World Journal of Medical Sciences 8 (4): 387-392, 2013 ISSN 1817-3055 © IDOSI Publications, 2013 DOI: 10.5829/idosi.wjms.2013.8.4.1128

Hypolipidemic and Antioxidant Activities of Anethum Graveolens Against Acetaminophen Induced Liver Damage in Rats

Waffa S.H. Ali

Department of Food Science and Nutrition, Faculty of Home Economics, Helwan University, Cairo, Egypt

Abstract: Twenty eight rats classified into four groups (7 rats each). The first group kept as control (-ve) fed standard diet only. The rest of rats were administered a single dose of 2 g/kg of paracetamol by stomach tube to induce liver injury then classified into control (+ve) and treated groups which were dill powder (10% of dill powder in the diet) and dill extract 300 mg/kg body weight by stomach tube)groups. The experiment period was 45 days. Nutritional results revealed that rat groups which treated with dill powder and dill extract showed a significant increase in weight gain and feed efficiency ratio (FER) comparing to control (+ve) rat group. Biochemical results showed a significant decrease in liver function enzymes and some lipid parameters in serum and also liver malondialdehyde, cholesterol and total lipids. On the other side, they showed increase in high density lipoprotein cholesterol (CHO) in serum and glycogen, triglyceride, glutathione peroxidase and superoxide dismutase in liver comparing to control (+ve) rat group. It is concluded that dill powder and extract can increase the activity of antioxidant and liver function enzymes in paracetamol induced liver injury.

Key words: Anethum graveolens • Antioxidant activity • Feed intake • Feed efficiency ratio (FER) • Liver function enzymes

INTRODUCTION

Liver is a vital organ has a wide range of functions in the body, including biotransformation and detoxification of endogenous and exogenous harmful substances, plasma protein synthesis and glycogen storage. Liver disease is associated with distortion of various metabolic functions. Extensive studies reported that natural products with antioxidant activity are effective to prevent the oxidative stress-related liver pathologies due to particular interactions and synergisms. Acetaminophen (paracetamol) is used as analgesic and antipyretic drug but excessive usage of acetaminophen can damage liver. Liver damage induced by the acetaminophen is a classical model for screening the hepatoprotective activity [1]. Dill belongs to the Umbelliferae family to which belong the herbs bay, cumin and parsley. Its name is derived from the Nordic word 'dilla' pointing to its soothing properties and its botanical name is Anethum graveolens. Although native to the Mediterranean region, dill is cultivated across Europe and America. The plant is 40 to 120 cm tall.

The stem is erect, round, smooth, dark-green and white-striped. The stem is branched above, with a bluish bloom. The leaves are double and more pinnate, feathery, white-tipped leaflets with a deep groove on the upper surface. Leaves have been used as a basic component in canning, soups and sauces and also flavouring salads and seafood while its seeds are used in tea, breads, soups, salads and preserves [2]. One of the most important nutrition benefits of dill leaves is its positive effect on blood sugar levels. Dill contains eugenol, an essential oil which is known to lower blood sugar levels in diabetes [3]. Dill leaves contains monoterpene which helps to protect the body from the harmful effects of free radicals. Dill has an antispasmodic effect on the smooth muscles of the gastrointestinal tract [4, 5]. Moreover, some pharmacological effects of dill have been reported, such as antimicrobial, antihyperlipidaemic, anticancer and antioxidant [6-8].

The present study was designed to investigate the effects of powder and extract of dill on paracetamol induced liver injury in rats.

Corresponding Author: Waffa S.H. Ali, Department of Food Science and Nutrition, Faculty of Home Economics, Helwan University, Cairo, Egypt.

MATERIALS AND METHODS

Materials: Paracetamol drug was obtained from Kahira Pharm. & Chem. Ind. Co. Cairo, Egypt. Kits for biochemical analysis were purchased from the Gamma Trade Company for Pharmaceutical and Chemicals, Dokki, Egypt. Dill was obtained from the local market of Cairo city, Egypt. Twenty eight rats were purchased from Helwan Farm of Laboratory Animals. The initial weight was 180±7g. The standard diet comprised of casein (200 g/kg), corn starch (497 g/kg), sucrose (100 g/kg), cellulose (30 g/kg), corn oil (50 g/kg), mineral mixture (100 g/kg), vitamins mixture (20 g/kg) and DL-methionine (3 g/kg) according to NRC [9].

