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In vitro Activity of Seaweed Extracts Collected from Gulf of Mannar Coast Islands, Tamilnadu on Clinical Isolates

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Abstract: In the present work we used ten seaweeds viz., Ulva lactuca, Ulva reticulate, Halimeda gracilis, Caulerpa peltata, Padina gymnospora, Stoechospermum marginatum, Spatoglossum asperum, Lobosphara variegate, Gracillaria grassa and Acanthophora spicifera were collected from Mandapam, Nallathanni theevu, Muyal theevu and Sayalkudi in the Gulf of Mannar Coast, Rameswaram, Tamil Nadu. Methanol:Chloroform (1:1v/v) was taken as a solvent extraction. The extraction of ten seaweeds were prepared for antibacterial activity against selected human pathogens such as Eschericha coli, Salmonella typhi, Klebsiella pneumonia and Vibrio cholera using agar disc diffusion method with positive control as Tetracycline. All the seaweeds extract were shown moderate antibacterial activity (22 to12mm) out of which maximum zone of inhibition (24mm) was exhibited by Gracilloria grassa against Vibrio cholera. Minimum zone of inhibition (11mm) was exhibited by Caulerpa peltata against Klebsilla pneumonia and (13mm) zone of inhibition by Caulerpa peltata against E. coli. Photochemical analysis of seaweeds may be useful for further antimicrobial activities of seaweeds and to confirm them as a better source for antimicrobial properties in the medical field.

Key words: Seaweeds • Clinical isolates • Zone of inhibition and antibacterial activity

INTRODUCTION

Seaweeds are considered a rich resource of bioactive compounds. Compounds with antibacterial, antiviral, antifungal, antimalarial, antifilarial, hypoglycaemic and antifertility activity have been detected in green (Chlorophyceae), brown (Phaeophyceae) and red algae (Rhodophyceae). They produce primary or secondary metabolites which are potentially bioactive compounds of interest in pharmaceutical industries such as carotenoids, dietary fiber, protein, essential fatty acids, vitamins and minerals. Important polysaccharides such as agar, alginates and carrageenans obtained from seaweeds are used in food Industries. The infectious disease is a major cause of morbidity and mortality worldwide (WHO, 2004). Pharmaceutical industries are interested in marine plants because of their rich and active molecules. This is a major concern and an urgent need for searching new and safe antibacterial agents. Several studies have been investigated about the biological activities of algae extracts. Different active molecules from seaweeds showed the antimicrobial activities against the pathogens e.g., S. aureus or P. aeruginosa that commonly cause infection in the human [1]. Enteric pathogens are the most frequent cause of diarrhea illnesses that account for an annual mortality rate of five million people worldwide, the second most common cause of death after cardiovascular disorders. Prominent pathogenic enteric include *Salmonella*, *Shigella* and strains of *E. coli* are responsible for diarrhea. Many novel molecules have been isolated from marine ecosystem and some of them are under investigation and are being used to developed new pharmaceuticals.

The usage of organic solvents provides a higher efficacy in extracting compounds always for antimicrobial activity assay [2]. Several extractable compounds such as cyclic-polysulfides and halogenated compounds are toxic to microorganism and they are responsible for antibiotic activity of seaweeds. Seasonal and geographical variation also contributes in the antimicrobial activity levels of marine algae. However, information is lacking on the seasonal and geographical variation in the specific metabolites of marine algae of South India. The extraction of major compounds from different species of seaweeds was solvent dependent. There are a lot of reports from around the world related to that seaweed species were extracted using organic solvents. The crude extracts of Indian seaweeds are active only against gram positive

bacteria [3]. To inhibit their activity new drug is required for this purpose sea source is screened for isolation of their novel compound. Photochemical analysis of seaweeds may be useful for further antimicrobial activities of seaweeds and to confirm them as a better source for antimicrobial properties in the medical field. Further work was needed to identify the principle compound which were responsible for antibacterial activity against pathogenic bacteria especially those causing the human diseases.

MATERIALS AND METHODS

Sample Collection: Seaweeds were collected by hands picking in period of November – December 2013 from the Islands for the first time from Nallathanni theevu, Muyal theevu and Sayalkudi along the Gulf of Mannar Coast. [8.47°N 79.02°E] Rameswaram, Tamilnadu, India.

Preparation of Seaweed Extract: The collected seaweeds were cleaned with sea water and fresh water to remove all epiphytes and sand particle and dried in shade at room temperature. The crude extracts from the seaweeds were prepared by solvent extraction method using Chloroform: methanol (1:1v/v) (Iverson *et al.*, 2001) and kept for 28 days, vigorously shaken at intervals. After the extraction the extracted samples were filtered through Waterman no.1 filter paper.

