© IDOSI Publications, 2021

DOI: 10.5829/idosi.jhsop.2021.289.300

# Comparative Study of Salinity Effects on Some Olive Cultivars

Abdthialjalal Z. Hassan

Department of Olive and Semi-Arid Zone Fruits, Horticulture Research Institute, Agricultural Research Center, Giza, Egypt

Abstract: This study was conducted during 2016 and 2017 seasons on eleven own-rooted olive (Olea europaea L.) cultivars (Picual, Koroneiki, Manzanillo, Coratina, Aggezi Shami, Aggezi Akse, Kalamata, Dolce, Maraki, Frantoio and Sewia). The selected cultivars were thirteen years old and were drip irrigated with ground saline water (EC 7.06 dS/m - 5648 ppm) to select the most salinity tolerant ones for the expansion of cultivated area in new reclaimed lands in Egypt. Chemical analysis of leaves and roots indicated that tolerant cultivars tended to have decreased Na content and increased K and N content percentages as well as K/Na ratio. Leaf analysis showed that higher Ca content percentage, Ca/Na and K+Ca/Na ratios distinguished tolerant ones. In both seasons, Picual contained decreased Na content in leaves and roots, total phenols percentage, proline content and turgid weight/fresh weight ratio in leaves, whereas it accumulated higher values of K and N contents, K/Na ratio in leaves and roots, Ca content, Ca/Na as well as K+Ca/Na ratios in leaves. On the contrary, Dolce significantly recorded the highest leaf and root Na contents and leaf turgid weight/fresh weight ratio, whereas it contained lowest K and N percentages as well as K/Na ratio in leaves and roots. Also, Dolce maintained minimum Ca percentage, Ca/Na, K+Ca/Na ratios as well as relative water content percentage in leaves. Aggezi Akse recorded the highest leaf total phenols content. Coratina, Aggezi Akse and Dolce contained higher values of leaf proline content. Results indicated that sensitive cultivars were not included in this study. Picual and Kalamata cvs. were the most tolerant cultivars while Aggezi Akse and Dolce cvs. were the least tolerant, other cultivars were in between when irrigated with saline ground water (5648 ppm).

Key words: Olea europaea · Cultivars · Salinity · Sodium · Potassium · Calcium

# INTRODUCTION

Salinity is a worldwide major problem facing crop production nowadays. It is estimated that approximately one-third of the world's irrigated lands and half the lands in semi-arid and coastal regions are affected by salinization; 10 million hectare (ha) of irrigated lands are abandoned annually because of excessive salinity. Hence, an effective way must be found to use soil of high salinity by the cultivation of tolerant cultivars [1]. The harmful effects of salinity on plants include negative effects on water relations, nutritional and osmotic imbalance, specific ion effects that lower photosynthetic rate and impair growth, oxidative stress or a combination of the previously mentioned effects [2-5].

Mediterranean and Middle East regions include a high olive (Olea europaea L.) plantation areas which

suffer from water shortage in semi-arid and arid zones. To overcome water scarcity, the use of saline water is adapted in olive production. the use of saline ground water as the only source for irrigation are currently, the main causes of salinization in Egypt [6]. Olive is moderately tolerant to salinity. Cultivated olive, a typical crop species of the semi-arid regions, could successfully face the new scenarios of climate change through tolerant varieties selection to salt and drought stresses [7]. In Egypt, olive harvested area was 214147.62 Feddans, producing 1080091 Tons in 2019 [8]. Approximately 0.9 million ha (2.1 million Feddans) suffer from salinization problems in the cultivated irrigated areas in Egypt. Furthermore, 60% of the cultivated lands in the northern Delta, 20% of the southern Delta and Middle Egypt and 25% of the Upper Egypt regions are all salt-affected [6].

**Corresponding Author:** Abdthialjalal Z. Hassan, Department of Olive and Semi-Arid Zone Fruits, Horticulture Research Institute, Agricultural Research Center, Giza, Egypt.

The response to salinity is a genotypic or cultivar dependent characteristic [9-11]. Demiral [12] found that salinity negatively affected the K<sup>+</sup> and Ca<sup>++</sup> contents of plant tissues. Ranking of cultivars on the basis of Na<sup>+</sup> accumulation and the K<sup>+</sup>:Na<sup>+</sup> ratio were more consistent than ranking based on shoot growth parameters [13]. Kchaou et al. [14]; Bader et al. [15] and Hassan et al. [16] indicated that the degree of tolerance to salinity varied between different olive cultivars. Much research has been devoted to the interaction between salinity and olive cultivation, however, these were made on seedling stages and more information is needed to assure sustainable long-term olive production based on advanced agro-techniques for saline conditions in the field [1]. Young trees are generally less tolerant to saline water than mature trees. Tolerant cultivars use is recommended in problematic saline sites [17, 14]. However, the number of evaluated cultivars is very limited if compared to the thousands of genotypes under cultivation worldwide [7]. Due to the expansion of cultivated area in new reclaimed lands in Egypt, salinity tolerant cultivars are of great importance but there is no sufficient literature about salt tolerance degree of local and imported olive tree cultivars, also there is shortage in studies comparing between cultivars when irrigated with saline ground water under Egyptian conditions. The response of eleven selected cultivars to salinity is comparatively studied.

### MATERIALS AND METHODS

Plant Materials and Experimental Conditions: This work was carried out during 2016 and 2017 seasons on eleven own-rooted olive (*Olea europaea* L.) cultivars (Picual, Koroneiki, Manzanillo, Coratina, Aggezi Shami, Aggezi Akse, Kalamata, Dolce, Maraki, Frantoio and Sewia). The selected cultivars were thirteen years old growing in a private farm. The selected cultivars were planted in sandy loam soil spaced at  $6 \times 3$  m and were irrigated with saline ground water (EC 7.06 dS/m – 5648 ppm) using drip irrigation system (4400 m³/fed.). The trees were subjected to the regularly recommended cultural practices and were free from pathogens and physiological disorders. Each cultivar was represented by three trees, each tree was a replicate.

Soil and water samples were collected. Soil samples were air-dried, grounded to pass through a 2.0 mm sieve and then mixed. Soil texture was sandy loam. Chemical

properties of the soil fractions at depth 0-30 cm and 30-60 cm and water sample were determined in Soils, Water & Environment Res. Ins. Labs and are presented in Tables 1, 2 and 3, respectively.

Fully expanded mature leaf samples from the middle portion of non-bearing shoots and fine root samples for each cultivar were collected in July for each season, then washed with tap water followed by distilled water. Leaf and root samples were oven dried at 70°C until completely dry. Leaf dry samples were digested in a mixture of sulfuric and perchloric acids according to Piper [18] to estimate Na, K, N and Ca percentages, the same was done for fine root samples to determine Na, K and N after digestion [19]. Also, K+/Na+ leaf and roots ratios, Ca+/Na+ and K+ Ca+/Na+ leaf ratios were calculated. Leaf Cl-percentage was also extracted from dry samples with distilled hot water and titrated with standard silver nitrate solution and then determined according to A.O.A.C. [20].

The total phenols percentage was determined in fresh leaf samples at 760 nm (JENWAY 6405UV/Vis Spectrophotometer), using the Folin–Ciocalteu reagent according to Singleton *et al.* [21].

Proline estimation (mg/100g) was performed using dried leaf samples and absorbance at 520 nm was measured spectrophotometrically [22].

Twenty punched leaf discs from the middle of the leaves of each cultivar were obtained using hole puncher, fresh weighted (FW) then placed in distilled water (20 ml) in petri dishes with lids to dehydrate for 24 h in a dark place, reweighted for turgid weight (TW), then placed in an oven at 70°C for 72 h and dry weighted (DW) for the calculation of relative water content percentage according to Ben-Ahmed *et al.* [23] using the equation:

RWC (%) =  $[(FW-DW)/(TW-DW)]_{\times} 100$ .

