

Response of Some Timber Tree Seedlings Grown in Calcareous Soil to Mycorrhizal and Phosphate Dissolving Bacteria Inoculation

Al-Atrash E.N., H.H. Hammad and Mona A. Amin

Timber Trees Department, Horticulture Research Institute, Agricultural Research Center, Giza, Egypt

Abstract: The present investigation was carried out at the experimental farm of Horticulture Research Station of El-Kanater El-Khayria, Qaluo-beia Governorate, Egypt, along two successive years (30 months starting from March 2018 to August 2020), aimed to study the effects of Vesicular-arbuscular mycorrhiza (VAM) and the phosphate dissolving bacteria *Bacillus megatherium* (B.M.) and *Bacillus polymexa* (B.P.) inoculation on seedlings of *Swietenia macrophylla* King, *Albizia lebeck* (L.) Benth.) and *Dalbergia sissoo* Roxb grown in calcareous soil. Results showed significant increases in all seedling growth parameters (seedling height, stem diameter, root length and fresh weight of shoots and roots) compared with un-inoculated seedlings. Also, growth parameters showed superiority for co-inoculation with VAM+B.M or with B.P compared to other treatments. Inoculation increased total chlorophyll content in leaves and carbohydrate percentages in stems compared to un-inoculated seedlings of all studied tree species. Also, results revealed increments in N, P and K % in leaves of inoculated seedlings with superiority to VAM + B.M and VAM+M.P. On the other hand, the results showed that mycorrhiza and phosphate dissolving bacteria treatments improved soil properties as it decreased pH and EC and increased organic matter (O.M.), beside increasing N, P and K availability forms especially P, which changed from insoluble to soluble form.

Key words: Mycorrhiza • *Bacillus megatherium* • *Bacillus polymexa* • *Swietenia macrophylla* • *Albizia lebeck* • *Dalbergia sissoo* • Calcareous soil

INTRODUCTION

During the last five decades, the reclamation and improvement of new lands in Egypt is an absolute must to face the ever-increasing demand of growing population. One of the most important problems of Egyptian desert soils is high pH level, leading to fix phosphorus in insoluble form such as tri-calcium phosphate. Microorganisms can stimulate, or inhibit root growth, depending on the type of microorganisms, plant species, and environmental conditions [1]. From this point of view Bowen and Rovira [2] classified the soil microorganisms into categories in relation to their effects on plant growth, negatives (detrimental): such as root pathogens, sub clinical pathogens; neutral rhizobacteria, cyanide producers and positives (beneficial): such as rhizobia, mycorrhizae; antagonists (biocontrol) hormone producers, plant growth promoting bacteria. It's well known that the

phosphate dissolving bacterium *Bacillus megatherium* is classified as beneficial, and it is widely used as biofertilizer.

Phosphorus availability in soils is one of the main factors that limiting vegetative growth. Under conditions of limited P, microbes aid in mitigating Puses and increasing its availability and vegetation clearly affects the microbial community and P cycling [3].

Mycorrhizae are obligate fungi that predominate in the roots and soil of higher plants. They form association with plant roots in a host-nonspecific manner. Seven types of mycorrhizae have been known i.e. arbuscular, ecto, arbutoid, ectendo, ericoid, monotropoid and orchidaceous mycorrhizae. Out of these, arbuscular and ectomycorrhizae are the most abundant and wide spread. They promote plant growth by enhancing nutrient acquisition and promoting growth hormones. They also increase the resistance in plants against plant pathogens

and surface area of root system for better absorption of nutrients from soil. Therefore, they can be used as biofertilizer and as biocontrol agent [4, 5].

The average soil phosphorus content is 0.05% (w/w) of which only 0.1% is available to plants due to soil pH [6]. Nearly 80% of applied phosphorus may be unavailable to plants [7]. Global P fertilizer consumption for 2010 was approximately 37.6 Mt with an annual 3% increase in demand thereafter to be over 45 Mt [8, 9]. Reserves of mineable rock phosphate (RP), provides the base raw material for inorganic fertilizer production, [10]. The release of P adsorbed on the solid phase of soil solution is very slow, and consequently, P fertilization is compulsory [11]. The availability of P to crops for uptake and utilization is declining in alkaline and calcareous soils due to the decreases of solubility of calcium phosphate minerals [12, 13].

Under Egyptian soils condition the conversion of applied, inorganic P-fertilizer to precipitated form of CO_3 (PO_4)₂, is a major problem, which is unavailable to the growing plants [14, 15]. VAM symbiosis can promote and increase the uptake of mineral nutrients such as P, Zn, and Cu by plant for growth [16].

On the other side *Bacillus megatherium* produces large amounts of organic acids, which increase the soil acidity and converts the insoluble forms of phosphorus into soluble ones [17, 18]. Consequently, the use of these bacteria as biofertilizers in the alkaline soils is very important and essential to increase the availability of soil phosphorus. Furthermore, the obligate symbiotic microorganisms vesicular- arbuscular mycorrhizal (VAM) fungi are associated with plant roots in a host monospecific manner [19].

Forest trees are renewable resources available to the mankind are not only necessary for ecology and aesthetics but also as a source for obtaining basic necessities for people. Though timber production as of now does not suffice the needs of the ever-growing human population, there is definitely a scope to increase the production of timber. Keeping these needs in view, increasing the production of timber seedlings at nursery level is most essential. This could be possible, if the production can be increased with the application of biofertilizers like phosphate solubilizing bacteria (PSB) and vesicular- arbuscular mycorrhiza (VAM), the mycobiont undergoes pronounced alterations of root system besides ensuring ecological sustainability [20]. VAM fungi offer a great potential for sustainable plant growth [21]. Recently, the potential of PSB and VAM fungal association on different plants has been well documented [22, 23].