Methods: Leaves and steams of dill were dried at 60°C, then crushed into powder and added as 10% to the diet as fibers. The hydro alcoholic extract of dried dill was prepared in the food and drug research laboratory of EMRI using following procedure. For the preparation of the hydro alcoholic extract, 100 gram of the dried grounded plant was suspended in 400 mL double distilled water- ethanol (2:1, v/v). The extract was filtered and the filtrate was evaporated to dryness with a rotatory vacuum evaporator. Dill extract was administered at dose levels of 300 mg/kg body weight of rats orally by stomach tube. After five days of adaptation period, the rats were randomly classified into four groups (7 rats each). The first group kept as control (-ve) fed standard diet only. The rest of rats were administered paracetamol drug at a single dose of 2 g/kg by stomach tube to induce liver injury [10]. The liver injured rats were classified into control (+ve) group and the other two treated groups which were dill powder and dill extract groups. Feeding and growth performance were carried out by determination of daily food intake, body weight gain and feed efficiency ratio (FER) according to method of Chapman et al. [11]. The rats were sacrificed at the end of the experiment (45 days) for collection of blood samples which centrifuged at 3000 rpm/15 minutes to obtain serum. The livers of rats were also collected for biochemical analysis and histopathological examination.

Serum aspartate and alanine amino transferase, alkaline phosphatase and gamma glutamyle transferase (AST, ALT, ALP& γ GT) enzymes activity were estimated according to Reitman and Frankel [12], Draper and Hadley [13] and Kind and King [14], respectively. Serum cholesterol (CHO), triglycerides (TG) and high density lipoprotein cholesterol (HDL-_c) were determined by using enzymatic colorimetric methods according to Abell *et al.*

[15], Buccolo and David [16] and Kostener [17], respectively. Low density lipoprotein cholesterol (LDL-c) and very low density lipoprotein cholesterol (VLDL-c) were calculated according to Fruchart [18]. Liver cholesterol (CHO), total lipids, triglyceride and glycogen were determined according to Richmond [19], Folch et al. [20], Scheletter and Nussel [21] and Rerup and Lundquist [22], respectively. Liver glutathione S-transferase (GST), superoxide dismutase (SOD), glutathione peroxidase (GPX) and malondialdehyde (MDA) were estimated according to Ellman [23], Beuchamp and Fridovich [24], Weiss et al. [25] and Uchiyama and Mihara [26], respectively. The fixed samples of liver in 10 % neutral buffered formalin were cleared in xylol and embeded in paraffin. 4-5 µM thick sections were prepared and stained with Hematoxylin and Eosin (H&E) for subsequent histopathological examination [27].

Statistical Analysis: Collected data were statistically analyzed using one way analysis of variance (ANOVA).Student "t" test was used for significance. Differences were considered significant at p < 0.05 according to Artimage and Berry [28].

RESULTS AND DISCUSSION

The results presented in Table 1 revealed that control (+ve) and rat group which treated with dill powder showed a significant decrease in weight gain, food intake and FER at P<0.01,0.001&0.05 while rat group which treated with dill extract showed a non significant decrease comparing to control (-ve) rat group. Rat groups which treated with dill powder and dill extract showed a significant increase in weight gain and FER comparing to control (+ve) rat group. These results are in agreement with those obtained by Singh et al. [6] and Stavri and Gibbons [29], who reported that dill contains substantial amounts of iron, manganese and calcium. It also contains flavonoids which are powerful antioxidants with antiviral and anti-inflammatory properties. Dill also provides zinc, iron, vitamin A and vitamin C which helps to maintain good health. Dill contains essential oils which stimulate the peristaltic movements of the digestive system as well as the secretion of digestive juices and also fiber which aids digestion. Hence, it improves the overall functioning of the digestive system and can be used as an appetizer.

Data in Table 2 showed that control (+ve) showed a highly significant increase in ALT, AST, ALP and γ GT at P<0.001 while rat groups which treated with dill powder and dill extract showed a significant increase in these liver

Table 1: Body weight gain, feed intake and FER of the experimental rat groups					
Groups					
Control (-ve)	Control (+ve)	Dill powder	Dill Extract		
92.41±3.11a	43.61±2.45c***	68.14±4.01b**	85.14±6.36a		
18.61±1.17a	15.67±1.20bc**	16.81±1.10b*	17.84±1.30ab		
0.118±0.003a	$0.061 \pm 0.004 d^{***}$	$0.090 \pm 0.003 c^{**}$	0.106±0.001ab		
	Groups 	Groups Control (-ve) Control (+ve) 92.41±3.11a 43.61±2.45c*** 18.61±1.17a 15.67±1.20bc**	Groups Dill powder 20.41±3.11a 43.61±2.45c*** 68.14±4.01b** 18.61±1.17a 15.67±1.20bc** 16.81±1.10b*		

Significant with control group * P<0.05 ** P<0.01 *** P<0.001.

abed Mean values in each raw having similar letters were not significantly different.