Also, the TW/FW ratio of leaf discs were estimated for each cultivar.

**Experimental Layout:** The experimental treatments (eleven olive cultivars) were arranged in randomized complete block design. Data recorded in 2016 and 2017 seasons were subjected to analysis of variance (ANOVA) according to Snedecor and Cochran [24] and the means were differentiated using Duncan multiple tests at the level of probability 5% [25].

Table 1: Chemical characteristics, available macro and micronutrients of the tested soil sample (0-30 cm) collected from the experimental area

			mEq/L.							
Ca <sup>++</sup>	Mg <sup>++</sup>	Na <sup>+</sup>	K <sup>+</sup>	CO <sub>3</sub> -	HCO <sub>3</sub> -	Cl <sup>-</sup>	So <sub>4</sub> -	pH (1:2.5)	EC dSm <sup>-1</sup> (1:5)	Texture class
18.42	16.91	29.31	0.64	-	4.72	27.12	33.44	8.00	7.08	Sandy loam
			mg/Kg							
N	P	K	Cu	Fe	Mn	Zn				
189.00	6.88	113.00	0.04	0.51	0.31	0.152				

Table 2: Chemical characteristics, available macro and micronutrients of the tested soil sample (30-60 cm) collected from the experimental area

			mEq/L.						
Ca <sup>++</sup>	Mg <sup>++</sup>	Na <sup>+</sup>	K <sup>+</sup>	CO <sub>3</sub> -	HCO <sub>3</sub> -	Cl <sup>-</sup>	So <sub>4</sub> -	pH (1:2.5)	EC dSm <sup>-1</sup> (1:5)
7.89	2.97	12.90	0.26	-	2.83	14.41	6.78	7.20	2.68
			mg/Kg						
N	P	K	Cu	Fe	Mn	Zn			
72.00	6.22	59.00	0.048	0.678	0.248	0.124			

Table 3: Chemical properties and available macro and micronutrients of the tested ground water sample collected from the experimental area

	1 1								1	
			mEq/L						EC	
Ca <sup>++</sup>	Mg <sup>++</sup>	Na <sup>+</sup>	K <sup>+</sup>	CO <sub>3</sub> -	HCO <sub>3</sub> -	Cl <sup>-</sup>	So <sub>4</sub> -	pН	dSm <sup>-1</sup>	ppm
16.06	11.74	38.26	0.17	0.00	1.75	29.15	35.33	8.30	7.06	5648.0
			mg/L							
N (NH <sub>4</sub> <sup>+</sup> )	N (NO <sub>3</sub> -)	P	В	Zn	Fe	Mn	Cu (µg/L)	SAR		
2.80	4.90	0.03	0.02	0.013	0.135	0.096	< 0.20	10.26		

## RESULTS AND DISCUSSIONS

Dealing with soil EC, it was 7.08 and 2.68 dSm<sup>-1</sup> in 0-30 and 30-60 cm soil fractions, respectively (Table 1 and 2). Concerning irrigation water EC (Table, 3), the estimated EC value was 7.06 dSm<sup>-1</sup> (5648 ppm). It was recommended for olive cultivation that soil EC in the root zone growth area at a level lower than 6 dSm<sup>-1</sup>. Also, it was suggested that in the long-term, 7.50 dSm<sup>-1</sup> water is not suitable for sustainable olive cultivation [26, 27]. In this study, no salt toxicity visual signs were observed such as tip burn, necrosis and/or shoot die back in all examined cultivars under farm conditions [28]. About 3500 m<sup>3</sup>/F. year was used for irrigation plus 900 m<sup>3</sup>/F. year (leaching methodology) using drip irrigation. It was reported that using drip irrigation allowed water applying with a high frequency and maintaining a high humidity in the soil, which avoided the harmful salt concentration for the roots [29].

**Leaf and Root Sodium Percentages:** Data presented in Table (4) show leaf and root sodium percentages of studied olive cultivars in 2016 and 2017 seasons. In both seasons, Dolce cv. showed the highest significant leaf

and root sodium values (1.40 and 1.42 % as well as 1.50 and 1.51 %, respectively), whereas Picual cv. leaves and roots statistically accumulated the lowest significant sodium values (0.56 and 0.63 % as well as 0.72 and 0.74 %, respectively).

It was concluded that the ability to regulate salt entry to the shoot can be used to screen genotypes for salt tolerance [17]. It's worthy to mention that in all studied cultivars, leaves sodium values were lesser than that of roots, which confirms Nabila et al. [30]; Olyaei et al. [5]; Kasırğa and Demiral [31] and Larbi et al. [32] findings, who noticed that sodium concentration level in the aerial parts of olive plants was lower than that of roots. In salt sensitive cultivars, sodium concentration increased more in the leaves than the roots [33, 34, 12, 35, 36, 15]. Mousavi et al. [7] mentioned that sodium ions concentration increase in leaves was higher than in shoots and roots of olive plants under severe conditions of salt stress. Moreover, Aparicio et al. [36] concluded that the most salt-tolerant genotypes (Ocal and Picudo cvs.) were characterized by an important accumulation of Na<sup>+</sup> in root and an important inhibition of translocation of Na+ to leaf in nutrient solution in a growth chamber pot experiment. They also mentioned that Na in leaves of six olive cultivars ranged 0.53-2.62%, while root Na ranged

Table 4: Leaf and root sodium and potassium percentages of olive cultivars in 2016 and 2017 seasons

		Na %				K %		
	Leaves		Roots		Leaves		Roots	
Cultivars	2016	2017	2016	2017	2016	2017	2016	2017
Picual	0.56 H	0.63 F	0.72 F	0.74 E	1.15 A	1.10 A	1.13 A	1.12 A
Kalamata	0.82 G	0.89 E	0.89 E	0.94 D	1.11 AB	1.08 A	1.08 AB	1.04 AB
Koroneiki	0.89 EF	0.95 DE	1.15 D	1.16 C	1.04 B	1.07 A	1.02 B	0.93 C
Aggezi Shami	0.86 FG	0.91 DE	0.95 E	0.99 D	1.07 AB	1.05 A	1.02 B	0.98 BC
Frantoio	0.84 G	0.90 DE	0.91 E	0.97 D	1.09 AB	1.06 A	1.07 AB	1.03 AB
Sewia	1.09 D	1.16 C	1.22 B-D	1.20 C	0.88 C	0.82 B	0.78 C	0.62 D
Maraki	0.92 E	0.96 D	1.20 CD	1.22 C	0.76 D	0.84 B	0.72 C	0.63 D
Coratina	1.17 BC	1.23 B	1.28 BC	1.27 C	0.55 E	0.53 C	0.51 D	0.47 E
Aggezi Akse	1.22 B	1.23 B	1.31 B	1.38 B	0.54 E	0.43 D	0.35 EF	0.31 F
Manzanillo	1.14 CD	1.20 BC	1.25 BC	1.26 C	0.62 E	0.56 C	0.42 E	0.40 E
Dolce	1.40 A	1.42 A	1.50 A	1.51 A	0.33 F	0.23 E	0.30 F	0.12 G

Means designated with the same letter within column in each season are not significantly different according to Duncan's Multiple Range Test (DMRT) at 0.05 level of probability.

