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Biological Control of Insect Pests in Iraq: 2) an Overview of Microbial Control Research Development

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Abstract: Chemical insect pest control agents are extensively used in Iraq though they are regarded as ecologically unacceptable. Therefore, there is an increased pressure to replace them gradually with environmental friendly alternatives including entomopathogenic micro-organisms, such as bacteria, fungi, viruses and nematodes which are safe to humans and non-target organisms. In Iraq, such field of biological control of insect pests is progressing significantly. There are increasing volume of research programs conducted on several crops with formulations of *Bacillus thuringiensis*, *Beauveria bassiana*, *Metarhizium anisopliae*, *Baculovirus*, *Steinernema* and *Heterorhabditis* spp. for controlling several agricultural pests. This review outlines the current status and knowledge on the potential use of entomopathogenic micro-organisms in insect pests control in Iraq.

Key words: Biological Control • Entomopathogenic Micro-Organisms • Iraq

INTRODUCTION

The use of pathogenic micro-organisms for pest control is not a new concept. Attempts were being made as early as the late 19th century to use fungal diseases to control a number of economic pests. However, interest in all forms of biological and cultural control of insects declined with the dramatic impact of synthetic chemical pesticides in the 1940s and 1950s[1]. Current concerns over resistance to chemicals, non-target effects of pesticides and the cost of production of new compounds have renewed interest in alternative forms of pest control, among which disease causing micro-organisms hold out particular promise as part of Bio-intensive Integrated Pest Management programs. The most commonly used living organisms, which are pathogenic for the pest of interest, include entomopathogenic fungi (EF) (Beauveria bassiana), bacteria (EB) (Bacillus thuringiensis), viruses (nuclearplyhedrosis) and nematodes (Steinernema spp.). The potential benefits to agriculture programs through the use of entomopathogenic microorganisms (EMO) are considerable [2].

In Iraq, such field of biological control of insect pests was started during early 1960s, with humble attempts of utilizing commercial preparations of EB, *B. thuringiensis* and since then several problems limit the crop area where

pathogenic micro-organisms are employed (will be illustrated in the last section). Hence, mentality of relaying upon pesticides as a magic solution is the most important element that limits the use of EMO, billions of dollars were spent on chemical pesticides in Iraq since 1960s, but not allocating enough financial support to survey, identify, formulation and field application of natural enemies (EMO) in well furnish facilities.

This review aims to show the progress accomplished in the field of BC technology research and development, mainly in this part the utilization of EMO (the first part was about parasites and predators[3]), by the Iraqi scientists and specialist since 1970s till the end of 2017. This will include some of the successful applications and finally the problems and challenges facing expansions and large implementations.

Present Status and Knowledge

Entomopathogenic Bacteria: Bacillus thuringiensis (Bt) is primarily a soil saprophyte bacteria and that association between Bt and insects is serendipitous [4]. In most cases ingestion of its toxin by insect is sufficient for death, though for some insects the presence of some spores is necessary for lethality [5]. Most commercial preparations are mixtures of spores and crystals. Bt is specific, killing only a limited range of insects (even Lepidopteron strains

Table 1: List of entomopathogenic bacteria and insect pest species tested

Bacterial species	Insect species
Bacillus thuringiensis (Mostly kurestaki)	Peach worm, Anarsia lineatella
	Apple worm, Laspeyresia pomonella
	Almond moth, Ephestia cautella
	Dried fruit moth, Ephestia calidella
	Lesser grain borer, Rhizopertha dominica
	Lesser date moth, Batrachedra amydraula
	Potato tuber moth, Phthorimaea operculella
	Wax moth, Galleria mellonella
	Khapra beetle Trogoderma granarium
	Peach green aphid, Myzus persicae
	Cotton leafworm, Spodoptera littoralis
	Beet armyworm, Spodoptera exigua
	Black cutworm, Agrotis ipsilon
	Tomato borer, Tuta absoluta
	Termite, Microcerotermes diversus
	Carob moth, Ectomyelios ceratoniae
	Cabbage worm, Pieris rapae
	Cucurbit fly, Dacus ciliatus
	Cotton bollworm, Heliothis armigera
	Spiny bollworm, Earias insulana
	Cowpea podborer, Etiella zinckenella
	Corn stalk borer, Sesamia cretica
	House mosquito, Culex pipiens, Aedes caspius & Theobaldia longiareolata
	Southern house mosquito, Culex quinquefasciatus
Bacillus sphaericus	House mosquito, Culex pipiens
	Aedes caspius
B. laterosporus	House mosquito, <i>Culex pipiens</i>
	Aedes caspius
Bacillus carotarum	Theobaldia longiareolata
Bacillus cereus	Theobaldia longiareolata

do not kill all Lepidoptera with equal facility) while having no effect on birds, mammals and fish. This selectivity is environmentally advantageous but creates problems in an agricultural setting where a complex of pests has to be controlled. Other drawbacks include slow speed of action, per os activity and U.Vs. sensitivity [6].