Swietenia macrophylla, belongs to family Meliaceae otherwise, known as Big Leaf Mahogany, is a slow-growing, tall, tropical tree reaching a height of about 40-60 m with a diameter of 1.5 to 2 m [24]. Various medicinal uses of this plant have been reported. The bark is used to treat diarrhea and fever. The tree is not cultivated for food for no plant part is edible. Else, it is used in reforestation projects or as a shade tree in plantation crops. Crushed fruit shells are used as a potting medium. The bark produces gums, and used for dyeing and tanning leather. Seed kernels yield oil which is very bitter and purgative. The wood is valued for high quality woodwork and furniture, musical instruments, veneer, etc. [24, 25].

Albizia lebeck (L.) Benth belongs to family Fabaceae, it is a multipurpose tree for semiarid regions. *A. lebeck* has been widely distributed around the tropics and mainly planted as a shade tree [26]. It is used in medicine in different purposes. Its wood is dense and used for making cabinet timber and also some types of furniture.

Dalbergia sissoo, belongs to family Fabaceae, and is one of the most useful timber species. It is used for high-quality furniture, cabinets, decorative, veneers, marine and aircraft grade plywood, ornamental turnery, carving, engraving, tool handle and sporting goods. Its root wood is used for tobacco pipes. In village industry, *D. sissoo* is popular for doors and windows. Oil obtained from the seeds is used to cure skin diseases. It is used as a wind break in mango, coffee and tea plantation. The tree nodulates, it therefore improves soil fertility [27].

Using mineral P along with slow release P fertilizer (rock phosphate) plus P bacterial transformants namely *Bacillus megatherium* var. Phosphaticum (phosphorene and mycorrhizal) was very effective in stimulating growth aspects and vine nutritional status as well as improving yield and quality of grapevine crops [28-31]. The solubility of P in triple calcium super-phosphate and slow release P fertilizer added to the soil can be governed by soil type, pH, organic matter and biofertilization. Using different organic manures enriched with B.M. var. phosphaticum as well as the application of mycorrhiza substantially was associated with enhancing the availability of P to plants [32, 33].

This study focused on the response of *Swietenia macrophylla* King, *Albizia lebeck* (L.) Benth.) and *Dalbergia sissoo* Roxb seedlings, grown in calcareous soil, to mycorrhiza and phosphate dissolving bacteria inoculation.

MATERIALS AND METHODS

The present investigation was carried out at the experimental farm of Horticultural Research Station of El-Kanater El-Khayria, Qaluoobia Governorate, Egypt, along two successive years (30 months starting from March 2018 to August 2020) to study the effects of Mycorrhiza (VAM) and the phosphate dissolving bacteria *Bacillus megatherium* (B.M.) and *Bacillus polymexa* (B.P) inoculation on seedlings of *Swietenia macrophylla* King, *Albizia lebbek* (L.) Benth. and *Dalbergia sissoo* Roxb grown in calcareous soil.

Plant Materials: One-year-old seedlings of *Swietenia macrophylla* king., *Albizia lebbek* (L.) Benth. and *Dalbergia sissoo* Roxb., were chosen. The seedlings used were in average of 15, 20 and 18 cm., in height and 0.7, 0.5 and 0.4 cm., in diameter respectively. The seedlings were transplanted on the first week of March 2018 in polyethylene bags of 20 cm. diameter, and 35 cm. depth, filled with 8 kg of calcareous soil transported from Bilpies desert, Sharkia Governorate, Egypt.

Physical and Chemical properties of the used soil were determined according to Black *et al.* [34] as shown in Table (1).

The seedlings were placed in shaded area and common cultural practices including irrigation were followed (twice weekly in winter and four times in summer). Every transplant received starter nutrition consisted of NPK 0.5 gm from Kristalon 19: 19: 19 for bacterial activity.

Microbial Inoculum: Mycorrhiza inoculum of *Glomus marrocarpum* fungi was supplied by the Microbiology Resources Centre (MIRCEN), Faculty of Agriculture, Ain Shams University, Cairo, Egypt. While, *Bacillus megatherium* and *Bacillus polymexa* were supplied by the Microbiology Department, Soil, Water and Environment Research Institute, Agricultural Research Center, Giza. *Bacillus* strains were grown in KB medium

(King's B Medium) and incubated at 25°C for three days until early log phase was developed and cell density reached about 10^9 ml⁻¹ container medium.

Inoculation of Seedlings: VMA was inoculated according to Menge *et al.* [35]. The gridlines intersect method was used to calculate VAM infection% according to Giovannetti and Mosse [36] which reached 90 %. While, the seedlings received suspensions of bacteria *Bacillus megatherium* (B.M.) and *Bacillus polymexa* (B.P.) three times on April 2018, 2019 and 2020 from planting as a soil drench at the rate of 10 ml per bag. The control plants were left without any biofertilization.

Data Concerned: At the end of the experiment) August 2020), the vegetative growth parameters were recorded including seedling height, stem diameter, root length (cm), root and shoot fresh weights (g), as well as shoot/root ratio on fresh weight basis.

Chemical Composition:

- Total chlorophylls, was determined according to the method described by A.O.A.C [37].
- Carbohydrates (% D.W) in stem was estimated using the method recommended by Dubois *et al.* [38].
- Nitrogen content was determined using the modified micro Kjeldahl method as described by Pregl [39].
- Phosphorus content was estimated using the method recommended by King [40].
- Potassium content was estimated using the flame spectrophotometer method of Piper [41].

