

## Effect of Exposure to Sodium Arsenite During Embryonic Development on Steroidogenic Hormones, Marker Enzymes and Gonadotrophine Hormones in Male Mice

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**Abstract:** The present study aimed to assess the possible interference of sodium arsenite in F1 generation male mice with special reference to steroidogenic marker enzymes. Mice were divided in to two groups. The mice in first were served as control and received normal tap water. Sodium arsenite administered orally to mice in the second group during pregnancy and lactation at a dose level of 0.4 ppm and analyzed for spermatogenesis and steroidogenesis in next generation adult male mice. The activity levels of selected steroidogenic marker enzymes (3 $\beta$ -hydroxysteroid dehydrogenase and 17 $\beta$ -hydroxysteroid dehydrogenase) decreased significantly in mice exposed to sodium arsenite. The circulatory levels of testosterone decreased in experimental mice with significantly increase in follicle stimulating hormone. The decreased levels of testosterone with elevated follicle stimulating hormone and luteinizing hormone levels in mice exposed to arsenic during early stages of development are indicative of intact pituitary-testicular axis. The results indicate that exposure to arsenic during early stages of development suppresses the male reproduction in adults. Thus, we conclude that the potential of reproduction is programmed, to some extent, in the early stages of development and hence any toxic insult during embryonic development and lactation suppresses male reproductive potential in adulthood.

**Key words:** Sodium Arsenite • Hormones • Enzymes • Male Reproduction • Testes • Gestation and Lactation

### INTRODUCTION

Arsenic, a non-essential trace element and a potent toxic metalloid has drawn increasing attention in recent years as a major pollutant of drinking water. Higher levels of inorganic arsenic occurs naturally in ground water of many parts of the world including India and millions of people are exposed worldwide to the drinking water containing this known carcinogen in excess amount [1]. Epidemiological data indicates that more than six million people residing in different areas of West Bengal, India are exposed to arsenic contaminated drinking water and more than 300,000 people were reported with signs of arsenic toxicity [2].

Arsenic exposure has been associated with an increased risk of dermatitis along with hyperkeratosis, gangrene and tumors of skin, bladder, liver, kidney, lung,

prostate and other tissues [3]. Epidemiological reports from Ukraine, Taiwan and Bangladesh revealed that the intake of arsenic contaminated food and water caused reproductive disturbances in woman [4], adverse pregnancy outcomes [5] and also spontaneous abortions [6]. Arsenic has been suspected to be the cause for reproductive failure in male workers at a copper smelter in [7] intoxication in experimental animals has been associated with inhibition of steroidogenesis [7, 8] and elevation of adrenocortical steroidogenesis [9].

There is a lack of literature and data related to the exposure to arsenic during prenatal and neonatal period on reproduction in adults, particularly at the dose levels occurring in drinking water in wide areas of India and in other countries where this element is present in the range above the admissible limit (0.01 ppm according to the World Health Organization) [10]. The present study was

conducted to assess the effect of exposure to 0.4 ppm of sodium arsenite through drinking water during embryonic development and lactation on reproductive hormones and enzymes of adult male mice (F1 generation).

## MATERIALS AND METHODS

**Animals:** Twenty Swiss Albino mice were bred at Department of Biotechnology, S.V. University, Tirupati. Animals were maintained in polypropylene cages lined with paddy husk under a well regulated light and dark (12h:12h) schedule at  $27 \pm 1^\circ\text{C}$  with relative humidity of 75%. Animals were given food and water *ad libitum*. The mice pellet feed was purchased from Kamadhenu Agencies, Bangalore, India. Healthy mice of 90 days age were selected for present study.

**Test Chemical:** Sodium Arsenite was purchased from S.D fine chemicals (Mumbai, India) used as test chemical. This compound was dissolved in normal water to obtain the final concentration of the 0.4 ppm/ kg body weight of the animal.

**Experimental Design:** Pregnant mice were divided randomly into two groups consisting of ten animals in each group. The animals in group 1 were allowed *ad libitum* access to tap water without sodium arsenite while the animals in group 2 were allowed *ad libitum* access to tap water containing 0.4 ppm of sodium arsenite during gestation and lactation periods. The mice were allowed to deliver the pups and pups, after weaning, were grown on normal diet and tap water (without sodium arsenite) up to 60 days and used for experimentation. The experimental animals were sacrificed by cervical dislocation. All animal procedures were approved by the Institutional Animal Ethics Committee at Sri Venkateswara University.