Table 2: Serum ALT, AST and ALP enzymes of the experimental rat groups

Groups			
Control (-ve)	Control (+ve)	Dill powder	Dill Extract
33.71±3.27c	99.61±5.61a***	52.11±4.96b**	49.71±4.22b**
44.20±4.75d	136.31±10.21a***	71.45±7.14b**	68.40±6.10b**
49.61±4.33c	152.13±15.16a***	78.85±7.20b**	73.36±8.11b**
4.96±0.43c	16.31±2.11a***	8.22±1.30b**	7.03±1.11b**
	Control (-ve) 33.71±3.27c 44.20±4.75d 49.61±4.33c	Control (-ve) Control (+ve) 33.71±3.27c 99.61±5.61a*** 44.20±4.75d 136.31±10.21a*** 49.61±4.33c 152.13±15.16a***	Control (-ve) Control (+ve) Dill powder 33.71±3.27c 99.61±5.61a*** 52.11±4.96b** 44.20±4.75d 136.31±10.21a*** 71.45±7.14b** 49.61±4.33c 152.13±15.16a*** 78.85±7.20b**

Significant with control group * P<0.05 ** P<0.01 *** P<0.001.

abed Mean values in each raw having similar letters were not significantly different.

Table 3: Serum lipids parameters of the experimental rat groups

	Groups			
Variables	Control (-ve)	Control (+ve)	Dill powder	Dill Extract
CHO(mg/g)	102.14±14.13c	185.11±17.26a***	125.40±12.12bc	139.61±13.67b*
LDLc(mg/g)	55.68±5.44d	145.26±13.26a***	79.05±8.37c*	93.67±9.16b**
T.G(mg/g)	61.31±7.22b	120.71±12.33a***	70.71±6.99b	73.61±7.14b
VLDLc(mg/g)	12.26±1.35c	24.14±3.44a***	14.14±1.51bc	14.72±1.40bc
HDLc(mg/g)	34.20±3.24a	15.71±1.77c***	32.21±4.31a	31.21±3.27a
Significant with control group * P<0.05 ** P<0.01 *** P<0.001.				

abed Mean values in each raw having similar letters were not significantly different.

Table 4: Liver cholesterol, total lipids, glycogen and triglyceride of the experimental rat groups.

	Groups				
Variables	Control (-ve)	Control (+ve)	Dill powder	Dill Extract	
CHO(mg/g)	4.21±0.38bc	7.33±1.03a***	4.96±0.41b	5.58±0.66b	
T. lipids (mg/g)	35.19±3.11b	55.96±5.17a***	39.61±4.21b	38.41±3.05b	
Glycogen(mg/100g)	6.17±1.07a	3.44±0.55c***	4.70±0.44b*	4.80±0.70b*	
TG (mg/g)	4.11±0.52a	2.99±0.31c***	3.75±0.47ab	3.44±0.32ab	
Significant with control group * P<0.05 ** P<0.01 *** P<0.001					

abed Mean values in each raw having similar letters were not significantly different.

enzymes at P<0.01 comparing to control (-ve) rat group. On the other side, rat groups which treated with dill powder and dill extract showed a significant decrease in these liver enzymes comparing to control (+ve) rat group. The elevated level of AST and ALT confirm in acute liver damage condition. In addition, the level of ALP and γ GT will rise with intrahepatic cholestasis and infiltrative diseases of the liver. The increase in these enzyme activities in our obtained results is indicative for liver damage and thus causes alteration in liver function after exposed to acetaminophen. These results are in agreement with those reported by Larson *et al.* [30]. The levels of these enzyme levels have been decreased by treatment with dill powder and dill extract indicating its hepatic treatment action. This may be due to dill contains vitamins and minerals. It also contains flavonoids which have antioxidant, anti-inflammatory and antimicrobial properties [5, 8].