2.73-5.03% at 200 mM salinity. The ion exclusion and compartmentation at the root level regulates ion concentration in the xylem so as to prevent the accumulation of potentially toxic ions in the aerial parts [37, 34, 28]. This mechanism works effectively in all studied olive tree cultivars, since these cultivars tended to accumulate higher Na ions at the root level than leaves, without detection of any toxicity symptoms after a long period of salt stress (since 2007). Similarly, Olyaei et al. [5] found that leaf and root Na content of Iranian olive cultivars ranged 0.45-1.21 and 1.56-2.32% respectively. It was reported that olive trees can tolerate irrigation water salinity of up to 5 dS/m with a SAR of 18 and can produce new growth when leaf Na levels ranged 0.4-0.5% dry weight [35]. In this study, Trees under study irrigated with 7.06 dS/m water salinity, with a SAR of 10.26 and still growing and yielding when leaf Na level reached up to 1.42%.

The above results are in line with Tabatabaei [38] and Chartzoulakis [39] who concluded that there is a genotypic difference among olive cultivars in their Na<sup>+</sup> accumulation. Na<sup>+</sup> uptake reduction and accumulation by plants is one of the most important mechanisms of plant resistance to salt stress [40]. Similar result was obtained by Benlloch *et al.* [41]; El-Sayed *et al.* [42]; Marin *et al.* [43]; Hassan *et al.* [16] and Hassan [44] who reported that Picual cv. is tolerant to salinity. Moreover, Abd Elhameed [45] found that Picual cv. was lesser than Kalamata in Na<sup>+</sup> leaf content. Chartzoulakis *et al.* [9] found that Kalamata cv. is resistant to salinity, while Koroneiki is less tolerant. Chartzoulakis [39] declared that salt-tolerant cultivar Kalamata has a more efficient mechanism regulating salt translocation to the shoot, keeping leaf Na<sup>+</sup> concentration

low, even exposed to 200 mM NaCl. The above results are in agreement with Melgar et al. [46] who found that root Na<sup>+</sup> content was higher in Koroneiki cv. than in Picual cv., reflecting differences in salinity tolerance. Similarly, Aly [47] mentioned that Aggezi transplants recorded the highest leaf Na content followed by Manzanillo then Koroneiki with significant differences. With respect to sodium allocation, Heimler et al. [49]; Rossi et al. [50] and Pandolfi et al. [11] reported that Leccino cv. (salt-sensitive) accumulated more sodium than Frantoio cv. (salt-tolerant) in all organs, especially in leaves. Rossi et al. [50] also mentioned that sodium is retained in root and translocated to aboveground organs in Frantoio lesser than in Leccino. Tabatabaei [1] reported that Manzanillo was sensitive to salinity. At 100 mM NaCl, Frantoio was able to accumulate less Na<sup>+</sup> than Manzanillo [13]. Hassan [44] found that Manzanillo cv. significantly achieved higher leaf Na percentages than Picual cv. in both seasons when irrigated with saline ground water.

Leaf and Root Potassium Percentages: Results of studied olive cultivars in 2016 and 2017 seasons are presented in Table (4). Dealing with leaf potassium percentage, higher percentages of Picual, Kalamata, Aggezi Shami and Frantoio cvs. were recorded with insignificant differences (1.15, 1.11, 1.07 and 1.09 % in the 1<sup>st</sup> season as well as 1.10, 1.08, 1.05 and 1.06 % in the 2<sup>nd</sup> season, respectively). The same trend of higher root potassium percentages was observed by Picual, Kalamata and Frantoio cvs. in the two studied seasons. However, Dolce cv. took the other way around in both seasons regarding leaf and root potassium content. Other cultivars were in between.

The decline of K concentration under salinity conditions was the case for olive trees, particularly in salt-sensitive cultivars [1]. Probably, the presence of K<sup>+</sup> enhances Na<sup>+</sup> exclusion by controlling channel selectivity. With high K concentration in leaves, the cultivar prevented osmotically Na transport from the roots to the aerial parts [51]. In contrast to Na<sup>+</sup>, the cultivars took and translocated a large quantity of K<sup>+</sup> to the canopy [12] and leaves [31]. Tabatabaei [38] concluded that there is a genotypic difference among olive cultivars in their ability to accumulate K+. Also, Kasırğa and Demiral [31] mentioned that the concentration of K was regarded as an indication of adaptation to salinity in olive. The above results are in agreement with Koubouris et al. [52] who revealed that the antagonistic role of Na and K was confirmed by plant tissue analysis. Similar results were obtained by Aparicio et al. [36] and Olyaei et al. [5] who mentioned that leaf and root K content ranged 0.75-1.43 and 0.42-0.90 % of six olive cultivars as well as 0.97-1.12 and 0.77-1.04% of Iranian olive cultivars, respectively.

Tattini et al. [33] found that K<sup>+</sup> content was always decreased to a greater extent in Leccino (salt-sensitive) than in Frontoio (salt-tolerant). El-Sayed et al. [42] noted that different accumulation of K content in 14 olive cultivar transplants, showing that Picual cv. significantly surpassed Aggezi, Koroneiki and Manzanillo cvs. in both seasons. Aly [47] found that Koroneiki surpassed in leaf K content followed by Manzanillo then Aggezi with significant differences. Perica et al. [13] demonstrated that Manzanillo accumulated less K<sup>+</sup> than Frantoio (0-100 mM salinity). Higher leaf K<sup>+</sup> content accumulated in Picual and Koroneiki cvs. than Manzanillo, Coratina and Eggazi Shami evs. transplants [48]. Nabila et al. [30] showed that Picual accumulated higher root K content than Manzanillo. Similarly, Abd Elhameed [45] and Hassan [44] demonstrated that leaf K+ content of Picual cv. was higher than that of Kalamata and Manzanillo cvs., respectively in both seasons. Mousavi et al. [7] found that higher leaf K<sup>+</sup> content of Royal de Cazorla cv. (tolerant) than Koroneiki cv. (susceptible) with insignificant difference in a hydroponic system.

**Leaf and Root** K\*/Na\* Ratios: Table (5) represents K\*/Na\* leaf and root ratios of different olive cultivars in 2016 and 2017 seasons. Both ratios were decreased drastically (2.04-0.23, 1.75-0.16, 1.58-0.20 and 1.50-0.08, in 2016 and 2017, respectively). The same trend was clear between olive cultivars in both seasons, as Picual cv. significantly maintained the highest leaf and root ratios (2.04 and 1.75

as well as 1.58 and 1.50 in the 1<sup>st</sup> season and the 2<sup>nd</sup> season, respectively) followed by Kalamata and Frantoio cultivars and they were at par, whilst Dolce cv. significantly exhibited the lowest ratios in both seasons. Studied cultivars tended to have higher leaf values than root.

Tattini [53] concluded that the resistance mechanism of salt-tolerant olive cultivars is probably related to the ability to maintain an appropriate K/Na ratio in actively growing tissues. Cultivars ranking based on Na<sup>+</sup> accumulation and K<sup>+</sup>/Na<sup>+</sup> ratio was more consistent than ranking based on shoot growth parameters [17, 13]. Olive owes its tolerance of salinity to its ability to restrict transport to shoots, isolate Na in vacuoles and maintain a high K/Na ratio to support tissue metabolism [54]. Demiral [12] suggested that the translocation rate of the ions might be evaluated as reliable criteria giving clues to salt-tolerance levels of O. europaea cultivars. Similar result was found by Kasırğa and Demiral [31] who reported that the K/Na ratio was found higher in leaves than in roots. They also added that higher K/Na ratio of the plant leaves can be accepted as key indicator reflecting the level of adaptation of the cultivar to salt stress. A well balanced K+:Na+ ratio is crucial for the proper adjustment of stomatal function, activation of enzymes, protein synthesis, cell osmoregulation, oxidants metabolism, photosynthesis and turgor maintenance [55].