Preparation of Media and Cultures: Media preparation: Peptone – 3g, NaCl-3g, Beef extract -0.18g, Agar- 12g, Distilled water 600ml, P pH adjusted to neutral (7.0). After the sterilization the bacterial culture is inoculate the nutrient broth. The inoculated broth has been incubated for 24 hours 37°C in incubator. The prepared nutrient medium was poured on to the sterilized Petri plates. After the solidification of the medium overnight bacterial cultures were prepared and inoculated by swap method.

Disc Diffusion Method: The crude of extracts of seaweeds such as Green algae (Ulva lactuca, Halimeda gracilis, Ulva reticulate, Caulerpa peltata) Brown algae: (Padina gymnospora, Stoechospermum marginatum, Spatoglossum asperum) and Red algae (Lobophora variegate, Gracillaria grassa and Acanthophora spicifera) were tested against five different human pathogenic bacteria namely Escherichia coli, Klebsiella pneumonia, Vibrio cholera and Salmonella typhi by disc diffusion method. The zone of inhibition was measured (mm in diameter). Chloroform as the negative control, antibiotic Tetracycline as positive control.

RESULT AND DISCUSSION

Marine algae are considered as source of bioactive compounds to produce great variety of secondary metabolites characterized by a broad spectrum of biological activities. In the present study antibacterial activities of 10 different types of seaweeds collected from the Rameshwaram, Mandapam, Nallathanni theevu, Uppu Thanni Theevu, Muyal theevu and Sayalkudi is Gulf of Mannar coast Tamil Nadu. Chloroform: methanol (1:1) extracts of 10 Green, brown and red seaweeds were tested at a concentration of 7.5 μ g/disc diffusion method against 4 clinical isolate of gram negative bacteria namely *Eschericha coli, Salmonella typhi, Klebsiella pneumonia and Vibrio cholera*, with positive control as Tetracycline.

Ulva lactuca: Extract obtained using chloroform: methanol (1:1) pointed out maximum zone of inhibition against pathogens like *Klebsilla pneumoneae* (15mm), *Vibrio cholera* (14mm), *Salmonella typhi* (14mm) and minimum zone of inhibition against *E. coli* (12mm).

Ulva reticulate: The extract obtained using methanol: chloroform (1:1) showed a maximum zone of inhibition against pathogen like *Klebsilla pneumonia* (18mm), *E.coli* (17mm) and minimum zone of inhibition against *salmonella typhi* (13mm), *Vibrio cholera* (13mm).

Caulerpa peltata: The methanol: chloroform (1:1) observed the maximum zone of inhibition against Vibrio cholera (20mm) and minimum zone of inhibition against E. coli (13mm) and Klebsilla pneumonia (11mm). Where no activity against Salmonella typhi.

Halimeda gracilis: The observation made from methanol: chloroform (1:1) showed a maximum zone of inhibition against Klebsilla pneumoneae (18mm), Salmonella typhi (15mm), Vibrio cholera (14mm) and minimum zone of inhibition against E. coli (11mm). Fig. 1-4.

Stoechospermum marginatum: The extract obtained using methanol: chloroform (1:1) showed a maximum zone of inhibition against pathogens like *Vibrio cholera* (22mm) and minimum activity shown against *E. coli* (20mm). Whereas no ant activities against pathogens like *Salmonella typhi and Klebsilla pneumonia* is observed.

Spatoglossum asperum: The methanol: chloroform (1:1) extract obtained showed maximum zone of inhibition against *Vibrio cholera* (16mm) and *Klebsilla pneumonia* (16mm) and minimum zone of inhibition against *Salmonella typhi* (15mm) and *E. coli* (14mm).

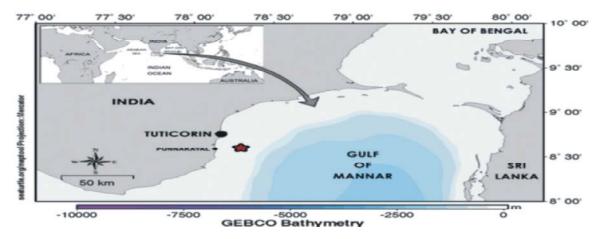


Fig. 1: Localization of the collection site of Gulf of Mannar Coast, Tamilnadu, India

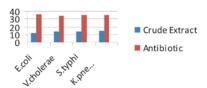


Fig. 1: Zone of Inhibition of crude extract of *Ulva lactuca* against clinical isolates

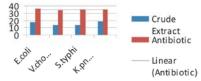


Fig. 2: Zone of Inhibition of crude extract of *Ulva* reticulata against clinical isolates isolates

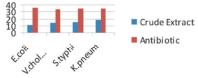


Fig. 3: Zone of Inhibition of crude extract of *Helimedia* gracils against clinical isolates

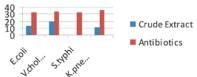


Fig. 4: Zone of Inhibition of crude extract of *Caulerpa* peltata against clinical isolates.