Data in Table 3 showed that control (+ve) showed a highly significant increase in CHO, LDLc, T.G and VLDLc at P<0.001 and a highly significant decrease in HDLc at P<0.001 comparing to control (-ve) rat group. Rat group which treated with dill powder showed a significant increase in LDLc at P<0.05 and non significant increase in other lipids parameters while rat group which treated with dill extract showed a significant increase in CHO and LDLc at P<0.05 &0.01 comparing to control (-ve) rat group. On the other side, rat groups which treated with dill powder and dill extract showed a significant decrease in CHO, LDLc, T.G and VLDLc and increase in HDLc comparing to control (+ve) rat group. In recent years, applications of dietary plants with antioxidative property have been the center of focus for improving the life quality of patients with hypercholesterolaemia. Yazdanparast and Alavi [31] reported that oral administration of dill leaf extract for 14 days reduced the levels of total cholesterol, triglycerides and low density lipoprotein (LDL) by a significant 20-50%. So, dill has shown promising results towards having a heartprotective function in rats. Valiollah and Naser [32] reported that daily oral administration of Anethum graveolens essential oil to rats at doses of 45, 90 and 180 mg/kg for 2 weeks significantly and in a dose-dependent manner reduced total cholesterol, triglyceride and LDLc and increased significantly HDLc. Anethum graveolens powder when added to the diet of animals showed similar effects on serum lipids.

The obtained results in Table 4 showed that control (+ve) showed a highly significant increase in liver CHO and total lipids at P<0.001 and a highly significant decrease in liver glycogen and liver triglyceride at P<0.001 comparing to control (-ve) rat group. Rat groups which treated with dill powder and dill extract showed a significant decrease in glycogen at P<0.05 comparing to control (-ve) rat group. On the other hand, rat groups which treated with dill powder and dill extract showed a significant decrease in liver CHO and total lipids and significant increase in both liver glycogen and triglyceride comparing to control (+ve) rat group. The obtained result may be due to antioxidant effects of dill. Previous studies have shown that dill's protective ability is comparable to that of alpha-tocopherol, ascorbic acid and quercetin. Protection from free radicals helps prevent serious health problems like cancer, liver and heart disease among

	Groups				
Variables	Control (-ve)	Control (+ve)	Dill powder	Dill Extract	
GST(µ/mg)	10.17±1.71a	6.31±0.86c***	8.96±1.03ab	10.27±1.32a	
GPX(µ/mg)	49.60±4.18a	28.11±2.60c***	39.96±3.98b*	41.14±4.17b*	
SOD(µ/mg)	39.36±4.55a	19.99±1.21d***	29.71±2.14bc*	33.68±3.11ab	
MDA(µ/mg protein)	10.21±1.24c	25.14±5.71a***	15.11±2.17b*	13.67±1.14b*	

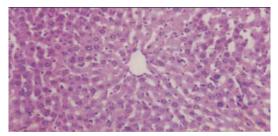
Table 5: Liver GPX, SOD and MDA of the experimental rat groups

Significant with control group * P<0.05 ** P<0.01 *** P<0.001.

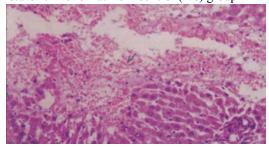
abed Mean values in each raw having similar letters were not significantly different.

others. Fruits and vegetables with bright colors contain properties that protect the body from free radicals. Dill is green in color and it can activate processes in the body which seek out and eliminate free radicals [6, 33].

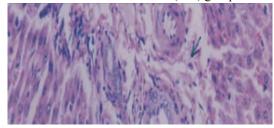
The obtained results in table 5 showed that control (+ve) showed a highly significant decrease in liver GST, GPX and SOD at P<0.001 and a highly significant increase in liver MDA at P<0.001 comparing to control (-ve) rat group. Rat groups which treated with dill powder and dill extract showed a significant decrease in liver GPX and SOD at P<0.05 and a significant increase in liver MDA at P<0.05 comparing to control (-ve) rat group but showed the opposite results comparing to control (+ve) rat group. These results are in line with the findings of Jaeschke et al. [34], who reported that in acetaminophen-induced hepatotoxicity, the balance between ROS generation and antioxidant defense mechanism may be lost thereby results in oxidative stress, which leads to hepatic necrosis. Increased levels of MDA in liver treated with acetaminophen suggest enhanced lipid peroxidation leading to tissue damage and failure to prevent formation of excess free radicals. Bahramikia and Yazdanparast [35] reported that treatment with different fractions of Anethum graveolens extract significantly increased hepatic antioxidant system activities such as SOD, catalase and GSH, along with decreased lipid peroxidation in feeding high-fat diet treated rats. Different fractions of Anethum graveolens extract especially different fractions diethyl ether besides its hypolipidaemic property could protect the liver against the feeding high-fat diet -induced oxidative damage in rats. Yazdanparast and Alavi [31] showed that Anethum graveolens crude extract can either increase the biosynthesis of glutathione or reduce the extent of oxidative stress leading to less glutathione degradation, or it may have both effects. In addition, the increase in glutathione peroxidase activities might be responsible for lowered hepatic MDA content. The elevated levels of both SOD and catalase with Anethum graveolens extract could be due to the influence of flavonoids and polyphenols. Anethum graveolens crude extract possesses in vivo antioxidant activity through decreasing the availability of lipid substances and increasing the activity of antioxidant enzymes.



