Global Veterinaria 12 (1): 98-103, 2014 ISSN 1992-6197 © IDOSI Publications, 2014 DOI: 10.5829/idosi.gv.2014.12.01.81157

Appraisal of Some Heavy Metals in Organ Meat from Non-industrialized Areas of Faisalabad, Pakistan

¹Ayesha Mehmood, ^{1,2}Raja Adil Sarfraz, ²Abdul Qudoos and ¹Fareena Akbar

¹Department of Chemistry and Biochemistry, Faculty of Sciences, University of Agriculture, Faisalabad, Pakistan ²Central Hi-Tech Laboratory, University of Agriculture, Faisalabad, Pakistan

Abstract: The aim of this investigation was to determine the current situation and distribution of heavy metals; cadmium (Cd), lead (Pb) and nickel (Ni) in organ meat of buffalo and goat reared in non-industrialized rural areas of district Faisalabad, Pakistan. Meat samples were subjected to wet digestion and analyzed by atomic absorption spectrophotometer. There was a significant difference (P<0.05) in the concentration of Ni, Pb and Cd in organ meat of buffalo and goat. Mean concentration of Cd, Pb and Ni in meat samples of buffalo was 0.04, 0.29, 1.58 μ g/g and of goat was 0.09, 1.28, 1.43 μ g/g, respectively. The distribution and concentration of metals varied in different organs of both species. Mean Pb concentration in goat organ meat was higher than the permissible limits set internationally. Mean Ni concentration was higher in meat of both species. Mean of Cd concentration in meat samples of both species were below the permissible limits. The results of the current investigation indicated polluted environment even in non-industrialized areas with different types of heavy metals approached to environment through various sources.

Key words: Heavy Metals • Organ Meat • Buffalo Goat • Non-Industrialized Areas • Faisalabad

INTRODUCTION

Meat is essential for growth and maintenance of good health and mainly composed of proteins, fat, carbohydrate and some important essential elements. The need for mineral compounds depends on age, physiological state and feed intake as well as on living conditions [1]. Direct exposure, polluted water and crops grown on irrigated sewerage water and industrial effluents are the sources of metal contamination to animals and humans [2]. Among various environmental pollutants, heavy metals are directly related to human health. Heavy metals have density more than 5g/cm3, atomic weight 63.546 to 200.590 [3] and a specific gravity greater than 4.0 [4]. Living organisms usually need some of heavy metals up to certain limits, while excess accumulation leads to severe harmful effects [3, 5]. Increasing industrialization facilitates entering of metals into the environment. These metals persist permanently as they cannot be degraded in the environment. Heavy metals enter into the food chain, make their passage into the tissues and often have direct

toxic effects [1]. Lead, cadmium, mercury and arsenic are among the main toxic metals which accumulate in food chain [6]. Contamination of meat and other edible tissues with hazardous metals is a matter of concern for humans. Heavy metals are toxic in nature and even at relatively lower concentrations can cause adverse effects [7, 8]. Processing of meat [8, 9] and rearing of livestock in proximity to polluted surroundings are key factors for their pollution in meat [10, 11].

Open literature documented the bioaccumulation and toxicity of heavy metals residues in animal tissue as hazardous potential [12-14]. Heavy metals contamination in meat is due to eating of spoiled feed by animals and also due to rearing of animals near heavy metal contaminated area [15-17]. Instances of heavy metals contamination in meat products during processing have also been reported [8, 9]. Similarly, the distribution and localization of some heavy metals in the tissues of some calf organs indicated higher levels of trace metals in livers, kidneys and small intestines [18]. The local community likes organ meat of animals including liver, kidney, heart

Corresponding Author: Raja Adil Sarfraz, Department of Chemistry & Biochemistry, University of Agriculture, Faisalabad, Pakistan. Tel: +92419200161-70 Ext/3602. etc. and used in different traditional recipes. Considering this point, present study aimed to evaluate the level and distribution of heavy metals (Cd, Pb, Ni) in organ meat of animals reared in non-industrialized areas of district Faisalabad, Pakistan.

MATERIALS AND METHODS

Collection of Organ Meat Samples: Fresh organ meat (lungs, livers, kidneys, hearts; 50 each) of buffalo and goat were purchased from the local meat shops located in non-industrialized rural areas of Faisalabad, Pakistan. It was insured that slaughtered animals were reared in these areas and not brought from unknown localities. Removal of fat from the collected samples was exercised prior to meat collection. The collected samples were kept frozen (-4°C) until further processed for elemental analysis.

