and donkeys. After PCR amplification of ITS1 gene, similar amplified DNA band of the same molecular size marker at 1115bp were recorded in the different isolates. No band variation of ITS1 gene could be detected by PCR-RFLP after using the two restriction enzymes, MSP1 and RSA1, This meaning absence of ITS1 variant which could not differentiated using these two restriction enzymes. This was in agreement with [28-29]. In the author's opinion and in agreement with [28] absence of variation in amplified ITS1 and indistinguishable genetic character in PCR-RFLP, meaning that the analyzed camel and donkey HC samples are infected with *E. granulosus* of sheep strain. This can be accepted as all examined animals are from the same localities. Moreover more research are continue aiming to further identification of hydatid infection of camel and donkeys based on PCR amplification and sequence of mitochondrial genes.

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