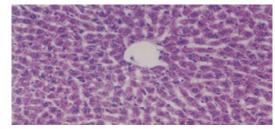
Picture 1: liver of rat from control (-ve) group



Picture 2: liver of rat from control (+ve) group



Picture 3: Liver of rat from dill powder group



Picture 4: Liver of rat from dill extract group

Microscopical examination of liver of rat from control negative group revealed the normal histological structure of hepatic lobule (Picture 1). Conversely, liver of rat from control (+ve) group showed vacuolar degeneration of hepatocytes and focal area of hepatic necrosis replaced by mononuclear leucocytic cells (Picture 2). However, liver of rat from dill powder group showed no changes except kupffer cells activation (Picture 3). No histopathological changes were noticed in liver of rat from dill extract group (Picture 4). Histopathological findings of liver samples were in agreement with the results obtained in biochemical studies, indicating that dill powder and extract are able to inhibit acetaminophen induced hepatotoxicity.

REFERENCES

- Björnsson, E. and R. Olsson, 2006. Suspected drug induced liver fatalities reported to the WHO database. Digestive and Liver Disease, 38(1): 33-38.
- Kmiecik, W., Z. Lisiewska and J. Slupski, 2004. Effects of freezing and storage of frozen products on the content of nitrate, nitrites and oxalates in dill (*Anethum graveolens* L.). Food Chem., 86: 105-111.
- Panda, S., 2008. The effect of *Anethum graveolens* L. (dill) on corticosteroid induced diabetes mellitus: involvement of thyroid hormones. Phytother Res., 22: 1695-1697.
- 4. Fleming, T., 2000. PDR for Herbal Medicines. New Jersey: Medical Economics Company, pp: 252-253.
- Kaur, G.J. and D.S. Arora, 2010. Bioactive potential of *Anethum graveolens, Foeniculum vulgare* and *Trachyspermum ammi* belonging to the family Umbelliferae - Current status. J. Med. Plant Res., 4: 87-94.
- Singh, G., S. Maurya, M.P. Lampasona and C. Catalan, 2005. Chemical constituents, antimicrobial investigations and antioxidative potentials of *Anethum graveolens* L. essential oil and acetone extract: J. Food Sci., 70: 208-215.
- Arora, D.S. and G.J. Kaur, 2007. Antibacterial activity of some Indian medicinal plants. J. Nat. Med., 61: 313-317.
- Kaur, G.J. and D.S. Arora, 2009. Antibacterial and phytochemical screening of *Anethum graveolens*, *Foeniculum vulgare and Trachyspermum ammi*. BMC Complement. Altern. Med., 9: 30.
- NRC (National Research Council), 1995. Nutrient Requirement. Fourth Reviser Edition. pp: 29-30 National Academy Press Washington, Animals, D.C. Environ. Sci. Health, 25: 487-494.
- Rafael, B., F. Daniela, S. Haim, A. Hussein, M. Zipora, P. Moshe, Z. Liliana, A. Yona, O. Ran and H. Zamir, 1999.Hypothyroidism protects rat liver from Aacetaminophen hepatotoxicity. Digestive Diseases and Sciences, 44(6): 1228-1235.
- Chapman, D.G., k R. Gastilla and T.A. Campbell, 1950. Evaluation of protein in food. I. A. Method for the determination of protein efficiency ratio. Can. J. Biochem. Physio., 1(37): 679-686.
- Reitman, S. and S. Frankel, 1957. Determination of glutamate pyruvat transaminase and glutamate oxaloacetate transaminase. Amer. J. Clin. Path., 28: 56-63.
- Draper, H.H. and M. Hadley, 1990. Malondialdehyde determination as index of lipid peroxidation. Methods Enzymol., 186: 421-431.