Reagents and Standard Solutions: Calibrated standards were prepared by using the commercially available stock solution (Applichem[®]). All the glass apparatus used throughout the process of analytical work were immersed in 8N HNO₃ overnight and finally rinsed with several changes of de-ionized water, air dried and stored.

Pre-treatment of Samples: The collected samples were decomposed by wet digestion method [19]. Accurately weighed meat samples (1gm) were placed in digestion flasks and concentrated nitric acid (10 ml) was added. The digestion flasks were heated (60-70 °C) on a hot plate for 20 minutes. After cooling, 5 ml of HCLO₄ was added and heated vigorously till the white fumes appeared and mixture volume reduced to 2-3 ml. The content of the flask was filtered into a 50 ml volumetric flask and diluted up to the desired volume by adding de-ionized water. The blank solution was also prepared in the similar acid matrix.

Analysis of Heavy Metals: Heavy metals (Cd, Pb, Ni) in digested meat samples were analyzed using atomic absorption spectrophotometer (Hitachi Polarized Zeeman AAS, Z-8200, Japan) following the conditions described in AOAC [20]. The instrumental operating conditions for the analysis of heavy metals are summarized in Table 1.

Data Analysis: The collected data is presented as mean and standard deviation and subjected to one-way analysis of variance (ANOVA). All statistical calculations were performed with SPSS 9.0 for Windows.

RESULTS

Cadmium: Cadmium level was highest in liver (0.08 μ g/g) and kidneys (0.14 μ g/g) whereas non-detectable in heart of both species (Table 2). Variation in the distribution of Cd in different organs of both species was significant (P<0.05). Mean Cd concentration was higher (0.09 μ g/g) in goat than buffalo (0.04 μ g/g) organ meat.

Lead: The mean concentration of Pb varied significantly (P<0.05) in organ meat of buffalo (0.29 μ g/g) and goat (1.28 μ g/g). Pb level was highest (0.54 μ g/g) and lowest (0.18 μ g/g) in buffalo kidneys and heart respectively (Table 2). Its concentration in heart, liver and lungs was non-significant (P>0.05) whereas, it was significant (P<0.05) in kidneys. In case of goat organ meat, it was maximum (2.67 μ g/g) in liver and minimum (0.44 μ g/g) in heart with a significant (P<0.05) variation in the examined organs.

Nickel: Nickel concentration in organ meat of buffalo and goat has been depicted in Table 2. In buffalo organ, highest concentration of Ni was found in lungs (2.49 μ g/g) and lowest in heart (0.90 μ g/g). Concentration of Ni varied significantly (P<0.05) between lungs and kidneys whereas; it was non-significant (P>0.05) in heart and liver.

Table 1: Operational conditions of atomic absorption spectrophotometer for the analysis of heavy metals

| Parameters | Cd | Pb | Ni | |
|--|-----------------------------------|-----------------------------------|-----------------------------------|--|
| Wavelength (nm) | 228.8 | 283.3 | 232.0 | |
| Slit Width (nm) | 1.3 | 1.3 | 0.2 | |
| Lamp Current (mA) | 7.5 | 7.5 | 10 | |
| Burner Head | Standard type | Standard type | Standard type | |
| Flame | Air-C ₂ H ₂ | Air-C ₂ H ₂ | Air-C ₂ H ₂ | |
| Burner Height (mm) | 5.0 | 7.5 | 7.5 | |
| Oxidant gas pressure (Flow rate) (kpa) | 160 | 160 | 160 | |
| Fuel gas pressure (Flow rate) (kpa) | 6 | 7 | 7 | |

 $0.89\pm0.01^{\circ}$

1.11±0.01^b

1.28±0.25^A

 0.97 ± 0.01^{d}

1.95±0.02b

1.40±0.02

 $1.40\pm0.02^{\circ}$

1.43±0.10^B

Concentration Elements Organs Buffaloes Goats Cadmium Heart $0.00{\pm}0.00^{f}$ $0.00{\pm}0.00^{f}$ Liver 0.08±0.00° 0.08±0.00° 0.05 ± 0.00^{d} 0.12±0.00^b Lungs 0.04±0.00e 0.14 ± 0.00^{4} Kidnev Mean $0.04{\pm}0.00^{B}$ 0.09±0.01^A Lead Heart $0.18{\pm}0.00^{\rm f}$ 0.44 ± 0.00^{6} Liver 0.22 ± 0.00^{f} 2.67±0.03ª

