American-Eurasian J. Agric. & Environ. Sci., 21 (1): 22-38, 2021 ISSN 1818-6769 © IDOSI Publications, 2021 DOI: 10.5829/idosi.aejaes.2021.22.38

Morphological and Productivity of Some Olive Genotypes Derived from a Breeding Program and Comparing with Their Parental Cultivars

M.A. Omran

Department of Olive and Semi-Arid Zone Fruits, Horticulture Research Institute, Agriculture Research Centre, Giza, Egypt

Abstract: An olive breeding program was initiated through the project of "Genetic improvement of Olive" (CFFC) IOC in 1994 aimed to obtain new cultivars from cross breeding between several local and foreign cultivars with desirable characteristics. The genotypes were planted by seedlings and established in the olive collection farm at Horticulture Research Institute Giza, Egypt. The present investigation was conducted during three successive seasons (2017, 2018 and 2019) to evaluate twelve of these genotypes (20 years old) and comparing these genotypes traits with the traits of their parental cvs., [Aggizi and Toffahi (the main table olive cvs., 'Egypt'); Chemlali (olive oil cv. 'Tunisia'); Arbequin (oil cv. and Manzanillo dual cv. 'Spain') as well as Kalamata (table olive cv. 'Greece')], to find out the most important characteristics of these genotypes comparing with parental and selecting the most superior ones that meeting the international market requirements and suitable for planting in this region. The genotypes and parental cvs. have been analyzed for traits of the olive tree according to the different parts of the tree (shoot, leaf, floral and fruits) in addition to fruit moisture, oil content and rooting ability. Herein, the greatest values of most the studied traits were significantly cleared in genotype 14. The earliest date of flowering fulfilled by Toffahi cv. (3rd season); whereas, Arbequin cv. was the latest one (1st season). Moreover, other genotypes and parental cvs. showed the greatest total number of flowers and perfect flowers/inflorescence in genotypes (13 & 14); the highest percent of perfect flowers and the minimal of staminate (male) flowers (14) during the experimental seasons. The highest fruit set % was clear in genotypes (14 & 79). A group of genotypes that derived from (Arbequin x Aggizi) gave utmost the highest records of yield crop and classificated as high oil content. Generally, we can classificated the tested genotypes as follow: the genotypes 15 and 68 as table olives, while the genotypes 53 and 69 for dual use and the genotypes 14, 79, 85 and 86 for oil production. In addition, the classification of rooting ability percent helps to divide the types of cuttings into easy, medium and hard to root. As that Arbequin cultivar was the highest in the ability to rooting %.

Key words: Olive • Evaluation • Genotypes • Cross breeding • Flowering • Fruiting • Yield • Oil and rooting

INTRODUCTION

Olive (*Olea europaea*) is one of the most important fruit species that thrive successfully on many arid and semi-arid lands and play an important role in the economy of many countries not only increases the land values where the soil is unsuitable for other crops, but also contributes to soil conservation. It helps to compact problems of environment and that are currently of concern to nation authorities and international organization [1, 2]. Three types of olive threes exist: for oil production, for table olives, for dual use (both oil and table) that have been generated from variability of olive germplasm, which accounted for more than 2000 cultivars [3]. Due to the technological advancement for harvesting the olive, the changes in agricultural policies and market liberalization, the olive research institutions in some producingcountries working to perform several breeding programs which searching for interesting genotypes with a high ecological plasticity, adaptable to new agronomical techniques, capable of producing high quality oil and for big table olive with good flavors and nutritional value of the olive product [4, 5]. In order to select new interesting genotypes, most of breeding programs have been focused on cross breeding among the main outstanding cultivars and selection within the progenies to increase the genetic variability [6, 7]. Evaluation of olive oil composition is considered as a compulsory task in any breeding program aiming at obtaining new olive cultivars [8].