In Iraq, researchers were start working with EB, specially Bt in mid 1960s, where [7] reported detailed study (for the first time) about the effect of Bt on spiny bollworm, Earias insulana in lab and field. In the 70s and 80s observations and field trials were also made on the effect of commercial formulations of Bt on peach worm, Anarsia lineatella [8], apple worm, Laspeyresia pomonella [9], Ephestia cautella [10], Galleria mellonella [11]. After that, utilization of Bt to control insect pests species was increased especially during period of 2010 till present time, where the number of published paper reached more than twenty articles dealt with more than ten pest species like tomato borer, Tuta absoluta [12], lesser date moth, Batrachedra amydraula [13]; corn stalk borer, Sesamia cretica [14], spiny bollworm, Earias insulana [15] and potato tuber moth, Phthorimaea operculella [16]. There were few articles dealt with Bacillus species to control mosquito larvae as a potential species transmitting malaria [17, 18]. Table 1 summarize the bacterial species utilized and the insect species tested.

The success cases in this field were as the following:

• The pioneered and comprehensive project concerned with isolation, identification, production, formulation and efficacy of *Bt* local isolates, was initiated at the Directorate of Agricultural and Biological Research, Iraqi Atomic Energy Commission during mid 1980s. The first *Bt* isolate was obtained from infected larvae of *Ephistia* spp. from rearing room and was identified as *Bt var. kaynea* by Pasture Institute, France, which was 1st record from Iraq [19]. Several experiments were conducted to determine the optimal conditions for growth and its efficacy (bioassays) using *Ephistia* larvae as a model and results were compared with that of commercial preparations. In 1991, patent was registered in Iraq about the best fermentation medium

support the growth of different Bt isolates identified up to that time [20]. Field trials of lab scale produced formula indicated that it could protect date fruits at commercial warehouses from infestation by Ephistia spp. larvae for up to six months [21]. Moreover, lab formula efficacy was field tested upon lesser date moth, Batrachedra amydraula [22], Carob moth, Ectomyelios ceratoniae [23] and Cabbage worm, Pieris rapae [24]. The next step was transferred to the of pilot scale production of tow local Bt formulations as wetable powder (WP) using kurstaki variety which showed good efficacies and their potencies were 14500 and 12000 IU/mg for Alnaser II and Alnaser I (brand name of the product respectively [25]. In first half of the 90s and after several large scale field trials conducted in provinces of Sallah-Aldin, Waset, Babel, Dyalh and Baghdad to control corn stem borer, Sesamia cretica at early stage of plant development in cooperation with the State Board of Plant Protection at the Ministry of Agriculture, the product (Alnaser) was adapted by the Ministry of Agriculture to control this insect pest in whole fields of corn cultivation instead of synthetic chemical insecticide usually used at that time. The total amount supplied to the Ministry of Agriculture reached more than 50 tons up to 1996 after that every things stopped according to the UN inspection team.

Large scale field application of the biological insecticide Bt in spray form, at a rate of 3 g/l (6-7 1/tree) to control lesser date moth, Batrachedra amydraula, was investigated [26, 27]. The results indicated that Bt was effective in reducing infestation of lesser date moth. However, the efficiency of the control was varied according to region being 79% in Basra and 35% in Al-Anbar. Good results were also obtained in Babil, Baghda, Diwanya and Wasit provinces. Other previous field trials showed that the efficiency of the biological insecticide Bt was about 66-75 during seasons of 2009 – 2011 [28, 29]. Trees treated with Bt resulted in various yield increase depending on location, cultivar and time of application. In addition, the efficacy of the egg parasitoid, T. evanescens and the bio-pesticide bacteria, Bacillus thuringiensis kurstaki, reduction of lesser date moth, B. amydraula infestations were studied during seasons of 2011 and 2012 and the results were encouraging, thus an IPM project was developed for this purpose at the Ministry of Science and Technology, Directorate of Agricultural Research [30].

Entomopathogenic Fungi: Entomopathogenic fungi are important natural regulators of insect populations and have potential as mycoinsecticide agents against diverse insect pests in agriculture. These fungi infect their hosts by penetrating through the cuticle, gaining access to the hemolymph, producing toxins and grow by utilizing nutrients present in the haemocoel to avoid insect immune responses [31]. Entomopathogenic fungi may be applied in the form of conidia or mycelium which sporulates after application. The use of fungal entomopathogens as alternative to insecticide or combined application of insecticide with fungal entomopathogens could be very useful for insecticide resistant management [32].

Revision of the available published Iraqi literatures on utilizing entomopathogenic fungi indicated that the earliest article was of 1982 measured the efficacy of Beauveria bassiana on long horned date palm stem borer, Jebusea hammerschmiditi (Pseudophilus testaceus) [33]. The peak of activity was during 2000 till now, with total number of 95 published articles. Table 2 shows that the most fungal species studied was Beauveria bassiana followed by Metarhizum anisopliae and other entomopathogenic fungal species.