Layout of Experiment: A number of 150 seedlings for each tree species was selected for this study then divided to 6 treatments, each treatment included 25 seedlings in 5 replicates (5 seedlings/replicate), which were arranged in a complete randomized design. All the obtained data were subjected to analysis of variance according to Snedecor and Cochran [42]. The new (LSD) was used to compare the average of the determined parameters.

Table 1: Soil physical and chemical analyses

pH	EC (dsm ⁻¹)	Calcium carbonate%	Coarse sand%	Fine sand%	Clay%	Silt%	Textural grade				
8.19	6.45	1.7	19.88	56.70	4.12	19.30	Calcareous				
							Available nutrients				
Water soluble cation (m mole L ⁻¹)				Water soluble anion (m mole L ⁻¹)				-----			
Ca ⁺⁺	Mg ⁺	Na ⁺	K ⁺	Co ₃ ⁻	HCO ₃ ⁻	Cl ⁻	SO ₄ ⁻	Available N (ppm)	Available P (ppm)	Available K (ppm)	S.P.
22.31	9.11	30.48	2.00	0.0	2.34	42.0	19.56	35.85	6.45	185.15	17.00%

RESULTS AND DISCUSSION

Vegetative Growth: Data in Tables (2) showed that inoculation with Vesicular- arbuscular mycorrhiza (VAM) or phosphate dissolving bacteria *Bacillus megatherium* (B.M.) or *Bacillus polymexa* (B.P.) to growing media had generally favorable effects on the vegetative growth parameters, compared to noninoculated seedlings (control). Also, results showed that the combined treatments of VAM with phosphate dissolving bacteria had significant effect on growth parameters, for all plant species.

Concerning the effect on seedling heights, the tallest plants were obtained with VAM + B.P. giving (104.00 and 95.00 cm.) for *Swietenia macrophylla* and *Albizia lebbek* seedlings with percentages of increases by 48.6 and 50.8% over control, respectively, while *Dalbergia sissoo* seedlings recorded the highest value when seedlings were inoculated by VAM+B.M (117.0 cm) achieving increase by 51.95% over control.

For stem diameter, inoculation with VAM + phosphate dissolving bacteria gave the thickest stems compared to control. The mixture of VAM + B.M. gave the highest value with *Swietenia macrophylla*, (2.60 cm) recording increase by 63% over control, while the mixture of VAM + B.P. recorded the highest values of stem diameter (2.40 and 1.70cm) for *Albizia lebbek* and *Dalbergia sissoo* seedlings with 118.18 and 112.5% increases over control respectively.

Regarding root length, data showed that inoculation of VAM or phosphate dissolving bacteria positively affected the root length for seedlings of all species comparing with control, but the highly significant values were obtained with VAM+B.P. recording (36.00, 33.00 and 33.00 cm) with 63.64, 83.33 and 94.12 % increases over control for *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* seedling respectively.

From the previous results it's worthy to notice that applying the mixture of VAM+ phosphate dissolving bacteria as soil drench biofertilizers produced healthy seedlings.

Data in Table (3) indicated highly significant increases in fresh weights of shoots and roots of seedlings inoculated by VAM and bacterial strains either alone or as a mixture. The highest increments of fresh weights of roots and shoots were observed with using the mixture of VAM + B.M., (28.00, 19.00, 16.00, 42.00, 36.00 and 41.00 g.) for *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* seedlings, respectively. It is

observed that the increments in fresh weights of roots represented 133.3, 116.7 and 75 % over control in the plants treated with VAM+B.M, VAM+B.P and VAM alone, respectively in *Swietenia macrophylla* during extended growing season (30 months), the same line was observed with *Albizia lebbek* and *Dalbergia sissoo* seedlings.

Also, data in Table (3) showed that the highest significant values in fresh weight of shoot of *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* seedlings were recorded by adding both VAM+B.M or VAM+ B.P compared with all used treatments. On the other hand, the lowest ones were obtained by control treatments.

Concerning shoot/roots fresh weight ratio of studied seedlings, data in Table (3) showed that this ratio in *Swietenia macrophylla* seedlings significantly reduced with inoculation by microorganism's strains, (VAM + B.M or VAM + B.P) compared with control consequently, the reduction in shoot/roots ratio F.W. was 9.87% than control, which mean that the roots increased in inoculated seedlings compared to non- inoculated ones. On the other hand, shoot/roots ratios of *Albizia lebbek* and *Dalbergia sissoo* seedlings exhibited that control gave the lowest values of 1.51 and 1.22. So the results of *Swietenia macrophylla* and *Albizia lebbek* more affected by inoculation with microorganism's strains as a mixture or alone, while *Dalbergia sissoo* seedlings were more affected by the inoculation with mycorrhiza fungi (VAM).

Chemical Analyses:

Total Chlorophylls (mg/g F.W.) and Total Carbohydrates (%D.W.): Regarding total chlorophylls, data in Tables (4) indicate that there were significant increases in leaves content of chlorophyll in seedlings inoculated with VAM+B.P. giving (4.91, 5.68 and 5.83 mg/ g F.W.), representing 22.44, 41.65, 41.85% increases over controls for *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo*, respectively. Statistically, all other treatments showed insignificant differences in the chlorophyll contents.

Concerning the effects of micro-organism inoculation on the content of total carbohydrates (%) of *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* seedlings, the obtained values ranged from 22.11, 10.79 and 9.42% in controls recording increases by (24.07% with B.P., 12.14% with VAM and 9.61% with B.M.) for the studied seedlings respectively.