In Egypt, olive is cultivated from ancient centuries. It is found in pharaoh's tombs and temples as pictures and fruits. Nowadays, olive trees play an important role in orchard establishment especially in new reclaimed areas that considered suitable for olive plantings [9]. So, the olive sector represents one of the most promising sectors in Egypt. As a result of increasing the local consumption of oil due to the awareness about the value of health and nutrient and failed of some fruit trees to succeed in the desert because of water salinity, For these reasons, the breeding programs are currently being carried out to obtain new olive cultivars with some of preferable traits [4]. The Olive breeding program has been initiated in Egypt since 1994 in horticultural research institute, by crossing between local and foreign cultivars to obtain new olive cultivars with some of preferable traits of oil and table cultivars. The objective of the breeding program was to improve the qualities of these cultivars and to obtain new olive cultivars with some of preferable traits such as early bearing, high productivity and oil content, resistance to pest and diseases, vigor suitability for mechanical harvesting and high quality of olive oil [10].

Therefore, the present study aimed to evaluate some morphological; floral and fruit characteristics; yield and oil content as well as rooting ability % of twelve olive genotypes derived through Genetic improvement of Olive (CFFC) IOC, of Horticulture Research Institute in Egypt and comparing these genotypes traits with the traits of parental cultivars to find out the most important desirable traits of these genotypes.

MATERIALS AND METHODS

This study was carried out through three growing seasons (2017, 2018 & 2019) of twelve olive genotypes derived through the project of "Genetic improvement of Olive" (CFFC) IOC, of Horticulture Research Institute in Egypt and comparing these genotypes traits with the traits of 6 parental cultivars [Aggizi and Toffahi (the main Egyptian table olive Cvs.); Chemlali olive oil Cv. (Tunisia); Arbequin oil Cv. and Manzanillo dual Cv. (Spain) as well as Kalamata table olive Cv. (Greece)]. The aim of the study was to find out the most important desirable characteristics of these genotypes that will be improving the quality of the fruits and productivity.

The seedlings of the selected genotypes were 20 years-old; planted in the olive collection farm of Horticulture Research Institute, Giza, Egypt at 4×4 m apart in clay loamy soil, under drip irrigation system by Nile water. The plants were grown under the same geographical conditions, received regularly the recommended cultural practices and free from pathogens and physiological disorders. The sources of genotypes according to the project map of Olive improvement program are cleared in Table (1).

Soil chemical; physical characteristics and water chemical properties were determined by Soil, Water and Environmental Res. Inst. Agric. Res. Center, according to the methods as described by Jackson [11] and was summarized in Tables (2 & 3).

Meteorological Data: Temperature and relative humidity data at location was obtained by the National Meteorology Laboratory, Ministry of Agriculture.

Morphometric characteristics were used of the 12 olive genotypes and parental cultivars according to the different parts of the olive tree (shoot, leaf, floral characteristics and fruit set, fruit characteristics; moisture and oil content), in addition to rooting ability, for comparing these genotype traits with the traits of parental cultivars to select the most superior ones that meeting the international market requirements and suitable for planting in this region.

Vegetative Characteristics: The following characters were addressed by using the methodology for primary and secondary characterizations of olive:

Shoot Characteristics: Twenty shoots (one-year-old) were randomly selected around each tree canopy and labeled to study:

Shoot length (cm).

Shoot thickness (cm).

Number of internodes/shoot.

Internodes length (cm): the internodes length on the labeled shoots was record in late of March.

Leaves characteristics:

Leaves density: calculated as a number of leaves per meter.

Leaf length (cm).

Leaf width (cm).

Am-Euras. J. Agric. & Environ	n. Sci., 21	(1): 22-38,	2021
-------------------------------	-------------	-------------	------

Tree number of the Genotypes project map	Crossing combination	Receptors
13	Chemlali x Toffahi	Chemlali 9
14	Chemlali x Toffahi	Chemlali 9
15	Aggizi x Arbequin	Aggizi 9
53	Manzanillo x Aggizi	Manzanillo ♀
68	Toffahi x Kalamata	Toffahi ♀
69	Toffahi x Kalamata	Toffahi ♀
75	Arbequin x Aggizi	Arbequin ♀
77	Arbequin x Aggizi	Arbequin ♀
79	Arbequin x Aggizi	Arbequin ♀
81	Arbequin x Aggizi	Arbequin ♀
85	Arbequin x Aggizi	Arbequin ♀
86	Arbequin x Aggizi	Arbequin ♀