Different local isolates and few commercial preparations of *Beauveria bassiana* and *Metarhizum anisopliae* efficacies were studied on 40 different insect pests species, such as Mediterranean fruit fly, *Ceratitis capitata* [34], southern cowpea weevil, *Callosobruchus maculates* [35], sunn pest, *Eurygaster integriceps* [36], two spotted mite, *Tetranychus urticae* [37], peach green aphid, *Myzus persicae* [38], corn stem borer, *Sesamia cretica* [39], termite, *Microcerotermes diversus* [40] and dubas bug, *Ommatissus lybicus* [41].

The success cases in this field were as the following:

Two local isolates of *Beauveria bassiana* were isolated from long horned date palm stem borer.

Jebusea hammerschmidti, one (BJH) from Mahaweel area (Babel province, south of Baghdad) and the second (Bb) from date palm orchard soil in Basra province. The efficacy and pathogenesity of both isolates have been tested on different species of-insects and mites, 1-10 days after spore spray. Both isolates showed 100% mortality after 5 days on cotton aphids, Aphis gossypii on cucumber, termites Microcerotermes diversus, scale insects, Aonidella orientalis, grape thrips Retithrips syriacus. The mortality reached 100% for green peach aphid, Myzus persica on potato, parlatoria scale insects, Parlatoria blanchardi on date palm and potato tuber

Table 2: List of entomopathogenic fugi and insect pest species tested

Fungal species	Insect species
Beauveria bassiana	Almond moth, Ephestia cautella,
	Mosquito, Culex quinquefasciatus
	Melon fly, Dacus frontalis,
	Mediterranean fruit fly, Ceratitis capitata
	Poplar leaf beetle, Melasoma populi
	Two spotted mite, Tetranychus urticae,
	Cotton aphid, Aphis gossypii,
	Peach green aphid, Myzus persica,
	White scale insect, Parlatoria blanchardi,
	Oriental yellow scale, Aonidiella orientalis,
	Cotton white fly, Bemisia tabaci,
	Metalic borer beetles Chalcophorella bagdadensis,
	Large stem borer, Capnodis miliaris,
	Termite, Microcerotermes diversus
	Sunn pest, Eurygaster spp.
	Black vine thrips, Retithrips syriacus,
	Angomis grain moth, Sitrotroga cerealella
	German cockroach, Blattela germanica
	Mole cricket, <i>Gryllotalpa</i> sp.,
	Southern cowpea weevil, Callosobruchus maculates
	Lesser date moth, Batrachedra amydraula,
	Fig leaf worm, Ocnerogyia Amanda,
	Citrus mealy bug, <i>Planococcus citri</i> ,
	Onion maggot, Delia alliria,
	Jasmini white fly Aleuroclava jasmine,
	Tuber moth, <i>Phthorimaea operculella</i> ,
	Wax moth, Galleria mellonella,
	House fly, Musca domestica,
	Khapra beetle, Trogoderma granarium,
	long horned date palm stem borer, Jebusea hammerschmiditi
	Corn stem borer, Sesamia cretica,
	Codling moth, Cydia pomonella,
	Cotton leaf worm, Spodoptera littoralis
	Dubas bug, Ommatissus lybicus
	Arabian Rhinoceros beetle, Oryctes agamemnon arabicus
	Fruit stalk borer, Oryctes elegans
	Lesser grain borer, Rhizopertha dominica
	Hyalopterus pruni
	Mealy Plum Aphid, Aphis pomi
	Broad bean black aphid, Aphis fabe
Metarhizium anisopliae	Saw toothed grain beetle, Oryzaephilitis suriuamensis,
	Mediterranean fruit fly, Ceratitis capitata,
	Mole cricket, Gryllotalpa sp.,
	Southern cowpea weevil, Callosobruchus maculates
	Tuber moth, Phthorimaea operculella,
	House fly, Musca domestica,
	Corn stem borer, Sesamia cretica
	Termite, Microcerotermes diversus
	Arabian Rhinoceros beetle, Oryctes agamemnon arabicus
	Fruit stalk borer, Oryctes elegans
	Lesser grain borer, Rhizopertha dominica