 0.22 ± 0.00^{f}

 0.54 ± 0.00^{d}

0 29±0 04^B

 $0.90{\pm}0.01^{d}$

 0.97 ± 0.01^{d}

2.49±0.03ª

1.98±0.02b

1.58±0.20^A

Lungs

Kidnev

Mean

Heart

Liver

Lungs Kidney

Mean

Nickel

| Table 2: | Concentration of heavy metals $(\mu g/g)$ in different organs of buffalo | |
|----------|--|--|
| | and goat | |

Mean of individual elements sharing similar letter in a row or in a column are statistically non-significant (P>0.05). Small letters represent comparison among interaction means and capital letters for overall means.

Highest concentration of Ni was observed in liver (1.95 μ g/g) and lowest (0.97 μ g/g) in heart of goat. Deposition of Ni was significant (P<0.05) in heart and liver whereas non-significant (P>0.05) in lungs and kidneys.

DISCUSSION

None of the organs of both species exceeded the permissible limit of 1 ppm recommended by FAO/WHO [21]. Highest Cd concentration was observed in goat kidneys that were also below the permissible limit. The relatively higher level of Cd in kidneys than in liver may be due to detoxification function of this organ where the metal is accumulated [22, 23]. Some previous studies conducted in the province of Punjab, Pakistan also reported under permissible Cd levels. Mariam et al. [24] reported under permissible limit of Cd in liver of animals at Lahore. Similarly, under permissible levels of Cd was also reported by Aslam [11] in liver, lungs and kidneys of goat at Faisalabad. In Egypt, Khalafalla et al. [25] also reported this trend of Cd accumulation in liver and kidneys of cattle. In contrast, higher levels of Cd have been recorded by Kramer et al. [26], Antoniou et al. [27], Jorhem et al. [28], Niemi et al. [29], Aranha [22], Roga-Franc et al. [30] and Lopez-Alonso et al.[31]. Akan et al. [32] reported 0.22, 0.17 and 0.44, 0.39 mg/g Cd in liver and kidneys of cow and goat. This reported high level of cadmium may be due to species, seasonal variation, feeding and rearing of livestock on contaminated land causing high accumulation of this metal in different organs and tissues.

Lead concentration in goat organs was higher than the permissible limit of one ppm set internationally. This higher trend was supported by the results of Sabir et al. [16] that indicated higher Pb concentration in mutton (≤ 2 ppm) and beef (up to 3 ppm). Mariam et al. [24] found higher levels of Pb than permissible limit of one ppm [33] in liver (2.18; 4.25 ppm) and kidneys (2.02; 3.85 ppm) of beef and mutton. Much high levels 7.72 and 14.38 mg/kg of Pb in lungs and kidneys of cattle and goat has been reported by Aslam [11] in Faisalabad, Pakistan. High trend of Pb accumulation in liver and kidneys of animals has been reported by Aranha [22] and Danev et al. [34] indicating that liver (86%) and kidney (100%) were contaminated above the limits set by the country regulations. Similarly, Maldonado et al. [35] noted significantly higher levels of lead in liver and kidneys during lactation in rats. The levels of Pb in liver, kidney and meat of beef, mutton caprine and chicken ranged from 0.1 to 1.34 µg/g [32]. Spierenburg et al. [36] reported significantly high amounts of Pb in liver and kidneys of cattle in Nigeria, Pb concentration in different organs and muscles of cow ranged from non-detectable to 1.22 mg/kg [37]. In contrast, under permissible levels of Pb has been noted by Stabel-Taucher et al. [38] and Jorhem et al. [28]. Low level in liver (42.70 µg/kg) and kidney (109.42 µg/kg) of cattle slaughtered in the Beni-Suef abattoir in Egypt was reported by Khalafalla et al. [25]. Accumulation of absorbed lead normally occurred in liver and kidneys [39, 40]. Airborne sources like industrial emissions and combustion of fuel having Pb additives may affect grazing animals [41]. Chances of Pb accumulation in different organs are mostly in animals that reared around refineries than reared in rural areas [42]. Abou Donia [43] pointed out that concentration of Pb was higher in livers and kidneys of animals from industrial areas. Indication of lower levels of Pb in some organs during the present investigation may be the results of the notable reduction in emission from automobiles resulting into reduced Pb level in meat tissues during the recent decades [42]. In spite of notable reduction in Pb level worldwide, the present results indicated that Pb residues still exists in non-industrialized rural localities. The entry route may be sewerage drains passing through this area containing untreated effluents from industrial or chemical processing units located way from this site. It may happen that from sewerage drains, the untreated waste water may find their way to irrigation channels and consequently through soil pollute the fodder.