Table 1: Sources of genotypes according to the project map of olive improvement program

Table 2: Physical and chemical properties of the soil under study

Property	Value	Property	Value
Sand (%)	27.48	Available micronutrients (mg kg ⁻¹)	
Silt (%)	34.22	Fe	6.71
Clay (%)	38.30	Mn	6.52
Texture	Clay loam	Zn	4.68
CaCO ₃ gkg ⁻¹ ₃	45.6	Soluble ions (meq/L)	
EC (dS m ⁻¹)	2.92	Ca ⁺⁺	13.8
pH (1:2.5) susp.	7.88	Mg++	10.5
Organic matter (%)	2.29	Na ⁺	4.6
Available macronutrients (mg kg ⁻¹)	K ⁺	0.70
N	33.30	HCO ₃ -	5.8
Р	5.50	Cl	8.0
K	360	SO_{4}^{-}	15.8

Table 3: The chemical analyses of the tested water sample (Nile water) collected from the experimental area

		Cations (meq/L)			Anions (Me	eq/L)		
E.C (dS/m)	pН	Ca ⁺⁺	Mg ⁺⁺	Na ⁺	 K ⁺	HCO ₃ -	Cl ⁻	SO4-	SAR
0.55	7.84	1.50	1.53	1.32	0.18	1.40	1.40	1.73	1.07
Some macro mic	ro nutrients (ppn	n)							
N	Р	K	Fe	Mn	Zn	Cu	Pb	Ni	В
1.36	0.54	7.02	0.02	0.04	0	0.04	0.01	0.01	0.07

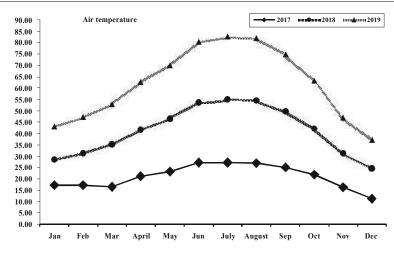
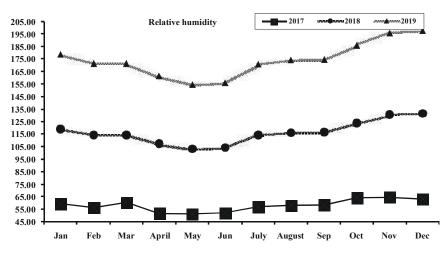


Fig. 1: Average air temperature from the experimental area during the three experimental seasons, 2017, 2018 and 2019



Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

Fig. 2: Relative humidity from the experimental area during the three experimental seasons, 2017, 2018 and 2019

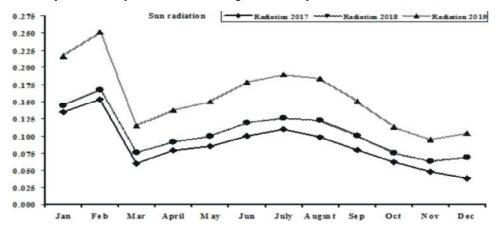


Fig. 3: Sun radiation degree from the experimental area during the three experimental seasons, 2017, 2018 and 2019

Leaf surface area (cm²): Samples of approximately 40 adult leaves taken from the middle section of one year old shoots to determine average leaf surface area, according to following equation:

Leaf area =0.53 (length x width) +1.66 [12]. Leaf shape index (L/W): Elliptic (< 4), Elliptic, -lanceolate (4-6) and lanceolate (> 6), according the characterization of IOC [13].

Floral Characteristics and Fruit Set %:

Flowering date and duration:

Start of flowering date: when 10-25% of flowers were opened.

Full bloom date: when 50-80% of flowers were opened. End of flowering date: developed when 25% of set fruits. Flowering period: was calculated by the days between beginning of flowering and end of blooming. Flowering density: calculated as a number of inflorescences/meter.

Inflorescence length (cm): short (< 2.5), medium (2.5-3.5) and long (> 3.5) according to IOC [14].