Table 2: Continued

Table 2: Continued	
Lecanicillium spp.	Soft scale insect, Exaerotopus tritici,
	Citrus mealy bug, Planococcus citri,
	Jasmini white fly, Aleuroclava jasmine,
	Termite, Microcerotermes diversus,
	White scale insect, Parlatoria blanchardi,
	Dubas bug, Ommatissus lybicus
	Cotton white fly, Bamisia tabaci
	Grain apid, Schizphis graminum
	Black legume aphid, Aphis craccivora
	Broad bean black aphid, Aphis fabe
	Grain English aphid, Sitobion avenae
	Oat aphid, Rhopalosiphum padi,
	Corn aphid, R.maids
Paecilomyces spp.	Poplar leaf beetle, Melasoma populi,
	Sunn pest, Eurygaster integriceps,
	Termite, Microcerotermes diversus
	Cotton white fly, Bamisia tabaci
	Broad bean black aphid, Aphis fabe
Tricoderma spp.	Cabbage aphid, Brevicoryne brassicae
	Grain apid, Schizphis graminum
	Lesser grain borer, Rhizopertha dominica
	Khapra beetle Trogoderma granarium
	Fig leaf worm, Ocnerogyia amanda
	Mole cricket, Gryllotalpa sp.
Isaria spp.	Mealy plum aphid, Hyalopterus pruni
	Apple aphid, Aphis pomi
	Citrus mealybug, Planococcus citri
Doratomyces stemonitis	House fly, Musca domestica
Emericella quadrilineata	House fly, Musca domestica
Zoophthora phytonomi	Alfalfa weevil, Hypera postica
Chaetomium elatum	Southern cowpea weevil, Callosobruchus maculates
Erynia phytonomi	Alfalfa weevil, Hypera postica
Alternaria alternate	Southern apple leaf worm, Targama (streblote) siva
Chaetomium sp.	Southern apple leaf worm, Targama (streblote) siva
спистин эр.	Southern apple real worth, Turgumu (streotole) siva

moth, *Phthorimaea operculella* after 7 days of spore spray and on stone fruit borers, *Chalcophorella bagdadensis* and capnodis, *Capnodis miliaris* larvae and adults after 10 days. The mortality on sunn pests, *Eurygaster integriceps* was 100% after 7 days when sprayed by BJH and after 10 days for Bb however, it was 100% after 7 days on mite, *Tetranychus spp.* for BJH and 10 days for Bb. Several solid and liquid production culture mediums have been tested and found that rice seed culture produced 3.2 x 10⁸ spores /gm while potato sucrose broth and date fruit syrup (debis) cultures produced 5x10⁷ and 3.9x10⁷ spores/ml respectively [42].

The presence of natural endophytic *Beauveria bassiana* within date palm tissues using molecular technique and measure their field efficacies in controlling dubas bug, *Ommatissus lybicus* (Deberg) were investigated recently [43]. Two entomopathogenic *B. bassiana* isolates (MARD 108 and 100) were isolated from

date palm, Phoenix dactylifera L. leaves; in addition, one isolate (MARD 92) originally isolated from soil was identified to have endophytic property. Concentration of 1×10^9 conidia/mL of each of three endophytic isolates was used in field experiments targeting dubas bug nymphs via injection tree trunks. The results indicated that the mortality rates reached 92%, 96% and 100% with infliction of the three endophytic isolates after 15 d from the treatment. The successful establishment of the fungal isolates in the date palm tissue was determined using B. bassiana species-specific primer for the first time via using conventional polymerase chain reaction (PCR) amplification technique before and after injection and the positive gel band representation was the identification signs. The novel results depicted for the first time the presence of natural endophytic B. bassiana isolates within date palm tissues and their field efficacies in controlling dubas bug, O.lybicus (Deberg) infestation.

Entomopathogenic Viruses and Nematodes: First welldocumented introduction of entomopathogenic virus into the environment which resulted in effective suppression of a pest occurred accidentally before the World War II. Along with a parasitoid imported to Canada to suppress spruce sawfly Diprion hercyniae, an nuclear polyhedrosis virus (NPV) specific for spruce sawfly was introduced and since then no control measures have been required against this hymenopteran species. In the past, the application worldwide of baculoviruses for the protection of agricultural annual crops, fruit orchards and forests has not matched their potential. The number of registered pesticides based on baculovirus, though slowly, increases steadily. At present, it exceeds fifty formulations, some of them being the same baculovirus preparations distributed under different trade names in different countries [44]

In Iraq, the published articles in this subject found to be limited (not more than 8 scientific papers). The earliest article was in 1999, which utilized local isolated granulosis virus to control tuber moth, *Phthorimea operculella* under field conditions [45]. Later, NPV was isolated and its efficacy on cotton leaf worm, *Spodoptera littoralis* was investigated [46 - 49]. Another study depicted the isolation of virus belong to family Entomopoxviridae infected long horn palm stem borer, *Jebusaea hammerschmidti* [50]. In addition, Oryctes -like virus was found infested larvae of palm stalk borer, *Orycts elegans* [51].

Nematodes that parasitize insects, known as entomopathogenic nematodes (EPNs), have been described from 23 nematode families [52]. Of all of the nematodes studied for biological control of insects, the Steinernematidae and Heterorhabditidae have received the most attention because they possess many of the attributes of effective biological control agents and have utilized as classical, conservational augmentative biological control agents. The vast majority of applied research has focused on their potential as inundatively applied augmentative biological control agents. Extensive research over the past three decades has demonstrated both their successes and failures for control of insect pests of crops, ornamental and lawn and turf. They can be considered good candidates for integrated pest management and sustainable agriculture due to a variety of attributes: some species can recycle and persist in the environment; they may have direct and/or indirect effects on populations of plant parasitic nematodes and plant pathogens; can play an indirect role in improving soil quality; and are compatible with a wide range of chemical and biological pesticides used in IPM programs [53].