**Assay of Testicular Steroidogenic Enzymes:** The  $3\beta$ -hydroxysteroid dehydrogenase (E.C. 1.1.1.51; conversion of NAD to NADH) and  $17\beta$ -hydroxysteroid dehydrogenase (E.C. 1.1.1.61; conversion of NADPH to NADP) activities in the testis were assayed using the method described [11]. The enzyme assays were made under the conditions following zero order kinetics after preliminary standardization regarding linearity with respect to substrate concentration, time of incubation and enzyme concentration.

The reaction mixture in a volume of 2.0 ml contained: 100  $\mu\text{moles}$  of sodium pyrophosphate buffer (pH 9.0), 0.5  $\mu\text{mol}$  of cofactor NAD for  $3\beta$ -HSD and NADPH for  $17\beta$ -HSD, 0.08  $\mu\text{mol}$  of substrate (dehydroepiandrosterone

for  $3\beta$ -HSD and androstenedione for  $17\beta$ -HSD) and 20 mg equivalent of microsomal protein as enzyme source. The reactions were carried out in a quartz cuvette of 1.0 cm path at  $23^\circ\text{C}$ . The change in the absorbance was measured at 340 nm at 20s intervals for 5 min in a UV-Vis spectrophotometer (Hitachi model U-2001) against controls. Protein content in the enzyme source was estimated by the method [12] using bovine serum albumin as standard.

## Determination of Serum Testosterone, FSH and LH

**Levels:** Radioimmuno assay of serum testosterone was performed by the method [13]. The sensitivity of the assay was calculated as 0.002 ng/ml. Serum FSH and LH were assayed [14]. Iodination of rFSH and rLH with  $\text{I}^{125}$  was performed by the method [15] using chloramineT as an oxidizing agent. The sensitivity of the assay was calculated as 0.004 and 0.006 for FSH and LH respectively. All the samples were run at the same time to avoid interassay variation.

**Statistical Analysis of the Data:** The data were presented as mean  $\pm$  SEM. Statistical analysis was performed using analysis of variance (ANOVA) followed by Dunnett's test, using SPSS 10.0 version.

## RESULTS

No mortalities were observed in control or in experimental groups. No behavioral abnormalities were observed in experimental mice.

The activity levels of  $3\beta$ -hydroxysteroid dehydrogenase decreased significantly in the testis of mice exposed to arsenic during early stages of development when compared with the corresponding controls (Table 1). Serum FSH, LH and testosterone concentrations in mice exposed to arsenic during early stages of development were shown in Table 1. The levels of serum testosterone decreased in adult mice exposed to arsenic during early stages of development when compared with the corresponding group of control animals. Whereas the levels of serum FSH and LH increased significantly in experimental mice with respect to the corresponding group of control mice (Table 1).

## DISCUSSION

The present study was aimed to determine the reproductive toxic effects of mice exposed to arsenic during embryonic development and lactation. The route chosen in this study for exposure was via drinking water

Table 1: Effect of gestational and lactational exposure to sodium arsenite on testicular 3- $\beta$  HSD and 17- $\beta$  HSD activity levels and serum FSH, LH and testosterone levels in adult mice

Parameters	Control	Arsenite	Anova
3 $\beta$ -HSD ( $\mu$ mol NAD converted to NADH/mg. Protein/h)	0.0262 $\pm$ 0.0045	0.0079 $\pm$ 0.0012 (- 99.21)	F <sub>2,21</sub> =14.063 P<0.01
17 $\beta$ -HSD ( $\mu$ mol NADPH converted to NADP/mg. Protein/h)	0.0168 $\pm$ 0.0019	0.0039 $\pm$ 0.0011 (- 99.61)	F <sub>2,21</sub> =14.063 P=0.1277 N.B.
Testosterone (ng/ml)	7.89 $\pm$ 1.21	4.25 $\pm$ 0.76 (-46.13)	F <sub>2,21</sub> =2.535 P=0.1652 N.B.
FSH (ng /ml)	5.26 $\pm$ 0.82	11.21 $\pm$ 1.96 (+113.11)	F <sub>2,21</sub> =5.713 P<0.05
LH (ng/ml)	1.25 $\pm$ 0.03	3.89 $\pm$ 0.11 (+211.2)	F <sub>2,21</sub> =13.444 P<0.01

Values are Mean  $\pm$  SEM of 8 animals. Values in parentheses are percent change from the control. Values are significantly different at \*p<0.05

through mothers to mimic human exposure and to reflect the impact on fertility of next generation. In the present study reproductive potential of male mice was measured using activity levels steroidogenic enzymes, circulatory levels of gonadotropins and testosterone as biological parameters.