Total number of flowers/inflorescence: Total number of flowers per inflorescence was counted and characterization according the IOC [14] into Low (< 18), medium (18-25), high (> 25).

Number of perfect flowers/inflorescence.

Number of staminate flowers/inflorescence.

Perfect flowerer percent: calculated according to Hegazi and Stino, [15]; Rallo and Fernández-Escobar [16] and Hegazi [17] as the following equation:

Perfect flower percentage= $\frac{\text{No. of perfect flowers}}{\text{No. of total flowers}} x100$

Fruit Set (%): Fruit set was calculated after 60 days from full bloom according to Hegazi and Hegazi [18] and Hegazi [19] as a formula:

Fruit set $\% = \frac{\text{No. of fruits}}{\text{No. of total flowers}} x100$

Yield (Kg/tree): Fruits were harvested during ripe stage (pigmentation on more than 50% of the skin) and the average tree yield of each genotype was calculated.

Fruit Characteristics: Thirty of fresh olive fruits were randomly hand-picked from the evaluated genotypes to determine fruit and seed parameters according to the International Olive Council standard method IOC, [14] as the following categories:

Fruit, seed and flesh weight (g).

Fruit polar length (mm); Fruit cross-sectional width (mm); and fruit shape index Fruit shape: determined according the ratio between the length and the width (L/W) as follow: Spherical (< 1.25), ovoid (1.25-1.45), elongated: (> 1.45) [20].

Seed polar length (mm); seed cross-sectional; seed shape index (L/W) and flesh/seed ratio, flesh/fruit (%) and seed/fruit (%).

Moisture content in the fruit: samples of 100 g of whole olives, desiccated in the oven at 105°C for 24 hours to tabulate the moisture content [13].

Percentage of oil content in the fruit (fresh and dry weight basis): samples were determined according to the A.O.A.C. [21].

The oil percent as dry weight is described as very low < 30, Low 30-40, medium 40-50, high 50-60 and very high > 60 [20].

Rooting Ability of Evaluated Genotypes Cuttings %: Sub-terminal cuttings of the selected twelve olive genotypes were taken (in the middle of spring or late summer) about (12-15 cm) long one-year shoots/season. Two pairs of terminal leaves on each cutting were retained and the basal cut was made just below the node. The cuttings were dipped in the solution of IBA at 4000 ppm for few seconds according to Kurd et al. [22] and Shereen [23] after that treated with benlate solution (3g /L) as fungicide. Then planted to a depth of 5 cm in a plastic box filled with a mixture of vermiculite and sand (1:2 volume), on rooting bench provided with basal heating and with mist system. After 70 days of planting, the cuttings were carefully excavated out of media and rooting percentage for each genotype was measured [24]. Rooting percent was described as: nil 0, very low 1-20, low 20 - 40, medium 40 -60, high 60 - 80 and very high 80 - 100 [20].

General Evaluation of the Twelve Evaluated Genotypes and Parental Cultivars under Egypt Condition: The final evaluation was calculated on basis of 100 units, which were shared between main growth, yield and fruit quality characteristics which were (5) units for No. of inflorescences/m, (5) units for perfect flowers (%), (15) unit for fruit set /m, (10) units for fruit weight (g), (5) units for flesh/fruit weight, (30) units for the total yield/tree and (30) units for fruit oil (%) (dry weight basis) according to Elhusseiny [25].

Statistical Analyses: The experiment was arranged in a randomized complete blocks design and the obtained data were subjected to analysis of variance and significant differences among means were determine according to Snedecor and Cochran [26]. In addition significant differences among means were distinguished according to the Duncan multiple tests range Duncan [27].

RESULTS AND DISCUSSION

Morphological traits, yield and fruit characteristics during (2017, 2018 and 2019 years) of twelve olive genotypes and their parental cultivars were evaluated as follow:

Vegetative Characteristics Shoot Characteristics

Shoot Length (cm): The values of shoot length significantly differed according to olive genotypes and parental cultivars during three studied seasons. As shown in Table (4), it was cleared that olive genotypes 68 (Toffahi x Kalamata) recorded the highest values of shoot length than other genotypes and superior on parental cultivars. On the other side, the shortest shoot length was observed by olive genotype 14 (Chemlali x Toffahi) comparing with other genotypes and parental cultivars. This was true during seasons of study.