Utilization of EPNs in controlling insect pests in Iraq back to late eighteens of the last century, when commercial preparations (as a gel) were brought from England and tested against onion fly, Delia antiqua and termite, Microcerotermes diversus infested citrus trees. The results were encouraged, hence the shutdown of the facility halted the project (Prof. Abdul-Sattar A. Ali (Al-Anbar Univ., Personal communication). The first documented isolation and identification of EPN Steinernema sp. in Iraq was from long horn palm stem borer and palm stalk borer [54]. This nematode species was utilized through injection of date palm trunk with suspension of nutrient medium and nematode, the result indicated the presence of infected borer larvae with nematode after 3 months [50]. The result of field treatment of olive trees with the nematodes S. carpocapsae and Heterorhabditis bacteriophora, indicated that such treatment inflect high mortality in termite Microcerotermes diversus worker and could protect the trees for 3 months [55]. Steinernema carpocapsae was isolated for the first time from apricot stem borer, Chalcophorella bagdadensis [56]. Laboratory study was conducted to evaluate the efficiencies of entomopathogenic nematode isolated from 13 different sites around Baghdad, using concentrations of 0.0, 25, 50 infective nematode juvenile /5 larvae of stem corn borer Sesamia cretica [57]. Results obtained showed that all nematode concentrations used were efficient in infecting and causing high mortalities of larvae of stem corn.

Moreover, recently a new entomopathogenic nematode Rhabdits blumi (Nematoda: Rhabditida) was isolated from Arabian rhinoceros beetle (ARB), Oryctes agamemnon arabicus larvae [58] . Laboratory results demonstrated that direct spray of 1000 infective juveniles (IJs) per mL of R. blumi on (ARB), caused 71.67% and 15% mortality in the larvae adults respectively. Treating the food source of the larvae (pieces of fresh tissue of the frond bases) with the same dose and period resulted in 48.33% mortality in larvae and 10% in the adults. Field experiments results showed that injection of 50 mL per palm tree with a concentration of 1000 IJs/mL of R. blumi inflected about 42% mortality in ARB larvae infested the tree. Research, nowadays concentrated upon culturing, mass rearing and storing the nematode. Therefore, different culture mediums (lab formulated and commercial products from abroad) were tested and finally reached to choose a lab formulated medium which support efficiently both the reproductive and pathogenic capacities of the nematode. Furthermore, the lab formulated medium could facilitate storing the nematode for around five months without any deleterious effect on nematode pathogenicity (Mohammed Z. Khlaf, Ministry of Science and Technology, Baghdad, Personal communication).

Problems and Challenges Facing Implementing BC Programs in Large Areas in Iraq: In addition to the problems and challenges mentioned before[3], it could be adding the following:

- There are two main areas of safety issues that must be considered when implementing a biological control program. The first question that must be answered is the introduction of the biocontrol agent will have adverse effects on non-target organisms? The second concern is the strength and duration of the biocontrol agent on the environment.
- In order to increase the utility of microbial pathogens in IPM programs, systematic surveys are required in different agroecological zones to identify naturally occurring pathogens. Detailed studies are necessary on the properties, mode-of-action and pathogenicity of such organisms. Ecological studies on the dynamics of diseases in insect populations are necessary because the environmental factors play a significant role in disease outbreaks and ultimate control of the pests.
- Acceptance and implementation of the technology by farmers needs more efforts by specialist and extension personals in order to persuade the farmers about the benefit that they would obtain when using safe alternative. Farmers Field School could be one of the approaches.

Request for Information: It has taken me a lot of time to obtain data on the use of biological control in Iraq and I convinced that this survey is still not complete. I encourage Iraqi researcher to send me all what they have of information, so I can provide a more reliable overview in the future and build up the data base which will be established in the net.