The salt-sensitive Leccino cv. showed a lower K<sup>+</sup>/Na<sup>+</sup> selectivity ratio than the salt-tolerant Frantoio cv. [33, 53, 49, 10]. Tabatabaei [1] found that difference in K selectivity among cultivars was also observed in the K/Na ratio. Reduction in leaf K/Na ratio was became more pronounced in Manzanillo and Zard cultivars. Perica et al. [13] gave evidence that in 0, 33, 66 and 100 mM NaCl, Frantoio maintained the highest K<sup>+</sup>/Na<sup>+</sup> ratio. At 166 mM NaCl, lower values were obtained by Leccino and Manzanillo. Aparicio et al. [36] found that the lowest decrease in K<sup>+</sup>/Na<sup>+</sup> ratio was shown by the most tolerant genotypes (Ocal and Picudo cvs.) grown in nutrient solution in a growth chamber pot experiment. They also mentioned that leaf and root K<sup>+</sup>/Na<sup>+</sup> ratio ranged 0.29-2.30 and 0.12-0.30% respectively. Abd Elhameed [45] showed that Picual had significant higher leaf K<sup>+</sup>/Na<sup>+</sup> ratios than Kalamata in both studied seasons. Mousavi et al. [7] showed that the K<sup>+</sup>/Na<sup>+</sup> ratio in Royal de Cazorla leaves was higher than in Koroneiki after the addition of 200 mM NaCl, confirming that salt tolerance as a consequence of both the ability to prevent Na+ accumulation and to maintain high levels of K<sup>+</sup> in leaves.

Table 5: Leaf and root K\*/Na\* ratios and nitrogen percentages of olive cultivars in 2016 and 2017 seasons

		K/Na ratio						
	Leaves		Roots		Leaves		Roots	
Cultivars	2016	2017	2016	2017	2016	2017	2016	2017
Picual	2.04 A	1.75 A	1.58 A	1.50 A	1.88 A	1.90 A	1.75 A	1.70 A
Kalamata	1.35 B	1.23 B	1.24 B	1.12 B	1.91 A	1.82 B	1.67 B	1.63 B
Koroneiki	1.16 C	1.12 B	0.89 D	0.80 D	1.49 DE	1.45 DE	1.45 GH	1.41 G
Aggezi Shami	1.24 BC	1.16 B	1.08 C	0.98 C	1.53 CD	1.49 CD	1.47 EF	1.42 FG
Frantoio	1.31 B	1.17 B	1.17 BC	1.08 BC	1.65 B	1.56 C	1.59 C	1.54 C
Sewia	0.81 D	0.71 D	0.65 E	0.51 E	1.56 C	1.53 C	1.51 D	1.48 E
Maraki	0.83 D	0.88 C	0.59 E	0.52 E	1.55 CD	1.53 C	1.50 DE	1.52 D
Coratina	0.47 E	0.43 EF	0.41 F	0.37 F	1.49 DE	1.45 DE	1.46 FG	1.42 FG
Aggezi Akse	0.44 E	0.35 F	0.27 GH	0.22 G	1.46 E	1.43 DE	1.44 GH	1.41 G
Manzanillo	0.54 E	0.47 E	0.34 FG	0.32 FG	1.52 C-E	1.45 DE	1.48 EF	1.43 F
Dolce	0.23 F	0.16 G	0.20 H	0.08 H	1.48 DE	1.41 E	1.43 H	1.40 G

Means designated with the same letter within column in each season are not significantly different according to Duncan's Multiple Range Test (DMRT) at 0.05 level of probability.

**Leaf and Root Nitrogen Percentages:** Data in Table (5) present leaf and root nitrogen percentages of different olive cultivars in 2016 and 2017 seasons. Data indicated that Kalamata cv. topped the others in leaf nitrogen percentage with insignificant variation with Picual cv. in 2016 season, but in 2017 season they switched their positions with statistical difference. Aggezi Akse cv. recorded the least value with insignificant differences with Koroneiki, Coratina, Manzanillo and Dolce cvs. in the 1st season, whilst in the 2<sup>nd</sup> season, it was Dolce cv. without statistical variation with the previously mentioned cultivars. Notably, Picual cv. gave the significantly highest nitrogen root percentages in both seasons. The same trend of lower values was observed by Koroneiki, Aggezi Akse and Dolce cultivars in both seasons.

Nabila *et al.* [30] and Kasırğa and Demiral [31] reported higher leaves nitrogen content than roots in Manzanillo and Picual cvs. as well as Gemlik cv., respectively. El-Sayed *et al.* [42] found that a higher leaf nitrogen content was recorded by Picual with significant differences with Aggezi, Koroneiki and Manzanillo cvs. in the 1<sup>st</sup> season, whilst in the 2<sup>nd</sup> season, significant difference was just with Aggezi. Aly [47] noted that Koroneiki statistically surpassed both Aggezi and Manzanillo in both seasons. Abd Elhameed [45] reported that higher N content in leaves of Picual cv. than Kalamata cv. with significant differences in both seasons. Hassan [44] confirmed that Picual leaf N content significantly exceeded that of Manzanillo in both seasons when irrigated with saline ground water.

**Leaf Calcium Percentage:** Data of 2016 and 2017 seasons showed that Picual cv. achieved significantly the highest

values (0.92 % in both seasons) and on the contrary, Dolce cv. acquired the lowest ones (Table, 6) regarding leaf Ca percentage.

Zidan et al. [56] stated that presence of K<sup>+</sup> and in particular Ca<sup>+2</sup> ions has been shown to reduce Na<sup>+</sup> influx to plant cells. Calcium maintains the integrity and function of cellular membranes. In that way root selectivity for K instead of Na is maintained [57]. Ca2+ is assumed to play an important role in retention mechanisms and sodium exclusion, which may be considered responsible for survival under salt stress conditions [58]. Jacoby [51] and Fernández-Escobar [59] noted that the presence of Ca<sup>+2</sup> enhances Na<sup>+</sup> exclusion. This presence can increase tolerance to salinity. According to Tattini et al. [33], Ca+2 tissue content was always decreased to a greater extent in salt-sensitive Leccino than in salt-tolerant Frontoio. Shaheen et al. [48] noted that Picual transplants gave the highest Ca+2 leaf content with significant difference just with Manzanillo in the 1st season. Kasırğa and Demiral [31] mentioned that leaf Ca concentration was regarded as an indicator of adaptation to salinity in olive. Mousavi et al. [7] found that higher leaf Ca<sup>++</sup> content of Royal de Cazorla (tolerant) than Koroneiki (susceptible) with significant difference in a hydroponic system. Similarly, Hassan [44] showed that Picual significantly surpassed Manzanillo in leaf Ca content in both seasons when irrigated with saline ground water.

**Leaf Ca<sup>++</sup>/Na<sup>+</sup> and K<sup>+</sup>+Ca<sup>++</sup>/Na<sup>+</sup> Ratios:** Data presented in Table (6) reveal that in both seasons, Picual cv. significantly surpassed the others recording 1.63 and 1.46 as well as 3.67 and 3.22, respectively regarding leaf Ca<sup>++</sup>/Na<sup>+</sup> ratio and K<sup>+</sup>+Ca<sup>++</sup>/Na<sup>+</sup> ratio, while Dolce cv. gave the minimum values.