It is obvious from the results that some organs contain higher amount of Ni. This element is present in a number of enzymes in plants and microorganisms. In humans, it regulates prolactine and stabilization of RNA and DNA structures [44]. In contrast, excessive intake of Ni can cause severe allergic reaction [45]. Majority of food products contains Ni less than 0.5 mg/kg fresh weight [46]. According to Solomons et al. [47], meat contains low levels of Ni ($<0.2 \mu g/g$). Open literature indicated that there is no set recommended amounts of Ni intake. However, the hypothetical human requirement for Ni would be 16 to 25 µg/1000 kcal or about 75 µg of elemental Ni per day [47]. In human, Ni deficiency has not been observed [48]. A wide variation in Ni levels in organ meat has been reported. Korenekova et al. [15] reported 0.23-0.35 mg Ni /kg in beef. Nwude et al.[37] documented 0.09 to 0.44 mg/kg of Ni level in liver and 0.30 to 0.57 mg/kg in kidneys. Sabir et al. [16] reported Ni concentration as 2ppm and 1ppm in mutton and beef, respectively. Akan et al. [32] found Ni concentration in liver $(0.29; 0.19 \ \mu g/g)$ and kidneys $(0.16; 0.13 \ \mu g/g)$ of cattle and goats. Aslam [11] documented Ni concentration in liver (12.18 mg/kg), lungs (13.27 mg/kg) and kidney (5.75 mg/kg) of goat meat obtained along the main sewerage drains of Faisalabad city. Higher level of Ni (101mg/kg) was also observed in livers of poultry at Faisalabad [49].

In conclusion, the results mentioned in preceding lines clearly indicated that concentration of Pb and Ni has been found higher in organ meat of buffalo and goat than the permissible limits of 1ppm and 0.2ppm, respectively. The presence of heavy metals residues in meat obtained from non-industrial areas is an indicative of continuous pouring of untreated industrial waste water from sewerage drains passing through these areas. Consequently, uptake of heavy metal residues by the fodder cultivated on this soil may occur that may be a cause of this pollution in non-industrialized rural areas.

ACKNOWLEDGEMENT

The authors thankfully acknowledge the financial support extended by Higher Education Commission (HEC) of Pakistan and technical support provided by Central Hi-Tech Laboratory staff, University of Agriculture, Faisalabad, Pakistan to carry out this work.

REFERENCES

- Baykov, B.D., M.P. Stoyanov and M.L. Gugova, 1996. Cadmium and lead bioaccumulation in male chickens for high food concentrations. Toxicological and Environmental Chemistry, 54: 155-159.
- Ward, N.I and J.M. Savage,1994. Elemental status of grazing animals located adjacent to the London Orbital (M25) motorway. Science of the Total Environment, 146(147): 185-189.
- Kennish, M.J, 1992. Ecology of Estuaries Anthropogenic Effects. CRC Press, Boca Raton, pp: 494.
- Connell, D.W. and G.J. Miller, 1984. Chemistry of Ecotoxicology Pollution. John Wiley and Sons, New York.
- Chitmanat, C. and S. Traichaiyaporn, 2010. Spatial and temporal variations of physical-chemical water quality and some heavy metals in water, sediments and fishes of the Mae Kuang River, Northern Thailand. International Journal of Agriculture and Biology, 12: 816-820.
- 6. Dermirezen, D. and K. Urue, 2006. Comparative study of trace elements in certain fish, meat and meat product. Meat Science, 74: 255-260.
- Mahaffey, K.R., 1977. Mineral concentrations in animal tissues: Certain aspects of FDA regulatory role. Journal of Animal Science, 44: 509-515.
- Santhi, D., V. Balakrishnan, A. Kalaikannan and K.T. Radhakrishnan, 2008. Presence of heavy metals in pork products in Chennai (India). American Journal of Food Technology, 3: 192-199.
- Brito, G., C. Diaz, L. Galindo, A. Hardisson, D. Santiago and F. Garcia Montelongo, 2005. Levels of metals in canned meat products: Intermetallic correlations. Bulletin of Environmental Contamination and Toxicology, 44: 309-316.
- Sedki, A., N. Lekouch, S. Gamon and A. Pineau, 2003. Toxic and essential trace metals in muscle, liver and kidney of bovines from a polluted area of Morocco. Science of the Total Environment, 317: 201-205.
- Aslam, B., 2010. Determination of heavy metal residues in the milk and meat of cattle and goat. Ph.D. Thesis. Department of Physiology and Pharmacology, University of Agriculture, Faisalabad, Pakistan.