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REFERENCES

- 1. Charn Ley, A.K., 1991. A Review in Microbial pathogens and insect pest control. Letters in Applied Microbiology. 12: 149-157.
- Gupta, S. and A.K. Dikshit, 2010. Biopesticides: An ecofriendly approach for pest control. J. Biopesticides, 3(1): 186-188.
- 3. Hussain F. Alrubeai, 2017. Biological Control of Insect Pests in Iraq: 1) an Overview of Parasitoids and Predators Research Development. Acad. J. Entomol., 10(2): 10-18.
- 4. Martin, P.A.W. and S. Traversr, 1989. Worldwide abundance and distribution of *Bacillus thuringiensis* isolates. Appl. Environ. Microbiol., 55: 2437-2442.
- Paynec, C., 1988. Pathogens for the control of insects: where next? Philosophical Transactions of the Royal Society, London, Series B, 318: 225-248.
- Jutsuma, R., N.J. Poole, A. Powellk and R.L. Bernier, 1989. Insect control using microbial toxins:status and prospects. In Progress and Prospects in Insect Control, ed. McFarlane, N.R. pp: 131-144. London: BCPC
- 7. Al-Azawi, A.F., 1964. Studies on the effect of *Bacillus thuringiensis* on the Spiny bollworm, Earias insulana and other lepidopterous insects. Entomophaga, 9(2): 137-145.
- Kheiri, I.M., T.R. Ahmed and M.K. Abdullah, 1982. Chemical control of *Anarsia lineatella*. Yearbook of plant protection research, Minist. Agri. & Agrar. Reform. Entomology Papers for 1977-1979, 2(1): 127-130.
- Kheiri, I.M. and M.K. Abdullah, 1982. Chemical control of *Laspeyresia pomonella*. Yearbook of plant protection research, Minist. Agri. & Agrar. Reform. Entomology Papers for 1977-1979, 2(1): 131-136.
- Al- Maadhidi, J.F. and H.F. Alrubeai, 1988. Susceptibility of *Ephestia calidella* larvae to *Bacillus thuringiensis* local isolates. Date Palm J. 6(1): 306-312.
- 11. Mahmoud, E.A., A.S.A. Ali and H.E. Abdullah, 1988. Influence *Bacillus thuringiensis* Berliner on survival and development of great wax moth, *Galleria mellonella*. J. Biol. Sci. Res., 19(2): 21-30.

- Ali, J.I.M., S.K. AL-Jamil and N.S. Othman, 2016.
 Isolation and identification of entomopathogenic bacteria companion to tomato moth *Tuta absoluta* Meyrik. J. Tikrit Univ. Agri. Sci., 16(4): 310-315.
- Alrubeai, H.F., B.Sh. Hamad, A.M. Abdullatif, H.Z. Ali and A. Abed, 2014. Efficacy of *Trichogramma evanescens* and *Bacillus thuringiensis kurstaki* to control lesser date moth *Batrachedra amydraula* Merck. J. Agric. Sci. Tech. B., 4: 281-284.
- 14. Al-Zubaidi, A.N., A.S.A. Ali and M.H.H. Al-Seidy, 2008. The combined effect of Agreen and the growth regulators Cascade and Insegar with some chemical insecticides against the corn stalk borer and the egg parasitoid *Telenomus busseolae* Gahan. Iraqi J. Agric., 13(1): 28-37.
- 15. Abdl-Razak, A.S., I.J. Aljboory and A.S.A. Ali, 2007. The effectiveness of the biological insecticide Agreen and some insecticides and growth inhibitors against *Earias insulana* (Lepidoptera: Noctuidae) in cotton field. Iraqi J. Agri. Sci., 38(6): 86-97.
- Al-Azawi, A.A.A., 2006. Study the effect of *Bacillus thuringiensis* to control tuber moth, *Phthorimaea operculella*. MSc. Thesis, College of Sci. for Women, Univ. of Baghdad.
- Mohsen, Z.H., M.A.K. Ibrahim and N.S. Al-Jadooa, 1986. Isolation of spore-forming bacilli from mosquitoes in natural breeding habitats in Iraq. Entomophaga, 31(2): 191-196.
- 18. Hameed, A.A.A., E.H.K. Al-Bander and F.A.G. Al-Obadi, 2017. Comparison the effect of fungi (*Beauveria bassiana*) and bacteria *Bacillus thuringiensis* (*israelensis*) on a cumulative mortality of *Culex Quinquefasciatus* (Diptera: Culicidae). S. Asian J. Life Sci., 5(1): 1-4.
- Al-Maadhidi, J.F. and H.F. Alrubeai, 2000. Production and Utilization of bacterial pesticides in insect control. 1st National Workshop on Biological Control of Insect Pests. IAEC, 25-26 Nov.2000.Baghdad, Iraq.
- Al-Maadhidi, J.F., H.F. Alrubeai, S.H. Khalaf and O.K. Abbass, 1991. Fermentation medium for *Bacillus* thuringiensis. Patent No.2323, Iraqi Central Organization for Standardization and Quality Control.
- Al-Maadhidi, J.F.H.F. Alrubeai, S.H. Khalaf and A.K. Abbas, 1989. Protection of date fruits from Ephestia calidella infestation by Bacillus thuringiensis. Proc. of 3rd National Conf. of Pests and Diseases of Vegetables and Fruits. Egypt., 2: 618-628.
- Alrubeai, H.F., J.F. Al-Maadhidi and L.G. Salim, 1997. Effects of lab. preparations of *Bacillus thuringiensis* on percent infestation of date fruits by *Batrachedra* amydraula. Iraqi. J. Agri. Sci., 28(2).