Table 6: Leaf calcium percentage, calcium/sodium ratio, potassium+calcium/sodium ratio and chloride percentage of olive cultivars in 2016 and 2017 seasons

	Ca %		Ca/Na ratio		K+Ca/Na r	atio	Cl %	
	Leaves		Leaves		Leaves		Leaves	
Cultivars	2016	2017	2016	2017	2016	2017	2016	2017
Picual	0.92 A	0.92 A	1.63 A	1.46 A	3.67 A	3.22 A	0.36 D	0.41 C
Kalamata	0.78 B	0.76 B	0.95 B	0.86 B	2.30 B	2.08 B	0.34 E	0.37 E
Koroneiki	0.69 BC	0.67 BC	0.77 CD	0.70 CD	1.94 E	1.83 D	0.32 F	0.36 E
Aggezi Shami	0.73 BC	0.74 B	0.85 BC	0.81 B	2.09 D	1.96 C	0.46 A	0.50 A
Frantoio	0.74 BC	0.71 BC	0.89 BC	0.78 BC	2.20 C	1.95 C	0.43 B	0.46 B
Sewia	0.68 BC	0.67 BC	0.62 EF	0.58 E	1.43 G	1.29 F	0.36 D	0.36 E
Maraki	0.66 CD	0.65 BC	0.72 DE	0.68 D	1.55 F	1.55 E	0.45 A	0.50 A
Coratina	0.55 DE	0.50 DE	0.47 GH	0.40 F	0.94 I	0.84 H	0.37 D	0.39 D
Aggezi Akse	0.46 EF	0.44 EF	0.37 HI	0.36 F	0.81 J	0.70 I	0.39 C	0.41 C
Manzanillo	0.63 CD	0.60 CD	0.55 FG	0.50 E	1.08 H	0.97 G	0.36 D	0.39 D
Dolce	0.39 F	0.36 F	0.28 I	0.25 G	0.51 K	0.41 J	0.43 B	0.46 B

Means designated with the same letter within column in each season are not significantly different according to Duncan's Multiple Range Test (DMRT) at 0.05 level of probability.

Table 7: Leaf total phenols percentage, proline content (mg/100 g dry weight), relative water content percentage and turgid weight/fresh weight ratio of olive cultivars in 2016 and 2017 seasons

	Leaf total phenols (%)		Leaf proline co	ntent (mg/100g)	Leaf relative w	vater content (%)	Turgid weight/Fresh weight ratio	
Cultivars	2016	2017	2016	2017	2016	2017	2016	2017
Picual	1.70 E	1.86 D	0.016 C	0.019 D	65.22 B	69.36 A	1.323 C	1.223 F
Kalamata	1.83 D	2.01 C	$0.020~\mathrm{BC}$	0.023 CD	67.43 A	67.21 B	1.333 C	1.283 EF
Koroneiki	2.06 C	2.18 B	0.023 B	0.025 B-D	60.05 E	58.62 F	1.360 BC	1.263 EF
Aggezi Shami	2.04 C	2.16 B	0.023 B	0.025 B-D	61.27 D	63.49 D	1.380 BC	1.323 DE
Frantoio	1.86 D	2.03 C	0.023 B	0.023 CD	62.07 C	65.74 C	1.363 BC	1.380 CD
Sewia	1.90 D	2.06 C	0.020 BC	0.024 B-D	58.99 F	54.30 H	1.390 BC	1.440 A-C
Maraki	2.19 B	2.24 B	0.021 BC	0.021 CD	61.03 D	60.56 E	1.420 B	1.417 A-C
Coratina	2.23 B	2.21 B	0.030 A	0.035 A	61.13 D	55.16 G	1.363 BC	1.370 CD
Aggezi Akse	2.47 A	2.50 A	0.030 A	0.034 A	57.12 G	54.40 H	1.413 B	1.453 AB
Manzanillo	2.24 B	2.22 B	0.025 AB	0.026 BC	57.11 G	55.15 G	1.343 BC	1.393 B-D
Dolce	2.26 B	2.22 B	0.030 A	0.030 AB	52.90 H	52.97 I	1.490 A	1.473 A

Means designated with the same letter within column in each season are not significantly different according to Duncan's Multiple Range Test (DMRT) at 0.05 level of probability

Increasing the Ca: Na ratio in the external solution has been reported to alleviate the effects of salinity on depolarization and selectivity of the plasma membrane [60]. Kasırğa and Demiral [31] found that higher K+Ca+Mg/Na ratio of the plant leaves can be accepted as key indicator reflecting the level of adaptation of the cultivar to salt stress.

**Leaf Chloride Percentage:** Referring to leaf chloride percentage, data presented in Table (6) show that both Aggezi Shami and Maraki cvs. significantly surpassed the others and on the contrary, Koroneiki cv. recorded the minimum values in both seasons.

The above results are in line with Tattini *et al.* [33] who concluded that Cl<sup>-</sup> uptake and transport to the shoot of olive trees is lower than Na<sup>+</sup>. Olive trees are less

sensitive to leaf Cl<sup>-</sup> than Na<sup>+</sup> especially at high salinities [27] and Cl<sup>-</sup> ion does not cause toxicity in olive trees [61]. Similarly, Manzanillo cv. has been described as very resistant to high concentrations of chlorides in irrigation water [62]. In the same direction, Melgar *et al.* [29] found that after eight years of treatment, salinity did not affect leaf Cl<sup>-</sup> concentration of Picual cv. trees and was always below the toxicity threshold (0.50%). Hassan *et al.* [16] reported that remarkable variations were noticed among one-year old own-rooted cultivars in relation to chloride content, Picual accumulated lowest leaf Cl<sup>-</sup> content followed by Manzanillo then Aggezi Shami.

**Leaf Total Phenols Content Percentage:** Table (7) represents leaf total phenols content of the studied olive cultivars in 2016 and 2017 seasons. In both studied

seasons, significant differences were observed, Aggezi Akse cv. recorded the highest content (2.47 and 2.50 %, respectively), Picual cv. took the other way around (1.70 and 1.86 %, respectively) and other cultivars were in between.

Saline water irrigation increases the contents of total phenols [63, 64]. It was concluded that high salinity induced the formation of total phenolic content in the leaves, which participate in the increase of antioxidant activity. Phenolic compounds play an important role in scavenging free radicals and protect plants against the damaging effects of reactive oxygen species due to salt stress [65]. In the same direction, Cavaca et al. [66] reported that phenols, as one of the major groups of secondary metabolites in olive, are important because of their involvement in the plant response against abiotic stressors. Olive cultivars may have a different response to abiotic factors, consequently leading to distinct olive leaf phenolic profiles and anti-oxidative activity. Similarly, Demir and Cetinkaya [67] concluded that cultivars may develop a defense mechanism against saline conditions through biosynthesizing polyphenols in different amounts.

Similarly, Farag et al. [68] found that Koroneiki had significantly higher leaf content than Picual. Also, Aparicio et al. [36] showed evidence that Picual cv. leaves contained lower total phenols than Frantoio cv. at 200 mM NaCl. Rossi et al. [10] mentioned that the concentration of total phenolic compounds was higher in old-leaves of Leccino (salt-sensitive) than in those of Frantoio (salt-tolerant). Higher leaf total phenols content of Dolce agogia than Frantoio was reported by Blasi et al. [69]. Moreover, Olmo-García et al. [70] found that Picual was the poorest variety of the eleventh studied cultivars, while Frantoio, Koroneiki and Manzanillo cvs. recorded higher total phenol values. Significant differences in leaf total phenolic content of 15 Italian olive cultivars were noted [71]. Pasković et al. [72] showed that temporal variation of olive leaf biophenolic composition may significantly depend on the cultivar.

Leaf Proline Content (mg/100g): Data presented in Table (7) shows leaf proline content (mg/100 g, dry weight) of olive cultivars in 2016 and 2017 seasons. Leaves of Coratina, Aggezi Akse and Dolce cvs. contained higher proline values than the others in both studied seasons (0.030 mg/100 g for all of them in 2016 season and 0.035, 0.034 and 0.030 mg/100 g in 2017 season, respectively). Picual cv. achieved the lowest values (0.016 and 0.019 mg/100 g, respectively) with

insignificant differences with Kalamata, Maraki and Sewia cvs. in both seasons.