The arsenic dose selected in the present study was not resulted in any toxic symptoms in mice. No mortality and no behavioral abnormalities were recorded in experimental mice indicating the arsenic do not exhibit any toxicity at the selected dose level. The results of the present investigation demonstrate the adverse effect of sodium arsenite on production of reproductive enzymes and hormones. Maintenance of spermatogenesis in mice depends upon adequate testosterone concentrations [16]. The levels of serum testosterone were decreased in adult mice exposed to arsenic during early stages of development. The decrease in serum testosterone could be due to diminished responsiveness of Leydig cells to leutinizing hormone and/or the direct inhibition of testosterone steroidogenesis. In steroidogenesis,  $\Delta^5$ , 3- $\beta$ HSD and 17- $\beta$ HSD are the key regulatory enzymes [17]. A significant decrease in the activity levels of these steroidogenic enzymes in testis of experimental mice indicate decreased steroidogenesis, which in turn may suppress the reproductive activities in the male mice. This is in agreement with the previous findings where arsenic treatment was associated with inhibition of testicular steroidogenesis in rat [18]. This alteration in steroidogenic enzyme activity in experimental mice may be the result of changes in the levels of plasma FSH and LH, since these are the regulators of Hydroxysteroid dehydrogenase activities [19]. The elevated levels of serum FSH and LH with lowered circulatory testosterone levels in experimental mice are indicative of intact pituitary-testicular axis. Testosterone plays an important role in attachment of the germ cells in seminiferous tubules. Low levels of intra-testicular testosterone may lead to detachment of germ cells from seminiferous epithelium and may initiate cell apoptosis [20]. The increase in the levels of serum FSH could be due to the

impairment of spermatogenesis by the arsenic on the spermatogenic compartment or through the inhibition of testosterone production. Thus, the increase in the levels of serum FSH reflect the germ cell loss in the spermatogenic compartment or damage to the sertoli cells, thereby affecting the feedback regulation of FSH secretion [21].

This study has relevance since embryos are more sensitive to arsenic toxicity and arsenic is a common toxicant in several parts of the world, including India.

## REFERENCES

1. Chappell, W.R., B.D. Beck, K.G. Brown, R. Chaney, C.C. Richard, K.J. Irgolic and D.W. North, 1997. Inorganic arsenic: A need and an opportunity to improve risk assessment. *Environ. Health. Perspect.*, 105: 1060-1065.
2. Chakraborti, D., M.M. Rahman, K. Paul, U.K. Chowdhury, M.K. Sengupta, D. Lodh, C.R. Chanda, K.C. Saha and S.C. Mukherjee, 2002. Arsenic calamity in the Indian subcontinent-What lessons have been learned? *Talanta*, 58: 3-22.
3. Gebel, T.W., 1999. Arsenic and drinking water contamination. *Sci.*, 283: 1458-1459.
4. Zadorozhanaja, T.D., R.E. Little, R.K. Miller, N.A. Mendel, R.J. Taylor, B.J. Presley and B.C. Gladen, 2000. Concentrations of arsenic, cadmium, copper, lead, mercury and zinc in human placenta from two cities in Ukraine. *J. Toxicol. Health A.*, 61: 2.
5. Yang, C.Y., C.C. Chang, S.S. Tsai, H.Y. Chuang, C.K. Ho and T.N. Wu, 2003. Arsenic in drinking water and adverse pregnancy outcome in an arseniasis-endemic area in northeastern Taiwan. *Environ. Res.*, 91: 29-34.
6. Ahmad, S.A., M.H. Sayed, S. Barua, M.H. Khan, M.H. Faruquee, A. Jalil, S.A. Hadi and H.K. Talukder, 2001. Arsenic in drinking water and pregnancy outcomes. *Environ. Health. Perspect.*, 109: 629-631.