- Zasadowski, A., D. Barski, K. Markiewicz, Z. Zasadowski, A. Spodniewska and A. Terlecka, 1999. Levels of cadmium contamination of domestic animals (Cattle) in the Region of Warmia and Masuria. Polish Journal of Environmental Studies. 8: 443-446.
- Satoh, M., H. Koyama, T. Kaji, H. Kito and C. Tohyama, 2002. Perspectives on cadmium toxicity research. Tohoku Journal of Experimental Medicine. 196: 23-32.
- Thompson, J. and J. Bannigan, 2008. Cadmium: Toxic effects on the reproductive system and the embryo. Reproductive Toxicology, 25: 304-315.
- Korenekova, B., S. Magdalena and P. Nai, 2002. Concentration of some heavy metals in cattle reared in the vicinity of a metallurgic industry. Veterinarski Arhiv, 72: 259-267.
- Sabir, S.M., S.W. Khan and I. Hayat, 2003. Effect of environmental pollution on quality of meat in district Bagh, Azad Kashmir. Pakistan Journal of Nutrition. 2: 98-101.
- Miranda, M., M. Lopez-Alonso, C. Castillo, J. Hernandez, F. Prieto and J.L. Benedito, 2003. Some toxic elements in liver, kidney and meat from calves slaughtered in Asturias (Northern Spain). European Food Research and Technology, 216: 284-289.
- Horky, D., J. Illek and A. Pechova, 1998. Distribution of heavy metals in calf organs. Veterinary Medicine. 43: 331-342.
- Richards, L.A., 1968. Diagnosis and Improvement of Saline and Alkaline Soils. 1st Ed, Agri. Handbook No. 60. IBH Publications Company, New Delhi, India.
- 20. AOAC., 1990. Official Methods of Analysis. Association of Official Analytical Chemists. Arlington, VA, USA.
- FAO., 2000. Food and Agriculture Organization/World Health organization. Report of the 32rd Session of the Codex Committee of the Food Additives, FAO, Rome, pp: 36-40.
- Aranha Kumar de., 1994. Environmental Chemistry, 3rd Ed. New Age International Limited Publisher, New Delhi, pp: 213-219.
- Stoyke, M., K.D. Doberschutz and K. Lusky, 1995. Heavy metal contents (cadmium, lead and mercury) in selected feedstuffs, organs and tissue in cattle from different sites of Brandenburg. Mengen-Spurenelemente Arbeitstag, 15: 269-276.

- Mariam, I., S. Iqbal and S.A. Nagra, 2004. Distribution of some trace and macrominerals in beef, mutton and poultry. International Journal of Agriculture and Biology, 6: 816-820.
- Khalafalla, F.A., F.H. Ali, F. Schwagede and M.A. Abd-El-Wahab, 2011. Heavy metal residues in beef carcasses in Beni-Suef abattoir, Egypt. Veterinaria Italiana, 47: 351-361.
- Kramer, H.L., J.W. Steiner and P.J. Vallely. 1983. Trace elements concentrations in the liver, kidney and muscle of Queensland cattle. Environmental Contamination and Toxicology, 30: 588-594.
- Antoniou, V., H. Baukali-Papadopoul, P. Epivatianos and B. Nathanael, 1989. Cadmium concentration in beef consumable tissues in relation to age of animals and area of their breeding. Bulletin of Environmental Contamination and Toxicology, 43: 915-919.
- Jorhem, L., S. Slarach, B. Sundstrom and B. Phlin, 1991. Lead, cadmium, arsenic and mercury in meat, liver and kidney of Swedish pigs and cattle in 1984-1988. Food Additives and Contaminants, 8: 201-212.
- Niemi, A., E. Venalainen, T. Hirvi, J. Hirn and E. Karppanen, 1991. The lead, cadmium and mercury concentration in muscle, liver and kidney from finishing pigs and cattle during 1987-1988. Z Lebensum Unters Forsch, 192: 427-429.
- Roga-Franc, M., T. Kosla and E. Rokicki, 1996. Cadmium concentration in organs of dairy cows depending on that element contents in meadow cover. Mengen-Spurenelem Arbeistag, 16: 393-397.
- Lopez-Alonso, M., J.L. Benedito, M. Miranda, C. Castill, J. Hernandez and R.F. Shore, 2000. Toxic and trace elements in liver, kidney and meat from cattle slaughtered in Galicia (NW Spain). Food Additives and Contaminants, 17: 447-457.
- 32. Akan, J.C., F.I. Abdulrahman, O.A. Sodipo and Y.A. Chiroma, 2010. Distribution of Heavy Metals in the Liver, Kidney and Meat of Beef, Mutton, Caprine and Chicken from Kasuwan Shanu Market in Maiduguri Metropolis, Borno State, Nigeria. Research Journal of Applied Sciences, Engineering and Technology, 2: 743-748.
- 33. ANZFA-Australia New Zealand Food Authority. URL:http://www.anzfa.gov.au
- Danev, M., V. Serafimovska, P. Sekulovski, E. Stojkovic, B. Krstic and M. Zoric, 1996. Cadmium contamination of beef. Tehnol Mesa, 37: 19-21.