It was mentioned that there are differences in leaf proline content between different olive cultivars [42, 47, 45, 16]. The proline accumulation recorded in stressed plants has allowed them to improve water uptake to actively growing tissues, to preserve appropriate leaf water status and to increase photosynthetic activity as noted by Ben Ahmed et al. [73]. Similar results were reported by El-Sayed et al. [42] who found that Manzanillo cv. recorded lower proline value than Aggezi cv. in the 1st season and the results of the 2nd season were as follows Picual < Koroneiki < Aggezi. Similarly, Aly [47] found that Koroneiki cv. recorded lesser leaf proline content than Manzanillo cv. in both seasons. Shaheen et al. [48] showed that leaves of Coratina accumulated higher proline content than Koroneiki in the 2<sup>nd</sup> season. The above mentioned result is in agreement with Abd Elhameed [45] who found that Kalamata cv. significantly surpassed Picual cv. in estimated proline content at 6000 ppm salinity. Lowest proline content of Picual was reported by Hassan et al. [16] in comparison with Manzanillo and Aggezi Shami cvs. at 4000 mg/L salinity. However, the results of this study didn't agree with some findings of El-Sayed et al. [42], Aly [47], Shaheen et al. [48] and Hassan et al. [16]. This contradiction may be due to the difference between young and mature plants, as young plants are less tolerant to salinity [74]. In the same direction, Marin et al. [43] noted that results obtained with young plants might not be related to the tolerance of mature plants growing under field conditions.

Leaf Relative Water Content Percentage and Leaf Turgid Weight/Fresh Weight Ratio: With respect to leaf relative water content percentage (RWC %), data tabulated in Table (7) cleared that in 2016 season, Kalamata cv. significantly surpassed the others (67.43%), whereas Picual cv. was significantly superior in 2017 season (69.36%). In both seasons, Dolce cv. achieved statistically the lowest values. Aly [47] recorded that the highest relative water content was shown by Koroneiki followed by Manzanillo then Aggezi with significant variations in both seasons. Hassan et al. [16] mentioned that mean leaf water content showed high significant differences among the studied one-year-old cultivars. Picual recorded the highest relative water content (83.00%), while Aggizi Shami (76.25%) showed the lowest percentage and Manzanillo showed an intermediate percentage (78.25%). They also added that relative water content is important physiological parameters for measuring the water status of the plants.

Leaf turgid weight/fresh weight ratio (T/F ratio) is presented in Table (7), it's also notorious that Dolce cv. recorded the highest values in both studied seasons (1.490 and 1.473, respectively). On the contrary, Picual and Kalamata cvs. in 2016 season as well as Picual and Koroneiki cvs. in 2017 season acquired lower values (1.323 and 1.333 as well as 1.223 and 1.263, respectively). This may be due to lower accumulation of Na<sup>+</sup> in leaves of the previously mentioned cultivars (Table, 4) which resulted in lower turgid weight and led to lower T/F ratio.

It's worthy to mention that despite the numerous studies on the salt tolerance of olive cultivars, the results of these studies remained confusing due to different comparisons between cultivars. For instance, many researchers defined Picual as a salt tolerant cultivar [41-44, 16], but it was reported to be the least tolerant [36].

#### **CONCLUSIONS**

- In all studied cultivars, Na content of leaves was lower than that of roots, revealing that sensitive cultivars were not included in this study.
- An effective salt-exclusion mechanism operating in the root system was confirmed. However, the ability to use this mechanism differed between tested cultivars. Tolerant cultivars tended to have decreased Na content in leaves and roots, while the opposite was true for the least tolerant ones.
- Remarkable differences among the tested cultivars irrigated with saline ground water (EC 7.06 dSm<sup>-1</sup>/5648 ppm) in their ability to accumulate K and Ca were clear, revealing selectivity for K and Ca which resulted in higher leaf and root K/Na ratio, Ca/Na and K+Ca/Na ratios in leaves of tolerant cultivars.
- Increased leaf total phenols and proline contents as well as leaf turgid weight/fresh weight ratio distinguished less tolerant cultivars.
- It seems that Na, K and Ca contents, besides K/Na, Ca/Na and K+Ca/Na ratios in leaves may be evaluated as reliable criteria for salt tolerance of olive cultivars.
- In spite of high irrigation water EC, no salt toxicity visual signs were observed among the studied cultivars. This may be due to following the recommended cultural practices including sufficient fertilization, drip irrigation and leaching that decreased the adverse effects of salinity.
- From the findings of this investigation, it seems that Picual and Kalamata cvs. were the most tolerant

cultivars, while Aggezi Akse and Dolce cvs. were the least tolerant ones, other cultivars were in between when irrigated with saline ground water (5648 ppm).

### REFERENCES

- Tabatabaei, S.J., 2006. Effects of salinity and N on the growth, photosynthesis and N status of olive (*Olea europaea* L.) trees. Sci. Hortic., 108: 432-438.
- Talhaoui, N., A. Taamalli, A. Gómez-Caravaca, A. Fernández-Gutiérrez, A. Segura-Carretero, 2015. Phenolic compounds in olive leaves: Analytical determination, biotic and abiotic influence and health benefits. Food Res. Int., 77(2): 92-108. https://doi.org/10.1016/j.foodres.2015.09.011.
- 3. Grattan, S.R. and C.M. Grieves, 1999. Salinity-mineral nutrient relations in horticultural crops. Sci. Hortic., 78: 127-157.
- 4. Munns, R., 1993. Physiological processes limiting plant growth in saline soils: some dogmas and hypotheses. Plant, Cell Environ., 16: 15-24.
- 5. Olyaei, F., B. Baninasab, C. Ghobadi and M. Gholami, 2015. Ion content and its correlation with some physiological parameters in olive cultivars in response to salinity Iran Agric. Res., 34(2): 61-70.
- Karajeh, F., T. Oweis, A. Swelam, A. El-Gindy, D. El-Quosy, H. Khalifa, M. El-Kholy and S.A. Abd El-Hafez, 2011. Water and agriculture in Egypt. Technical paper based on the Egypt-Australia ICARDA workshop on on-farm water-use efficiency. ICARDA, Egypt.
- Mousavi, S., L. Regni, M. Bocchini, R. Mariotti, C. Nicolò, S. Mancuso, J. Googlani, M. Chakerolhosseini, C. Guerrero, E. Albertini, L. Baldoni and P. Proietti, 2019. Physiological, epigenetic and genetic regulation in some olive cultivars under salt stress. Sci. Rep., 9:1093. https://doi.org/10.1038/s41598-018-37496-5.
- 8. F A O statics, 2019. http://www.fao.org/faostat/en/#data/QC
- Chartzoulakis, K., M. Loupassaki, M. Bertaki and I. Androulakis, 2002. Effects of NaCl salinity on growth, ion content and CO<sub>2</sub> assimilation rate of six olive cultivars. Sci. Hort., 96: 235-247.
- Rossi, L., A. Francini, A. Minnocci and L. Sebastiani, 2015. Salt stress modifies apoplastic barriers in olive (*Olea europaea* L.): a comparison between a salttolerant and a salt-sensitive cultivar. Sci. Hortic., 192: 38-46 http://dx.doi.org/10.1016/ j.scienta.2015.05.023.