- Alrubeai, H.F. and J.F. Al-Maadhidi, 1988. Susceptibility of *Ectomyelios ceratoniae* to *Bacillus thringiensis* isolates under laboratory and field conditions. J. Agric. Water Reso. Res., 7(2): 125-136.
- Alrubeai, H.F., Al-Maadhidi, B.H. Hassan and A.H. Salman, 2000. Efficacy of local and commercial preparation of *Bacillus thuringiensis* in controlling cabbage worm *Pieris rapae* L. IAEC. Sci. J., 2: 105-111.
- Alrubeai, H.F., J.F. Al-Maadhidi, Z.A. Al-Gahrbawi and L.G. Salim, 1999. Standardization of two new formulations of *Bacillus thuringiensis* for the use against Lepidopterous pests. Al-Mustansiriyah J. Sci.
- Ali, A.S.A. and N.N. Hama, 2016. Integrated management for major date palm pests in Iraq. Emirate J. Food and Agri., 28(1): 24-33.
- Ali, A.S.A. and J.K. Mohammad, 2014. Potentials of utilizing biological measures for the management of lesser date moth *Batrachedra amydraula* in Iraq, pp: 299-304. In: Proceedings of the 5th International Date Palm Conference. Abu-Dhabi, UAE, March 16-18, 2014.
- 28. Mohammad, J.K., 2011. Some integrated means for the control of the lesser date moth. *Batrachedra amydraula* Meydrick. PhD. Dissertation, College of Agriculture, University of Baghdad.
- Mohammad, J.K., R.F. Al-Jassani, A.A. Ali and M. El-Bouhssini, 2013. The efficacy of the biological insecticides *Bacillus thuringiensis* and Spinosad against the lesser date moth. Iraqi J. Agri. Sci., 44(2): 220-222.
- Alrubeai, H.F., B.Sh. Hamad, A.M. Abdullatif, H.Z. Ali and A. Abed, 2014. Efficacy of Trichogramma evanescens and Bacillus thuringiensis kurstaki to control lesser date moth Batrachedra amydraula Merck. J. Agric. Sci. Tech. B, 4: 281-284.
- 31. Hajeck, A.E. and S.T. Leger, 1994. Interactions between fungal pathogens and insect hosts. Annual Review of Entomology, 39: 293-322.
- 32. Hoy, M.A., 1999. Myths, models and mitigation of resistance to pesticides. In: Insecticide Resistance: From Mechanisms to Management (Denholm, I., Pickett, J.A. and Devonshire, A.L., eds.), New York, CABI Publishing, 1999, pp. 111-119.
- 33. Dhiab, I.M., I.A. Swayir and I. Abdul-Ahad, 1982. Investigations on palm stem borer *Pseudophilus testaceus* (Coleoptera: Cerambycidae). Yearbook of Plant Protection Research, Ministry of Agriculture and Agrarian Reform. 2(1) Entomology papers, 97-112.

- 34. Khlaywi, S.A., M.W. Khudhair, H.F. Alrubeai, A.K. Shbar and S.A. Hadi, 2014. Efficacy of Beauveria bassiana and Metarhizum anisopliae to control Mediterranean fruit fly, Ceratitis capitata. Int. J. Entomol. Res., 02(01): 169-173.
- 35. Salih, H.A.A.A., E.A.M. Al-Ukary and E.A. Al-Bayaty, 2010. Effect of *Beauveria bassiana* on different stages of southern cowpea weevil, *Callosobruchus maculates*. Waste J. Sci. Med., 3(1): 26-33.
- 36. Assaf, L.H.A., 2007. Ecological Study and evaluation of activity of *Beauveria bassiana* (Bals.) Vuill. and *Paecilomyces farinosus* (Dicks ex Fr.) on some biological aspects of sunn pest on wheat. Ph.D. Dissertation, College of Agriculture and Forestry, Mosul University, pp: 231.
- Mahdi, H.M.R., 2002. Chemical and biological control of two spotted mite, *Tetranychus urticae* on tomato crop. MSc. Thesis, College of Agriculture, Univ. of Basrah.
- Al-Jomaily, S.A.R., H.M. Al-Rubeae and N.S. Twaj, 1998. Production of biopesticide from *Beauveria* bassiana (Balsamo) Vuillemin to control green peach aphid Myzus persicae Sulzer. Al-Basrah J. Sci., 4: 44-52.
- Al-Hydari, A.T.A., 2000. Lab and field studies on the effects of *Beauveria bassiana* on corn stem borer *Sesamia cretica*. M.Sc. Thesis, College of Agriculture, Univ. of Baghdad, pp: 66.
- Al-Jassany, R.F. and M.A.A. Al-Salehi, 2011. Efficacy of the fungus *Beauvaria bassiana* (Bais.) Vuil on termite workers and soldiers of *Microcrotermes diversus* (Silvestri) at different temperatures. Arab J. Pl. Prot., 29: 206-213.
- 41. Jassim, H.K., 2007. Biological studies on *Ommatissus lybicus* (DeBergevin) Asche and Wilson (Homoptera: Tropiduchidae) and its biological control using isolates of *Beauveria bassiana* and *Lecanicillium lecanii*. PhD. Dissertation, College of Agriculture, Univ. of Baghdad.
- 42. Al-Jboory, I.J., I.A. Ismail and S.S. Al-Dahwe, 2006. Evaluation of two isolates of *Beauveria bassiana* against some insects and mites and testing the efficiency of some culture media.. Univ. Aden J. Nat. & Appl. Sci., 10(1): 6.
- 43. Khudhair, M.W., H.F. Alrubeai and M.Z. Khalaf, 2016. Innovative method to control dubas bug, *Ommatissus lybicus* (Deberg) (Homoptera: Tropiduchidae) in date palm orchards using endophytic *Beauveria bassiana* isolates. J. Agri. Sci. & Tech. A, 6: 394-402.