Table 2: Effect of mycorrhiza and phosphate dissolving bacteria treatments on increment of seedling height, stem diameter and root length at the end of the experiment

Treatments	<i>Swietenia macrophylla</i>			<i>Albizia lebbeck</i>			<i>Dalbergia sissoo</i>		
	Seedling height (cm)	Stem diameter (mm)	Root length (cm)	Seedling height (cm)	Stem diameter (mm)	Root length (cm)	Seedling height (cm)	Stem diameter (mm)	Root length (cm)
Control	70.00	1.60	22.00	63.00	1.10	18.00	77.00	0.80	17.00
VAM	83.00	1.90	33.00	72.00	1.40	23.00	85.00	1.00	25.00
B.M	80.00	2.30	26.00	70.00	1.70	27.00	90.00	1.01	22.00
B.P	81.00	2.02	27.00	74.00	1.90	29.00	93.00	1.06	27.00
VAM+B.M.	102.00	2.60	34.00	93.00	2.20	31.00	117.00	1.06	31.00
VAM + B.P.	104.00	2.50	36.00	95.00	2.40	33.00	110.00	1.70	33.00
LSD at 5 %	0.56	0.08	0.65	0.58	0.09	0.67	0.59	0.09	0.66

VAM = Vesicular Arbuscular Mycorrhiza B.M. = *Bacillus megatherium* B.P.=*Bacillus polymexa*

Table 3: Effect of mycorrhiza and phosphate dissolving bacteria treatments on, fresh weights of roots and shoots and shoot / root ratios at the end of the experiment

Treatments	<i>Swietenia macrophylla</i>			<i>Albizia lebbeck</i>			<i>Dalbergia sissoo</i>		
	Root fresh weight (g)	Shoots fresh weight (g)	Shoot/roots ratio fresh weight (g)	Root fresh weight (g)	Shoots fresh weight (g)	Shoot/roots ratio fresh weight (g)	Root fresh Weight (g)	Shoots fresh weight (g)	Shoot/roots ratio fresh weight (g)
Control	12.00	21.00	1.75	10.00	15.00	1.51	9.00	11.00	1.22
VAM	21.00	34.00	1.62	12.00	24.00	2.02	12.00	19.00	1.59
B.M	15.00	28.00	1.89	11.00	29.00	2.64	10.00	26.00	2.64
B.P	16.00	29.00	1.82	16.00	31.00	1.95	14.00	27.00	1.96
VAM+B.M.	28.00	42.00	1.58	19.00	36.00	1.59	16.00	41.00	2.06
VAM+B.P.	26.00	41.00	1.58	19.00	34.00	1.79	15.00	33.00	2.76
LSD at 5 %	1.07	0.72	0.08	1.01	0.77	0.10	0.99	0.71	0.11

VAM = Vesicular Arbuscular Mycorrhiza B.M. = *Bacillus megatherium* B.P.=*Bacillus polymexa*

Table 4: Effect of mycorrhiza and phosphate dissolving bacteria treatments on total chlorophyll in leaves (mg/g F.W.) and total carbohydrates contents in stem at the end of the experiment

Treatment	<i>Swietenia macrophylla</i>		<i>Albizia lebbeck</i>		<i>Dalbergia sissoo</i>	
	Total chlorophyll (mg/g.F.W.)	Total carbohydrates (%D.W.) in stem	Total chlorophyll (mg/g.F.W.)	Total carbohydrates (%D.W.) in stem	Total chlorophyll (mg/g.f.w.)	Total carbohydrates (%D.W.) in stem
Control	4.01	22.11	4.01	10.79	4.11	9.42
VAM	4.81	24.01	4.37	12.14	4.22	9.33
B.M	4.62	23.91	4.81	11.75	4.43	9.61
B.P	4.50	24.07	4.87	12.03	4.48	9.58
VAM+B.M.	4.73	20.76	5.30	11.60	5.40	8.50
VAM+B.P.	4.91	21.40	5.68	11.77	5.83	8.40
LSD at 5 %	0.13	0.16	0.14	0.14	0.12	0.15

VAM = Vesicular Arbuscular Mycorrhiza B.M. = *Bacillus megatherium* B.P.=*Bacillus polymexa*

Table 5: Effect of mycorrhiza and phosphate dissolving bacteria treatments on N, P and K percentages in leaves at the end of the experiment

Treatments	<i>Swietenia macrophylla</i>			<i>Albizia lebbeck</i>			<i>Dalbergia sissoo</i>		
	N (%)	P (%)	K (%)	N (%)	P (%)	K (%)	N (%)	P (%)	K (%)
Control	2.01	0.22	0.91	2.12	0.18	1.21	1.27	0.26	0.66
VAM	2.55	0.35	0.96	2.30	0.26	1.23	1.38	0.52	0.66
B.M	2.11	0.33	1.01	2.19	0.20	1.30	1.40	0.50	0.68
B.P	2.42	0.35	1.02	2.22	0.23	1.32	1.40	0.44	0.67
VAM+B.M.	2.50	0.36	1.04	2.73	0.25	1.36	1.41	0.60	0.69
VAM+B.P.	2.47	0.37	1.08	2.55	0.27	1.40	1.41	0.61	0.69
LSD at 5 %	0.11	0.02	0.02	0.11	0.02	0.03	0.10	0.02	0.02

VAM = Vesicular Arbuscular Mycorrhiza B.M. = *Bacillus megatherium* B.P.=*Bacillus polymexa*

Nitrogen, Phosphorus and Potassium (%D.W.): Data in Table (5) revealed that the application of mycorrhiza and phosphate dissolving bacteria treatments caused significant elevation of N% compared to control in all studied species, and the highest N % values occurred in the seedlings treated with VAM for *Swietenia macrophylla* 2.55% in leaves whilst, the highest values of N% for *Albizia lebbek* and *Dalbergia sissoo* seedlings were 2.73 and 1.41% in leaves, for the treatment of VAM+B.M compared to their control.