- 11. Pandolfi, C., N. Bazihizina, C. Giordano, S. Mancuso and E. Azzarello, 2017. Salt acclimation process: a comparison between a sensitive and a tolerant *Olea europaea* cultivar. Tree Physiol., 37: 380-388. doi:10.1093/treephys/tpw127.
- 12. Demiral, M.A., 2005. Comparative response of two olive (*Olea europaea* L.) cultivars to salinity. Turk. J. Agric. For., 29(4): 267-274.
- Perica, S., S. Goreta and G.V. Selak, 2008. Growth, biomass allocation and leaf ion concentration of seven olive (*Olea europaea* L.) cultivars under increased salinity. Sci. Hortic., 117(2): 123-129. https://doi.org/10.1016/j.scienta.2008.03.020.
- Kchaou H., A. Larbi, K. Gargouri, M. Chaieb, F. Morales and M. Msallem, 2010. Assessment of tolerance to NaCl salinity of five olive cultivars, based on growth characteristics and Na<sup>+</sup> and Cl<sup>-</sup> exclusion mechanisms. Sci. Hortic., 124: 306-315. https://doi.org/10.1016/j.scienta.2010.01.007.
- 15. Bader, B., F. Aissaoui, I. Kmicha, A. Salem, H. Chehab, K. Gargouri, D. Boujnah and M. Chaieb, 2015. Effects of salinity stress on water desalination, olive tree (*Olea europaea* L. cvs. 'Picholine', 'Meski' and 'Ascolana') growth and ion accumulation. Desalination, 364: 46-52. https://doi.org/10.1016/j.desal.2015.01.002.
- Hassan, I.F., G. Maybelle, A. Bedour, P. Proietti and L. Regni, 2020. Salinity stress effects on three different olive cultivars and the possibility of their cultivation in reclaimed lands. Plant Arch., 20(2): 2378-2382.
- 17. Gucci, R. and M. Tattini, 1997. Salinity tolerance in olive. Hort. Rev., 21: 177-214.
- 18. Piper, C.S., 1950. Soil and plant analysis. Inter. Sci. Publisher, Inc. New York, pp. 368.
- Cottenie, A., M. Verlso, L. Kilkens, G. Velghe and R. Camerlynck, 1982. Chemical Analysis of Plants and Soils. Lab. Agroch. State Univ. Gent, Belgium.
- A.O.A.C., 1970. Official Methods of Analysis of Assiociation of official Agriculture Chemists 10<sup>th</sup> Ed. Washington D.C., U.S.A.
- Singleton, V.L., R. Orthofer, R.M. Lamuela-Raventós, 1999. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Meth. Enzymol., 299: 152-178.
- 22. Bates, S.L., 1973. Rapid determination of free proline for water stress studies. Plant and Soil, 39: 205-207.
- Ben-Ahmed, C., B. Ben-rouina and M. Boukhriss,
  Changes in water relations, photosynthetic activity and proline accumulation in one-year-old

- olive trees (*Olea europaea* L., cv. "Chemlali") in response to NaCl salinity. Acta Physiol. Plant., 30:553-560.https://doi.org/10.1007/s11738-008-0154-6.
- Snedecor, G.W. and W.G. Cochran, 1980. Statistical Methods. 6<sup>th</sup> ed. Iowa State Univ. Press, Ames, Iowa, U.S.A., pp: 507.
- Duncan, D.B., 1955. Multiple range and multiple F. tests. Biometrics, 11: 1-42.
- FAO (Food and Agriculture Organization), 1985.
  Water quality for agriculture. FAO Irrigation and Drainage Paper 29, Rome, Italy.
- 27. Aragues, R., J. Puy, A. Royo and J.L. Espada, 2005. Three-year field response of young olive trees (*Olea europea* L., cv. Arbequina) to soil salinity: Trunk growth and leaf ion accumulation. Plant Soil, 271: 265-273.
- Chartzoulakis, K., 2005. Salinity and olive: Growth, salt tolerance, photosynthesis and yield. Agr. Water Manage., 78: 108-121.
- Melgar, J.C., Y. Mohamed, N. Serrano Castillo, P.A. García-Galavís, C. Navarro, M.A. Parra, G. Beltrán, M. Benlloch and R. Fernández -Escobar, 2012. Response of Olive Trees to Irrigation with Saline Water. Acta Hortic., 928: 281-286. 10.17660/ActaHortic.2012.949.40.
- Nabila, E.K., M.S. Abourayya and M.H. El-Sheikh, 2013. effect of salinity treatments on mineral content of Manzanillo and Picual olive leaves, shoots and roots. J. Appl. Sci. Res., 9(1): 258-262.
- 31. Kasırğa, E. and M.A. Demiral, 2016. Salt stress-mineral nutrient relations in olive (*Olea europaea* L.) plant. Eurasian J. Soil Sci., 5(4): 307-313.
- Larbi, A., H. Kchaou, B. Gaalichec, K. Gargouria, H. Boulald and F. Morales, 2020. Supplementary potassium and calcium improves salt tolerance in olive plants. Sci. Hortic., 260: 108912 https://doi.org/10.1016/j.scienta.2019.108912.
- 33. Tattini, M., P. Bertoni and S. Caselli, 1992. Genotipic responses of olive plants to sodium chloride. J. Plant Nut., 15(9): 1467-1485. DOI: 10.1080/01904169209364412.
- 34. Gucci, R., L. Lombardini and M. Tattini, 1997. Analysis of leaf water relations in leaves of two olive (*Olea europaea*) cultivars differing in tolerance to salinity. Tree Physiol., 17: 13-21.
- Chartzoulakis, K., 2011. The Use of Saline Water for Irrigation of Olives: Effects on Growth, Physiology, Yield and Oil Quality. Acta Hortic., 888: 97-108. DOI: 10.17660/ActaHortic.2011.888.10.

- Aparicio, C., M. Urrestarazu and M. Cordovilla, 2014.
  Comparative Physiological Analysis of Salinity Effects in Six Olive Genotypes. HortScience, 49(7): 901-904. https://www.researchgate.net/ publication/264337055.
- 37. Tattini, M., R. Gucci, M.A. Coradeschi, C. Ponzio and J.D. Everard, 1995. Growth, gas exchange and ion content in *Olea europaea* plants during salinity stress and subsequent relief. Physiol. Plant., 95: 203-210.
- 38. Tabatabaei, S.J., 2007. Salinity stress and olive: An overview. Plant Stress, 1(1): 105-112.
- 39. Chartzoulakis, K., 2014. The Potential of Saline and Residual Water Use in Olive Growing. In the Proceeding VII<sup>th</sup> IS on Olive Growing. F. Vita Serman *et al.* (eds.) Acta Hort. 1057, ISHS. DOI: 10.17660/ActaHortic.2014.1057.29.
- Zargar, S.M., R. Mahajan, J.A. Bhat, M. Nazir and R. Deshmukh, 2019. Role of silicon in plant stress tolerance: opportunities to achieve a sustainable cropping system. 3 Biotech, 9(73): 1-16. https://doi.org/10.1007/s13205-019-1613-z.
- Benlloch, M., L. Marin and R. Fernández-Escobar, 1994. Salt tolerance of various olive varieties. Acta Hortic., 356, 215-217. DOI: 10.17660/ActaHortic.1994.356.46.
- 42. El-sayed, E.H., M.E. El-said, A.H. El-sherif and S.A. Sari El-Deen, 1996. Chemical studies on the salt tolerance of some olive cultivars. Olivae, 64: 52-57.
- 43. Marin, L., M. Benlloch and R. Fernandez-Escobar, 1995. Screening for olive cultivars for salt tolerance. Sci. Hortic., 64: 113-116.
- 44. Hassan, A.Z., 2021. Minimizing the adverse effects of irrigation with saline water on olive trees, Picual and Manzanillo cultivars. Hort. Sci. & Ornamen. Plants, 13(2): 210-225. https://www.idosi.org/jhsop/13(2)21/11.pdf.
- 45. Abd Elhameed, E.E., 2018. Tolerance of some olive strains to salinity stress. Ph.D. Thesis, Fac. Agric., Cairo Univ., Egypt.
- Melgar, J.C., J.P. Syvertsen and F. Garcı'a-Sa'nchez, 2008. Can elevated CO<sub>2</sub> improve salt tolerance in olive trees? J. Plant Physiol., 165: 631-640. DOI: 10.1016/j.jplph.2007.01.015.
- 47. Aly, A.A., 2005. Physiological studies on the effect of salt stress on some olive cultivars. Ph.D. Thesis, Fac. Agric., Benha Univ., Egypt.