Padina gymnospora: The methanol: chloroform (1:1) extract showed the maximum zone of inhibition against pathogen like *Vibrio cholera* (22mm) and minimum zone of inhibition showed against *E. coli* (20mm). Where as no any activity against pathogens like *Salmonella typhi and Klebsilla pneumonia* were recorded (Fig. 5-7).

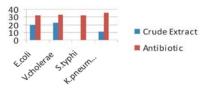


Fig. 5: Zone of Inhibition of crude extract of *Padina* gymnospora against clinical



Fig. 6: Zone of Inhibition of crude extract of Stoechospermum marginatum against clinical isolates

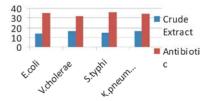


Fig. 7: Zone of Inhibition of crude extract of Spatoglossum asperum against clinical isolates

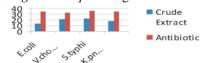


Fig. 8: Zone of Inhibition of crude extract of *Lophophora* variegate against clinical isolates.

Lobophora variegate: The methanol: chloroform (1:1) posses maximum zone of inhibition against *Salmonella typhi* (22mm), *Vibrio cholera* (21mm) and minimum zone of inhibition against *Klebsilla pneumonia* (18mm), *E. coli* (13mm).

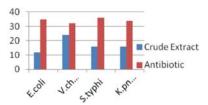


Fig. 9: Zone of Inhibition of crude extract of *Gracilaria* grassa against clinical isolates

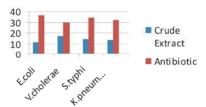


Fig. 10: Zone of Inhibition of crude extract of Achanthophora spicifera against clinical isolates.

Gracillaria grassa: The extraction of methanol: chloroform (1:1) observed maximum activity against Vibrio cholera (24mm) and moderate activity against Salmonella typhi (16mm) and Klebsilla pneumonia (16mm) and less activity obtained against E. coli (12mm).

Acanthophora spicifera: The extract of methanol: chloroform (1:1) showed maximum zone of inhibition against *Vibrio cholera* (17mm) and moderate activity against *Salmonella typhi* (14mm) and *Klebsilla pneumonia* (13mm) and low activity (11mm)obtained against *E. coli* (Fig. 8-10).

In our study, ten different marine algae collected from the Coastal area of Gulf of Mannar. Rameswaram, Tamil Nadu were screened for their antibacterial activity using Methanol: Chloroform extract against four different gram negative human pathogens (E. coli, V. cholera. S. typhi and K. pneumonia) these organisms are considered as danger pathogens because of their fatal causes. According to the past collected data, the different zone of inhibition observed for the same selected test organism with different solvent extract of algae. First time algae is tested for antibacterial activity using methanol: chloroform extract of Lobophora variegate. These algae show antibacterial compound for tested gram negative bacteria showed (18mm) zone of inhibition against the K. pneumonia bacteria, (22mm) zone of inhibition against the S. tvphi bacteria, (21mm) zone of inhibition against the V. cholera, (13mm) zone of inhibition against the E. coli bacteria.

The overall result showed that 1:1 proportion of seaweed extract: antibiotic Tetracycline such as Gracillaria grassa, Lobophora variegata, of red algae and Stoechospermum marginatum, Padina gymnospora of brown algae, Caulerpa peltata of green algae were more active compared to other groups of algae tested. Similar results were also obtained [4, 5]. The tested solvent was determined to be the best solvent for isolation of antimicrobial compounds from the tested marine algae. These results were in close agreement with those obtained by [6]. It was revealed that the chloroform: methanol extract Gracillaria grassa, Padina gymnospora, Stoechospermum marginatum, Lobophora variegate and Ulva lactuca were found active against Vibrio cholera, E. coli, Klebsiella pneumonia, Salmonella typhi. This may indicate that the extraction method had definite effect on the isolation of bioactive principles by using Chloroform: Methanol (1:1 v/v), when compared with other solvents. Some author showed that methanol extraction yield higher antimicrobial activity than n-hexane and ethyl acetate [7, 8]. This difference in results may be firstly due to difference in species used, time and place of sample collection, secondly; there may also be difference in the capability of the extraction protocols to recover the active metabolite and finally, differences in the assay methods that would result in different susceptibilities of the target strains. But, further studies may be made to identify and evaluate the actual substances which are responsible for the antibacterial property.

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