Data presented in Table (5) showed that P% in leaves of *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* significantly increased by different inoculation treatments compared to control. The best results were obtained with the treatments of VAM + B.P. which gave 0.37,0.27 and 0.61 P%, followed by those treated with VAM+B.M 0.36, 0.25 and 0.60 P% of seedling of *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* respectively.

Results in Table (5) showed that K% in leaves of *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* seedlings increased under all inoculation treatments compared to controls of these species which recorded the lowest values of 0.91,1.21 and 0.66 K% respectively. This increase was significant only with the treatment of VAM+B.P which reached to maximum values 1.08, 1.40 and 0.69 K% followed by those treated with VAM+B.M. giving 1.04,1.36 and 0.69 K% respectively.

Effect of Mycorrhiza and Phosphate Dissolving Bacteria Treatments on Soil Properties: Results obtained in Table (6) revealed that, the inoculation by mycorrhiza and phosphate dissolving bacteria had beneficial influences on soil properties.

As regard soil pH, inoculation decreased it with all treatments and the lowest pH was obtained with VAM+B.P recording decrease percentages of 5.11, 0.85, 0.97% compared to control.

For EC values, also the VAM+B.P inoculation recorded the lowest percentages (1.80, 1.66, 1.65%) over the untreated soil for *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo*, respectively.

Concerning soil organic matter (O.M), the data in Table (6) indicated positive effects in increasing the percentage of (O.M) to reach 15.38,15.09, 11.76% for VAM+ B.P inoculation over the control for *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo*, respectively.

Generally, VAM, B.M and B.P. play the main role in facilitating the presence of phosphorus in the soil in dissolving form through the material it produces, that reduce the acidity of the soil, which brings the triphosphate to a unique phosphate that can be absorbed by plant roots.

Concerning the effect of mycorrhiza and phosphate dissolving bacteria inoculation on N.P and K contents of the soil, results showed increments with all treatments and the high superiority of the values were obtained with VAM+B.P recording 7.69, 8.26 and 8.34 % for N, 31.21,30.07 and 39.52% for P and 22.73, 23.12 and 20.87% for K over control for *Swieteniamacrophylla*, *Albizia lebbek* and *Dalbergia sissoo*, respectively. These results agree with Mahfouz and Sharaf- Eldin [43]; Hasaneen *et al.* [44]; Runyan and Odorico [3]; Sushanta *et al.* [11] and Ghafoor [13].

It is worthy to mention that the results of the present study indicated that the inoculation of the studied seedlings with VAM and Phosphate dissolving bacteria (PDB) exhibited positive effects on growth parameters of the inoculated plant seedlings. This was previously reported by Mahfouz and Sharaf-Eldin [43] on (*Foeniculum vulgare* Mill.); Hasaneen *et al.* [44] on *Lactuca sativa*; Ahmed and Abada [29]; Shaheen *et al.* [30] and Shaaban [31] on superior grapevine.

Also, it's evident that microorganism's strains, mixture or alone was beneficial for the growth of both roots and shoots also, biofertilization promoted and formed more secondary hairs roots. These results agreed with Motosugi *et al.* [45] on some annual plants as they reported that the highest ratio of roots / shoots and leaves was given by adding biofertilizers and its evident that resulted in more formation of hair roots.

Concerning the effect of biofertilizer on enhancing the content of total chlorophylls and total carbohydrates, the obtained results were in harmony with Shanan and Higazy [46] who found that algalization enhanced the biochemical characters such as carbohydrates and chlorophyll in leaves. Also, Grzesik *et al.* [47] on (*Salix viminalis* L.) reported that the biofertilizers treatments increased the stability of chlorophyll content, intensity and of net photosynthesis.

Prasanna *et al.* [48] on wheat (*Triticum vulgar* L.) and Anand *et al.* [49] on maize crop showed significant increase in chlorophyll a, and Mohsen *et al.* [50] on lettuce plants. The authors found that all different biofertilizer treatments considerably increased chlorophyll content and total carbohydrate.

Table 6: Effect of mycorrhiza and phosphate dissolving bacteria treatments on soil properties at the end of the experiment

<i>Swietenia macrophylla</i>						
Treatment	pH	EC(dsm ⁻¹)	O.M. (%)	N ppm	P ppm	K ppm
Control	8.60	6.65	0.52	35.75	7.05	179.9
VAM	8.18	6.55	0.59	36.93	9.11	216.0
B.M.	8.20	6.56	0.59	36.77	8.33	212.2
B.P.	8.19	6.56	0.59	36.80	8.35	212.2
VAM+B.M.	8.23	6.53	0.60	38.46	9.26	220.2
VAM+B.P.	8.16	6.53	0.60	38.50	9.25	220.8
<i>Albizzia lebbbeck</i>						
Control	8.22	6.64	0.53	35.84	7.15	179.5
VAM	8.17	6.58	0.54	38.11	9.58	215.0
B.M.	8.19	6.59	0.54	37.70	8.46	216.3
B.P.	8.19	6.59	0.58	37.20	8.48	216.0
VAM+B.M.	8.16	6.53	0.58	38.80	9.26	220.6
VAM+B.P.	8.15	6.53	0.61	38.80	9.30	221.0
<i>Dalbergia sissoo</i>						
Control	8.23	6.66	0.51	35.87	7.06	179.2
VAM	8.17	6.59	0.54	34.40	9.80	215.1
B.M.	8.20	6.60	0.54	36.80	8.41	214.0
B.P.	8.19	6.60	0.54	36.78	8.35	214.60
VAM+B.M.	8.17	6.58	0.57	38.80	9.99	216.0
VAN+B.P.	8.15	6.55	0.57	38.86	9.85	216.6

VAM = Vesicular Arbuscular Mycorrhiza B.M. = *Bacillus megatherium* B.P.=*Bacillus polymixa*

These results are in agreement with Hammad *et al.* [18] on *Taxodium disticum*; Nelson and Achar [19] on *Brassica oleracea*, Al Harbi *et al.* [12] and Ghafoor [13] on wheat, who explained that improving effects arising from microbial inoculation are due to dissolving phosphate to be available form by plants, and producing growth promoting substance such as auxins.