- Maldonado, V.M., S.J. Cerbon, M.A. Albores, L.C. Hernandez and J.V. Calderonsalinas, 1996. Lead: intestinal absorption and bone mobilization during lactation. Human and Experimental Toxicology. 15: 872-877.
- Spierenburg, T.J., G.J. De-Graaf and A.J. Braars, 1988. Cadmium, zinc, lead and copper in livers and kidney of cattle in the neighborhood of zinc refineries. Environmental Monitoring and Assessment. 11: 107-114.
- Nwude, D.O., J.O. Babayemi and O. Abhulimen, 2011. Metal quantification in cattle: A case of cattle at slaughter at Ota Abattoir, Nigeria. Journal of Toxicology and Environmental Health Sciences. 3: 271-274.
- Stabel-Taucher, R., E. Nurmi and E. Karppanen,1975. Content of copper, zinc, lead, cadmium and mercury in muscle, liver and kidney of fresh cattle. Journal of Scientific AgriculturalSocietyofFinland, 47: 469.
- Buck, W.B., 1975. Toxic materials and neurologic disease in cattle. Journal of American Veterinary Medical Association, 33: 2395-2401.
- Hoseini, H. and M.S. Tahami, 2012. Study of heavy metals (Pb and Cd) concentrations in liver and muscle tissues of *Rutilus frisii kutum*, Kamenskii, 1901 in Mazandaran province. Global Veterinaria. 9: 329-333.
- Humphreys, D.J., 1991. Effects of exposure to excessive quantities of lead on animals. British Veterinary Journal, 147: 18-30.

- Erivo, R., R. Makela-Kurtto and J. Sippala, 1990. Chemical characteristics of Finnish agricultural soils in 1974 and 1987. In Acidification in Finland (P. Kauppi, P. Antilla andKenttamies, eds). Springer-Verlag, Berlin, pp: 217-234.
- Abou Donia, M.A., 2008. Lead concentration in different animals muscles and consumable organs at specific locations in Cairo. Global Veterinaria. 2: 280-284.
- Khurshid, J.S. and H.Q. Iqbal, 1984. The role of inorganic elements in the human body. Nucleus. 21: 3-23.
- 45. Murtic, C.R.K. and P. Viswanathan, 1989. Toxic Metals in the Indian Environment. Wiley Eastern Ltd., New Delhi.
- 46. IARC., 1990. Nickel and nickel compounds. In Chromium, Nickel and Welding. Lyon, International Agency for Research on Cancer. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. 49: 257-445.
- Solomons, N.W., F. Viteri, T.R. Shuler and F.H. Nielsen, 1982. Bioavailability of nickel in man: effects of foods and chemically-defined dietary constituents on the absorption of inorganic nickel. Journal of Nutrition, 112: 39-50.
- EVM, 2003. Nickel, risk assessment. Expert Group on Vitamins and Minerals (EVM). Food Standards Agency, London, United Kingdom.
- Rehman, K.U., S. Andleeb, A. Mahmood, S.M. Bukhar, M.M. Naeem and K. Yousaf. 2012. Translocation of Zinc and Nickel from poultry feed to broilers and their excretion through litters. Global Veterinaria, 8: 660-664.