- Kind, P.R. and E.J. King, 1954. Estimation of alkaline phosphatase activity by determination of hydrolyzed phenol with aminoantipyrene. J. Clin. Path., 7(4): 322-326.
- Abell, L.L., B.B. Levy, B.B. Brodie and R. Kendal, 1952. A simplified method for the estimation of total cholesterol in serum and demonstration of its specificity. J. Biol. Chem., 195: 357-366.
- Buccolo, G. and H. David, 1973. Ouantitative determination of serum triglycerides by use enzymes. Clin. Chem., 19: 419-32.
- Kostener, C.M., 1977. Enzymatic determination of cholesterol high density lipoprotein fraction prepared by polyanion precipitation. J. Clin. Chem., 22: 695.
- Fruchart, G.C., 1982. LDL-cholesterol determination after separation of low density lipoprotein. Rev. Fr. Des. Laboratories, 103: 7-117.
- Richmond, N., 1973. Colorimetric method of determination of total cholesterol and high density lipoprotein cholesterol (HDLc). Clin. Chem., 19: 1350-1356.
- Folch, J., M. lees and G.H. Stanley, 1957. A simple method for isolation and purification of total lipid from animal tissue. J. Biol. Chem., 266: 497-509.
- 21. Scheletter, G. and E. Nussel, 1975. Arbeitsmed Sozialmed Praventimed, 10: 25.
- Rerup, E. and S. Lundquist, 1967. Precipitation and purification of liver glycogen in rats. Acta Pharmmic. Tox., 25: 47-51.
- Ellman, G.L., 1958. Liver glutathione. A colorimetric method for determining low concentration of glutathione. Arch. Biochem. Biophys. 78: 443-450.
- 24. Beuchamp, C. and J. Fridovich, 1971. Superoxide dismutase. Improved assay an assay applicable to acryloamide gels. Anal. Biochem., 44: 276-287.
- Weiss, C., H.S. Marker and G.M. Lehrer, 1980. Sensitive fluorometric assays for glutathione peroxidase and reductase. Anal. Biochem., 106: 512-516.
- Uchiyama, M. and M. Mihara, 1978. Determination of malondialdhyde precursor in tissues by thiobarbituric acid test. Anal. Biochem., 86(1): 271-278.
- Bancroft, J.D.,A. Stevens and D.R. Turner, 1996. Theory and Practice of Histological Technique. 4th Ed. New York, Churchill, Livingstone.
- Artimage, G.Y. and W.G. Berry, 1987. Statistical Methods. 7th Ed. Ames, Iowa State University Press, pp: 39-63.
- 29. Stavri, M. and S. Gibbons, 2005. The antimycobacterial constituents of dill (*Anethum graveolens*). Phytother Res., 19(11): 938-41.

- Larson, A.M., J. Polson, R.J. Fontana, T.J. Davern, E. Lalani and W.M. Lee, 2005. Acute Liver Failure Study Group (ALFSG). Acetaminopheninduced acute liver failure: results of a United States multicenter, prospective study. Hepatology, 42(6): 1364-72.
- Yazdanparast, R. and M. Alavi, 2001. Antihyperlipidaemic and antihypercholesterolaemic effects of *Anethum graveolens* leaves after the removal of furocoumarins. Cytobios. 105(410): 185-91.
- 32. Valiollah, H. and A. Naser, 2008. Hypolipidemic activity of *Anethum graveolens* in rats. Phytotherapy Research, 22(3): 372-375.

- Satyanarayana, S., 2004. The extracts of dill fruits show antioxidant activities in an *in vitro* study. J. Herb. Pharmacother, 4(2): 1-10.
- Jaeschke, H., T.R. Knight and M.L. Bajt, 2003. The role of oxidant stress and reactive nitrogen species in acetaminophen hepatotoxicity. Toxicology Letters, 144(3): 279-288.
- 35. Bahramikia, S. and R. Yazdanparast, 2009. Efficacy of different fractions of *Anethum graveolens* leaves on serum lipoproteins and serum and liver oxidative status in experimentally induced hypercholesterolaemic rat models. Am. J. Chinese Med., 37: 685-699.