Many results are inconclusive, but encouraging enough to improve selection procedures and the production of quality inocula for practical application.

Despite only a small proportion of angiospermic species having been examined, mycorrhiza form a ritualistic of relationship with the roots of nearly eighty percent of such plant species [5]. Arbuscular mycorrhiza (AM) symbiosis can promote host plant growth by increasing the uptake of mineral nutrition such as P, Zn, and Cu [16]. AM Fungi and plant roots, improve water and nutrient uptake like phosphorus, nitrogen and micronutrients and thus enhance plant growth [51].

As mycorrhizal inoculation mediated processes involved the availability of phosphate and other nutrients in the soil. Arbuscula rmycorrhizal (AM) fungi are found among the soil flora and interact with approximately 85% of the plants on the ground [52].

Mycorrhizae are obligate fungi that predominate in the roots and soil of higher plants. They form association with plant roots in a host-nonspecific manner. They also

increase the resistance in plants against plant pathogens and surface area of root system for better absorption of nutrients from soil. Therefore, they can be used as bio-fertilizer and as biocontrol agents. The primary establishment effect of AM is the improvement of phosphate uptake by plants due to the ability of the external mycelium of AM fungi to act as a bridge between roots and the surrounding soil microhabitats. This gives access to the phosphate ions from the soil solution beyond the phosphate- depletion zone surrounding the roots [53]. The AM fungi can contribute to P capture and supply, by linking the biotic and geochemical portions of the soil ecosystem, therefore affecting P cycling rates and patterns in both agricultural and natural ecosystems [54]. Phosphate dissolving bacteria (PDB) solubilize insoluble P by producing chelating substances and various organic acids and hence this available P is taken up by plants [55-57]. The hyphae length density of plants inoculated with PDB and AMF was longer than the other plants inoculated with AMF alone and there was significant difference between them. That is could be attributed to the role of PDB which increased the total percentage of root colonization by native AMF by producing phytohormones which apparently stimulate mycorrhizal infection, the phosphate made available by PSB (phosphate solubilizing bacteria) acting on sparingly soluble P sources may not reach the root surface due to

limited diffusion, it was proposed that if the solubilized phosphate was taken up by an AM mycelium. This synergistic microbial interaction should improve P supply to the plant [58].

In the present investigation, results revealed that the combined inoculation of VAM+ B.P. or B.M. significantly increased seedling height, over the treatments of PSB or VAM. These results are consistent with earlier reports of Kalavathi *et al.* [59]. It is relevant to mention that the possible synergistic effect would be the uptake by AM fungal hyphae and translocation into the plant of P released by PSB in soil [21, 23].

Certain plant hormones like IAA and GA produced by the bacterial culture might induce the growth of other associated organisms like VAM [58].

Thus, using of these biofertilizers treatments VAM +B.P. or B.M. produced vigour growth of *Swietenia macrophylla*, *Albizia lebbek* and *Dalbergia sissoo* seedlings as well as healthy timber seedling stock at nursery level, for forest plantation projects.

CONCLUSION

It is necessary to mention that the use of biofertilizers is important for cheap production and, also to provide hard currency for the import of mineral fertilizers, in addition to improve the soil and reducing air pollution resulting from the oxidation of mineral nutrients, in addition to the utilization of non-absorbable phosphorus. In order to get the most out of it, you must combine various microorganisms that are useful for feeding the plant. From the results of our study it's worthy to be recommended that applying VAM+ phosphate dissolving bacteria to the plants as bio fertilizers for the production of healthy seedlings of *Swietenia macrophylla* *Albizia lebbek* and *Dalbergia sissoo* seedlings especially when grown in calcareous soil.