- 44. Mazids, J., Ch. Kalita and R. Ch. Rajkhowa, 2011. A review on the use of biopesticides in insect pest management. Int. J. Sci. Advanc. Tech., 1(7): 169-178.
- 45. Salih, H.A.A. and H.F. Hassan, 1999. Isolation and utilization of granulosis virus in controlling of tuber moth, *Phthorimea operculella*. Absracts of the meeting on Biological Control of Agricultural Pests, Aleppo, Syria 1999. Arab J. Pl. Prot., 17(2): 101.
- 46. Rabee, S.K.J., R.F. Al-Jassany and S.H. Samir, 2009. Evaluation the efficiency of some insecticides in controlling cotton leaf worm *Spodoptera littoralis* (Boisd) (Lepidoptera:Noctuidae) on potato (*Solanum tubersum* L.) and survey the natural enemies of the insect. J. Iraqi Agri., 14(3): 153-160.
- 47. Jaafar, S.K.J., R.F. Al-Jassany and S.H. Samir, 2003. Isolation and identification of nuclear polyhydrosis virus from cotton leafworm, *Spodoptera littoralis*. J. Iraqi Agri., 8(3): 65-66.
- 48. Salih, H.A.A.M., L.Kh. Obaid and E.A. Al-Bayyar, 2015. Study the effect of three levels of the virus sponpy as biocontrol roles on the second and fourth instars larvae of the cotton leafworm, *Spodoptera littoralis* in Iraq. J. Al-Anbar Univ. Pure Sci., 9(6): 1-6.
- Salih, H.A.A.M., 2003. Isolation and identification of nuclear polyhydrosis virus from cotton leafworm, *Spodoptera littoralis*. Proc.the 2nd Scientific Conference of Karbala Univ., pp: 806-810.
- 50. Al-Jboory, I.J., 2007. Survey and identification of the biotic factors in the date palm environment and its application for designing IPM-program of date palm pests in Iraq. J.Adan Univ. Nat. Appli. Sci., 11(3): 28.
- 51. Salih, H.A.A.M. and I.J. Al-Jboory, 2001. Isolation and identification of Oryctes -like virus from palm stalk borer, *Oryctes elegans* as first record in Iraq. Al-Basra Date Palm Res. J., 1(2): 114-118.
- 52. Koppenhofer, A.M., 2007. Nematodes. pp: 249-264 in L.A. Lacey and H.K. Kaya, eds. Field Manual of Techniques in Invertebrate Pathology: Application and Evaluation of Pathogens for Control of Insects and Other Invertebrate Pests, Second ed. Dordrecht: Springer.
- 53. Lacey, L.A. and R. Georgis, 2012. Entomopathogenic nematodes for control of insect pests above and below ground with comments on commercial production. J. Nematol., 44(2): 218-225.
- 54. Al-Jboory, I.J. and S.J. Salih, 2001. New record of entomopathogenic nematode on long horn date palm stem borer and palm stalk borer from Iraq. Al-Basra Date Palm Res. J.,1(1): 38-45.

- 55. Al-Jassany, R.F. and M.A.A. Al-Salehi, 2015. Efficacy of some biological agents in protection of olive trees from infestation of termite *Microcerotermes diversus* (Silv.) (Isoptera: Termatidae). Arab J. Pl. Prot., 29(1): 68-76.
- 56. Al-Jboory, I.A. and I.A. Al-Zubaii, 2006. New record of entomopathogenic nematode from Iraq.Arab J. Pl. Prot., 24(1): 56.
- 57. Saleh, H.M., 2015. Detection of entomopathogenic nematodes in agricultural soils in Iraq.Al-Anbar J. Agri. Sci., 13(1): 387-392.
- 58. Khalaf, M., H. Alrubeai, F. Naher and M. Jumaa, 2016. Biological control of the date palm tree borers, *Oryctes* spp.(coleopteran: Scarabaidae:Dynastinae). Book of Proceedings, VII International Scientific Agriculture Symposium (Agrosym 2016) Jahorina, October 06-09, 2016, Bosnia and Herzegovina, pp: 1561-1566.