- 48. Shaheen, M.A., A.A. Hegazi and I.S.A. Hammam, 2011. Effect of salinity treatments on vegetative characteristics and leaves chemical content of transplants of five olive cultivars. J. Hort. Sci. & Ornamen. Plants, 3(2): 143-151.
- Heimler, D., M. Tattini, S. Ticci, M.A. Coradeschi, M.L. Traversi, 1995. Growth, ion accumulation and lipid composition of two olive genotypes under salinity stress. J. Plant Nutr., 18: 1723-1734.
- Rossi, L., M. Borghi, A. Francini, X. Linb, D. Xie,
  L. Sebastiani, 2016. Salt stress induces differential regulation of the phenylpropanoid pathway in *Olea europaea* cultivars Frantoio (salt-tolerant) and Leccino (salt-sensitive). J. Plant Physiol., 204: 8-15. http://dx.doi.org/10.1016/j.jplph.2016.07.014.
- Jacoby, B., 1999. Mechanisms involved in salt tolerance of plants. In: Handbook of plant and crop stress. Ed., Pessarakli, M. Marcel Dekker Inc., New York, USA, pp: 97-123.
- Koubouris, G., N. Tzortzakis, N. Kourgialas, M. Darioti and I. Metzidakis, 2015. Growth, photosynthesis and pollen performance in saline water treated olive plants under high temperature. Int. J. Plant Biol., 6: 6038. DOI: 10.4081/pb.2015.6038.
- 53. Tattini, M., 1994. Ionic relations of aeroponically-grown olive plants during salt stress. Plant Soil, 161: 251-256.
- Connor, D. and E. Fereres, 2005. The Physiology of Adaptation and Yield Expression in Olive. Hortic. Rev., 31: 155-229.
- 55. Abbasi, H., M. Jamil, A. Haq, S. Ali, R. Ahmad, Z. Malik and Z. Parveen, 2016. Salt stress manifestation on plants, mechanism of salt tolerance and potassium role in alleviating it: a review. Zemdirbyste, 103(2): 229-38.
- Zidan, I., B. Jacoby, I. Ravisna and P.M. Neumann, 1991. Sodium does not compete with calcium in saturating plasma membrane sites regulating Na<sup>22</sup> influx in salinized maize roots. Plant Physiol., 96: 331.
- 57. Vigo, C., I. Therios and A. Bosabalidis, 2005. Plant growth, nutrient concentration and leaf anatomy in olive plants irrigated with diluted seawater. J. Plant Nut., 28: 101-102.
- Melgar, J.C., M. Benlloch and R. Fernández-Escobar, 2006. Calcium increases sodium exclusion in olive plants. Sci. Hortic., 109: 303-305.
- Fernández-Escobar, R., 2019. Olive Nutritional Status and Tolerance to Biotic and Abiotic Stresses. Front. Plant Sci., 10: 1151. doi: 10.3389/fpls.2019.01151.

- Rinaldelli, E. and S. Mancuso, 1996. Response of young mycorrhizal and nonmycorrhizal plants of olive tree (*Olea europaea* L.) to saline conditions.
  I. Short-term electro- physiological and long-term vegetative salt effects. Adv. Hort. Sci., 10: 126-134.
- Melgar, J.C., Y. Mohamed, N. Serrano, P.A. García-Galavís, C. Navarro, M.A. Parra, M. Benlloch and R. Fernández-Escobar, 2009. Long term responses of olive trees to salinity. Agric. Water Manage., 96: 1105-1113. Doi: 10.1016/j.agwat.2009.02.009.
- 62. Robinson, F.E., 1987. Growth potential of young olive with high chloride irrigation water. HortScience, 22: 509.
- 63. Ben Ahmed, C., B. Ben Rouina, S. Sensoy and M. Boukhriss 2009. Saline Water Irrigation Effects on Fruit Development, Quality and Phenolic Composition of Virgin Olive Oils, cv. Chemlali. J. Agric. Food Chem., 57(7): 2803-2811. DOI: 10.1021/jf8034379.
- Stefanoudaki, E., M. Williams, K. Chartzoulakis and J. Harwood, 2009. Olive Oil Qualitative Parameters after Orchard Irrigation with Saline Water. J. Agric. Food Chem., 57(4): 1421-1425. DOI: 10.1021/jf8030327
- 65. Petridis, A., I. Therios, G. Samouris and C. Tananaki, 2012. Salinity-induced changes in phenolic compounds in leaves and roots of four olive cultivars (*Olea europaea* L.) and their relationship to antioxidant activity. Environ. Exp. Bot., 79: 37-43. https://doi.org/10.1016/j.envexpbot.2012.01.007.
- 66. Cavaca, L.A., I.M. López-Coca, G. Silvero and C.A. Afonso, 2020. The olive-tree leaves as a source of high-added value molecules: Oleuropein In Studies in Natural Products Chemistry: Bioactive Natural Products. Ed., Atta-Ur-Rahman, Elsevier: Amsterdam, The Netherlands, 64: 131-180.
- 67. Demir, S. and H. Cetinkaya, 2020. Effects of saline conditions on polyphenol and protein content and photosynthetic response of different olive (*Olea europaea* L.) cultivars. Appl. Ecol. Environ. Res., 18(2): 2599-2610. http://dx.doi.org/10.15666/aeer/1802 25992610.

- 68. Farag, R.S., G.S. El-Baroty and A.M. Basuny, 2003. The influence of phenolic extracts obtained from the olive plant (cvs. Picual and Kronakii), on the stability of sunflower oil. Int. J. Food Sci. Technol., 38: 81-87.
- Blasi, F., E. Urbani, M.S. Simonetti, C. Chiesi and L. Cossignani, 2016. Seasonal variations in antioxidant compounds of *Olea europaea* leaves collected from different Italian cultivars. J. Appl. Bot. and Food Qual., 89: 202-207. DOI:10.5073/JABFQ.2016.089.025.
- Olmo-García, L., A. Bajoub, S. Benlamaalam, E. Hurtado-Fernández, M.G. Bagur-González, M. Chigr, M. Mbarki, A. Fernández-Gutiérrez and A. Carrasco-Pancorbo, 2018. Establishing the Phenolic Composition of *Olea europaea* L. Leaves from Cultivars Grown in Morocco as a Crucial Step Towards Their Subsequent Exploitation. Molecules, 23(10): 2524. doi: 10.3390/molecules23102524. PMID: 30279368; PMCID: PMC6222472.
- Nicolì, F., C. Negro, M. Vergine, A. Aprile, E. Nutricati, E. Sabella, A. Miceli, A. Luvisi and L. De Bellis, 2019. Evaluation of Phytochemical and Antioxidant Properties of 15 Italian *Olea europaea* L. Cultivar Leaves. Molecules, 24(10):1998. DOI: 10.3390/molecules24101998.PMID: 31137706; PMCID: PMC6572269.
- 72. Pasković, I., I. Lukić, P. Žurga, V.M. Germek, M. Brkljaèa, O. Koprivnjak, N. Major, K. Grozić, M. Franić, D. Ban, Š. Marcelićm and S.G. Ban, 2020. Temporal variation of phenolic and mineral composition in olive leaves is cultivar dependent. Plants, 9(9) https://doi.org/10.3390/plants9091099
- 73. Ben Ahmed, C., S. Magdich, B. Ben Rouina, M. Boukhris and F. Ben Abdullah. 2012. Saline water irrigation effects on soil salinity distribution and some physiological responses of field grown Chemlali olive. J. Environ. Manage., 113: 538e544.
- Sanzani, S., L. Schena, F. Nigro, V. Sergeeva, A. Ippolito and M.G. Salerno, 2012. Abiotic diseases of olive. J. Plant Path., 94(3): 469-491.