REFERENCES

1. Marschner, H., 1995. Mineral Nutrition of Higher Plants. Second Edition, Academic Press, Harcourt Brace and Company Publishers, New York, pp: 985.
2. Bowen, G.D. and A.D. Rovira, 1991. The rhizosphere, hidden half in plant roots. pp: 641-669, Marcel Dekker, New York.
3. Runyan, C.W. and P.D. Odorico, 2013. Positive feedbacks and bistability associated with phosphorus vegetation microbial interactions. *Advan. Water Res.*, 52: 151-164.
4. Harley, J.L. and S.E. Smith, 1983. Mycorrhizal symbiosis (1st ed.). Academic Press, London.
5. Wang, B. and Y.L. Qiu, 2006. Phylogenetic distribution and evolution of mycorrhizas in land plants. *Mycorrhiza*, 16(5): 299-363.
6. Achal, V., V.V. Savant and R.M. Sudhakara, 2007. Phosphate solubilization by wide type strains and UV induced mutants of *Aspergillus tubingensis*. *Soil Biol. Biochem.*, 39: 695-699.
7. Holford, I.C.R., 1997. Soil phosphorus: its measurement, and its uptake by plants. *Aust. J. Soil Res.*, 35: 227-239.
8. Heffer, P. and M. Prud'homme, 2010. Fertilizer outlook 78th IFA Annual Conference, Paris IFA, France, 31 May 2 June 2010.
9. FAO., 2017. World fertilizer trends and outlook to 2020. Summary Report. Rome, 38: 11-15.
10. Cordell, D. and S. White, 2011. Peak phosphorus: clarifying the key issues of a vigorous debate about long-term phosphorus security. *Sustainability*, 3(10): 2027-2049.
11. Sushanta, S., S. Bholanath, M. Sidhu, P. Sajal and D. Partha, 2014. Grain yield and phosphorus uptake by wheat as influenced by long-term phosphorus fertilization. *African Journal of Agricultural Research*, 9(6): 607-612.
12. Al-Harbi S.F., A.M. Ghoneim, A.S. Modaihsh and M.O. Mahjoub, 2013. Effect of foliar and soil application of phosphorus on phosphorus uptake, use efficiency and wheat grain yield in calcareous soil. *J. Appl. Sci.*, 13(1): 188-192.
13. Ghafoor, A.M.R., 2016. Effect of phosphorus fertilizer application on some yield components of wheat and phosphorus use efficiency in calcareous soil. *J. Dynam. Agric. Res.*, 3(4): 46-52.
14. El-Gibaly, M., F. El-Rewiny, M. Abdel Nasser and T.H. Dahtory, 1977. Studies on phosphate solubilizing bacteria in soil and rhizosphere of different plants, I – Occurrence of bacteria, acid producers and phosphate dissolvers. *ZbIBakt*, II, 132: 233- 239.
15. Zayed, G., 2005. Bio-production of compost with low pH and high soluble phosphorus from sugar cane bagas seenriched with rock phosphate. *World Journal of Microbiology & Biotechnology*, 21: 747-752.
16. Javot, H., N. Pumplin and M.J. Harrison, 2007. Phosphate in the arbuscular mycorrhizal symbiosis: transport properties and regulatory roles. *Plant Cell Environ.*, 30: 310-322.

17. Zayed, G., 1997. Can immobilization of *Bacillus megatherium* cells in alginate beads protect them against bacteriophages? Journal of Plant and Soil, 197: 1-7.
18. Hammad, H.H., A.A. Awad and O.S. El-Kobisy, 2011. Influence of some plant growth promoting rhizobacteria (PGPR) on vegetative growth, nitrogen and phosphorus contents and anatomical characteristics of *Taxodium distichum* Rich. transplants. Bull. Agric., Cairo Univ., 62: 29-39.
19. Nelson, R. and P.N. Achar, 2001. Stimulation of growth and nutrient uptake by VAM fungi in *Brassica oleracea* var. capitata. Biolo. Plant., 44(2): 277- 281.
20. Mukerji, K.G. and M. Sharma, 1996. Mycorrhizal relationships in forest eco-system. In: Forests -A Global Perspective. Eds. S. K. Majumdar, E.W. Miller and F. J. Brenner. The Pennsylvania Academy, of Science, U.S.A., pp: 95-125.
21. Lakshman, H.C., 2010. Bioinoculants for Integrated Plant Growth. M.D. Publications (Pvt.) Ltd. New Delhi. India, pp: 549 pages.
22. Poonguzhali, S., M. Madhiyan and T.M. Sa, 2008. Isolation and identification of phosphate solubilizing bacteria from Chinese cabbage and their effect on growth and phosphorous utilization of plants. J. Micro. Biol. Biotechnol., 18: 773-777.
23. Lakshman, H.C., 2009. Growth response of and nitrogen fixation of *Phaseolus lunatus* (Lima bean) with the inoculation of AM Fungi and Rhizobium. Asian. Sci., 4(1-2): 37-41.
24. Rise, 1994. Some tannin producing tree species. Ecosystem Research and Development Bureau College. Laguna Philippines, 5(5).
25. Rise, 1995. Mahogeny (*Swietenia macrophylla* king.) and Narra (*Preracarpus spp.*) Ecosystem research and Development Bureau College. Laguna Philippines, 7(1).
26. Prinson, J.H., 1986. Potential of *Albizia lebbek* as a tropical fodder tree-A review of literature, Tropical Grasslands, 29: 78-83.
27. Orwa, C., A. Mutua, R. Kindt, R. Jamnadass and S. Anthony, 2009. Agroforest tree Database: a tree reference and selection guide version 4.0. World Agroforestry Centre, Kenya.
28. Ibrahim, H.M., 2009. Response of flame seedless and superior grapevines grown in sandy calcareous soil to phosphate dissolving bacteria treatments. J. Agric. Res., 87(1): 285-300.
29. Ahmed, F.F. and M.A.M. Abada, 2012. Response of Thompson seedless grapevines to some slow release N, P and K fertilizers. Egypt. J. Agric. Res., 90(3): 1-16.
30. Shaheen, M.A., S.M. Abd El-Wahab, F.M. El-Morsy and A.S.S. Ahmed, 2013. Effect of organic and bio-fertilizers as a partial substitute for NPK mineral fertilizer on vegetative growth, leaf mineral content, yield and fruit quality of superior grapevine. J. Hort. Sci. and Ornamental Plants, 5(3): 151-159.
31. Shaaban, A.S., 2014. Effect of organic fertilization on growth and quality of superior grapevine. Ph.D. Thesis, Fac. Agric., Cairo Univ., Egypt.
32. Cabrera, O., G.I. Valera and M.I.F. Aguirre, 2003. The role of biofertilizers in Agricultural Crops in the Central Region of Mexico. Agric. Techn. en Mexico Inst. national de Investigaciones Forestales agricolas y ecuatorias (INFAP), 2: 231-250.
33. Kannaiyan, S., 2003. Biotechnology of Biofertilizers. Alpha Sci. Int. Ltd., PO Box 4067 Pangbourne R. 68 UK, 1-275.
34. Black, C.A., D.D. Evans, L.E. Ensminger, J.L. White and F.E. Clark, 1965. Methods of Analysis for Soils, Plants and Water. Agric. Pub., Univ. of California, Riverside, USA.
35. Menge, J.A., V.X. Lembringt and E.L. Jahson, 1977. Utilization of Mycorrhizal fungi in citrus nursery. Proc. Int. Soc. Citriculture, 1: 129.
36. Giovannetti, M. and B. Mosse, 1980. An evaluation of techniques for measuring vasicular-arbuscular mycorrhizal infection in root. New Phytologist, 84: 489-500.
37. A.O.A.C., 1980. Association of Official Agricultural Chemists, Official Methods of Analysis Benjamin Franklin Station, Washington, D.C. USA, pp: 495-510.
38. Dubios, M., K.A. Gilles, J.K. Hamilton, P.A. Reber and F. Smith, 1956. Colorimetric method for determination of sugars and related substances. Anal. Chem., 28: 350-356.
39. Pregl, P., 1945. Quantitative Organic Microanalysis 4th Ed., Churchill Publishing Co., London. Press, Boca Raton, FL.
40. King, E.J., 1951. Micro-Analysis in Medical Biochemistry. 2nd Ed. Churchil. London.
41. Piper, C.S., 1950. Soil and Plant Analysis. Inter. Sci., Pulb, New York, pp: 368.
42. Snedecor, G.W. and W.G. Cochran, 1980. Statistical Methods. Oxford and J.B.H. Bub com. 7th Edition, USA.