7. Sarkar, M., G. Ray Chaudhuri, A. Chattopadhyay and N.M. Biswas, 2003. Effect of sodium arsenite on spermatogenesis, plasma gonadotrophins and testosterone in rats. *Asian. J. Androl.*, 1: 27-31.
8. Chattopadhyay, S., S. Ghosh, S. Chaki, J. Debnath and D. Ghosh, 1999. Effect of sodium arsenite on plasma levels of gonadotrophins and ovarian steroidogenesis in mature albino rats: Duration dependent response. *J. Toxicol. Sci.*, 24: 425-431.
9. Vijaya Bhaskar Reddy, M., S.D. Sudheer, P. Sasikala P. Sreenivasula Reddy, S. Hemadri Reddy and A. Karthik, 2011. Effect of Transplacental and Lactational Exposure to Arsenic on Male Reproduction in Mice, Jr. *Rep and Infertility*, 2(3): 41-45, ISSN 2079-2166, © IDOSI Publications.
10. Rahman, M.M., B.K. Mandal, T.R. Chowdhury, M.K. Sengupta, U.K. Chowdhury, D. Lodh, C.R. Chanda, G.K. Basu, S.C. Mukherjee and K.C. Saha, 2003. Arsenic groundwater contamination and sufferings of people in North 24-Parganas, one of the nine arsenic affected districts of West Bengal, India. *J. Environ. Sci. Health Part A Tox. Hazard Subst. Environ. Eng.*, 38: 25-59.
11. Bergmeyer, H.U., 1974. 3- $\beta$ HSD hydroxysteroid dehydrogenase. In: Bergmyer ed. *Methods of Enzymatic Analysis*, Vol. I. New York: Academic Press, pp: 447-489.
12. Lowry, O.H., M.J. Rosebrough, A.L. Farr and R.J. Randall, 1951. Protein measurement with the folin phenol reagent. *J. Biol. Chem.*, 193: 265-275.
13. Rao, A.J., R. Chakraborti, S.G. Kotagi and N. Ravindranath, 1990. Effect of constant infusion of gonadotrophin releasing hormone (GnRH) agonist buserelin and antagonist CDB 2085 A using osmotic mini pumps on testicular function in adult male bonnet monkey (*Macaca radiata*). *Andrologia*, 22: 567-573.
14. Miyachi, Y., J.L. Vaitukaitis, E. Nieschlag and M.B. Lipsett, 1972. Enzymatic radioiodination of gonadotropins. *J. Clin. Endocrinol.*, 34: 23-28.
15. Greenwood, F.O., W.M. Hunter and J.S. Clover, 1963. The preparation of  $^{131}\text{I}$  labeled human growth hormone of high specific activity. *Biochem. J.*, 89: 114-123.
16. Sharpe, R.M., 1987. Testosterone and spermatogenesis. *J. Endocrinol.*, 113: 1-17.
17. Hinshelwood, M.M., M. Demter-Arlotto, G.D. Means and E.R. Simpson, 1994. Expression of genes encoding steroidogenic enzymes in the ovary. In: *Molecular Biology of the Female Reproductive System* (J.K. Findlay, Ed.), Academic Press, London, pp: 129-145.
18. Sarkar, M., N.M. Biswas and D. Ghosh, 1991. Effect of sodium arsenite on testicular  $\Delta^5$ -3 $\beta$  and 17 $\beta$ -hydroxysteroid dehydrogenase activities in albino rat: dose and duration dependent response. *Med. Sci. Res.*, 19: 789-790.
19. Odell, W.D., R.S. Swerdloff, J. Bain, F. Wallesen and P.K. Grover, 1963. The effect of sexual maturation testicular response to LH stimulation of testosterone secretion in the intact rat. *Endocrinology*, 72: 452-464.
20. Blanco-Rodriguez J. and C. Martinez-Garcia, 1997. Apoptosis pattern by oestradiol treatment of the seminiferous epithelium of the adult rat. *J. Reprod. Fertil.*, 110: 61-70.
21. Van Theil, D.H., R.J. Sherins, G.H. Jr. Myers and V.T. Jr. De Vita, 1972. Evidence for a specific seminiferous tubular factor affecting follicle stimulating hormone secretion in man. *J. Clin. Invest.*, 51: 1009-1019.