43. Mahfouz, S.A. and M.A. Sharaf-Eldin, 2007. Effect of mineral vs. biofertilizer on growth, yield, and essential oil content of fennel (*Foeniculum vulgare* Mill.) International Agrophysics, 21(4): 361-366.
44. Hasaneen, M.N.A., M.E. Younis and S.M.N. Tourky, 2009. Plant growth, metabolism and adaptation in relation to stress conditions. XXV. Salinity-biofertility interactive effects on nitrogen and phosphorus metabolites and enzyme activities in *Lactuca sativa*. Agrochimica, 58: 273-283.
45. Motosugi, H., Y. Yamamoto, T. Naruo, H. Kitabayash and T. Ishii, 2002. Comparison of the growth and leaf mineral concentrations between three grapevine rootstocks and their corresponding tetraploids inoculated with an arbuscular mycorrhizal fungus *Gigaspora margarita*. Vitis., 4(1): 21-25.
46. Shanan, A.T. and A.M. Higazy, 2009. Integrated biofertilization management and cyanobacteria application to improve growth and flower quality of *Matthiolaincana*. Res. J. Agric. and Biolog. Sci., 5(6): 1162-1168.
47. Grzesik, M., Z. Romanowsk, A. Duda and H. Kala, 2017. The role of biofertilizers in agricultural crops in the central region of Mexico. Agricultural Technical en Mexico Institutronational de Investigaciones Forest Alesagricolasyecuaris (INFAP), 2: 231-250.
48. Prasanna, R., N. Lata, A. Radhika, S. Jadhav, J. Monica and D. Brahma, 2009. Rhizosphere dynamics of inoculated cyanobacteria and their growth-promoting role in rice crop. Egyptian Journal of Biology, 11: 26-3.
49. Anand, M., B. Kumar and D. Nath, 2015. Cyanobacterial consortium in the improvement of maize crop .International Journal of Current Microbiology and Applied Sciences, 4(3): 264-274.
50. Mohsen, A.A.M., A.S.A. Salamaand and F.M.A. El-Saadony, 2016. The effect of foliar spray with Cyanobacterial extracts on growth, yield and quality of lettuce plants (*Lactuca sativa* L.). Middle East Journal of Agriculture Research, 5(1): 90-96.
51. Goussous, S.J. and M.J. Mohammad, 2009. Comparative effect of two arbuscular mycorrhiza and N and P fertilizers on growth and nutrient uptake of onions. Int. J. Agric. Biol., 11: 463-467.
52. Brachmann, A. and M. Parniske, 2006. The most widespread symbiosis on earth. PloS Biol., 4: 239- 240.
53. Smith, S.E. and D.J. Read, 1997. Mycorrhizal Symbiosis. Biologia Plantarum, 40: 154.
54. Jeffries, P and L.M. Barea, 2001. Arbuscular Mycorrhiza; A key Component of Sustainable Plant-soil Ecosystems Fungal Associations, pp: 51-75.
55. Zubillaga, M.M., J.P. Aristi and R.S. Lavado, 2002. Effect of phosphorus and nitrogen fertilization on sunflower (*Helianthus annuus* L.) nitrogen uptake and yield. J. Agron. Crop Sci., 188: 267-274.
56. Ekin, Z., F. Ouz, M.Erman and E. Qun, 2009. The effect of Bacillus sp. OSU-142 inoculation at various levels of nitrogen fertilization on growth, tuber distribution and yield of potato (*Solanumtu berosum* L.). Afr. J. Biotechnol., 8(18): 4418-4424.
57. Chen, Y.P., P.D. Rekha, A. B. Arun, F.T. Shen, W.A. Lai and C.C. Young, 2006. Phosphate solubilizing bacteria from subtropical soil and their tricalcium phosphate solubilizing abilities. Appl. Soil Ecol., 34: 33-41.
58. Barea, M., R. Azcon and C. Zcon, 1983. Interaction between phosphate-solubilizing bacteria and VA mycorrhiza to improve the nutrition of rock-phosphate by plants in non-acidic soils". Third International congress on Phosphorus compounds U.S.A. Nov. 15-19: 127-152.
59. Kalavathi, B.P., P. Santhakrishnan and M.P. Divya, 2000. "Effect of VA mycorrhizal fungi and phosphorus solubilizing bacterium in Neem". Indian Forester, 72: 67-70.