Shoot Thickness (cm): The maximum thickness of shoots (Table 4), is concomitant to olive genotype 53 (Arbequin x Aggizi) in both first and third season shared with Aggizi and Manzanillo cvs. in the first and second season respectively, following with genotype 15 in the first and second seasons. On the other side, there were an intermediate records represented between genotypes and their parental cultivars, the minimal thinnest values were recorded by genotype 81 during three studied seasons.

Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

Table 4: Shoot length (cm), shoot thickness (cm), No. of internodes/shoot and No. of internodes/meter of twelve olive genotypes and parental cultivars during 2017. 2018 and 2019 experimental seasons

	Shoot leng			Shoot thi		,		ternodes /s			les length	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	28.00B	26.40 E	32.10B	3.00B	2.90C	2.90D	15.60J	14.60K	14.60L	1.79E	1.81B	1.96B
14	15.40 J	19.20M	20.40N	2.80D	2.90C	2.80E	12.60N	16.05I	17.33H	1.22L	1.24IJ	1.18P
15	23.85 G	25.60F	27.82E	3.05AB	3.00B	2.90D	11.40Q	12.45N	14.18M	2.09A	2.06A	2.20A
53	25.30 DE	27.60C	29.25C	3.10A	3.00B	3.20A	12.200	13.40L	15.30J	2.07B	2.06A	1.91C
68	29.40 A	30.20A	32.80A	2.50G	2.60F	2.70F	21.70B	23.20A	22.70B	1.39J	1.30GH	1.44I
69	26.40 C	25.10G	28.65D	2.60F	2.50G	2.60G	22.35A	22.18B	23.27A	1.18M	1.18K	1.23N
75	25.40 D	23.60IJ	26.40HI	2.70E	2.50G	2.70F	21.05C	19.30E	21.63C	1.21L	1.22I-K	1.22N
77	22.70 H	23.90H	25.30K	2.40H	2.70E	2.60G	19.05F	20.30D	21.08D	1.21IL	1.19JK	1.29M
79	23.50 G	22.70L	26.10J	2.60F	2.50G	2.60G	18.25F	17.10G	19.63C	1.29K	1.33FG	1.33L
81	25.10 DE	28.20B	27.55F	2.30I	2.40H	2.50H	16.35I	19.30E	17.23H	1.54G	1.46E	1.60H
85	24.30 DE	23.40J	26.75G	2.70E	2.50G	2.70F	15.15K	15.15J	15.15JK	1.60F	1.54D	1.77E
86	25.30 DE	23.90H	26.65GH	2.80D	2.90V	2.90D	17.85G	18.70F	19.38F	1.42I	1.28G-I	1.38K
Chemlali	22.40 H	23.60IJ	24.60L	2.90C	3.00B	2.90D	14.45L	14.55K	15.10K	1.55G	1.62C	1.63G
Aggizi	21.22 I	23.70HI	26.24IJ	3.10A	2.80D	3.00C	11.70P	12.98M	14.60L	1.81D	1.83B	1.86D
Toffahi	26.20 C	27.30D	29.10C	2.50G	2.60F	2.65FG	22.36A	23.18A	23.24A	1.19M	1.20JK	1.200
Arbequin	24.90 E	22.94K	24.64L	2.90C	2.70E	2.70F	17.30H	16.75H	17.90G	1.44H	1.37F	1.38K
Kalamata	25.42 D	25.66F	22.75M	2.51G	2.60F	2.58G	20.80D	21.10C	16.17I	1.22L	1.27HI	1.41J
Manzanillo	24.90 E	23.10K	26.65GH	3.00B	3.10A	3.10B	13.10M	12.90M	15.21JK	1.90C	1.79B	1.75F

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level

As comparing genotype 53 with parental cultivars it could be noticed that, the genotype 53 which recorded the maximum shoot thickness was superior to Arbequin cultivar during three studied seasons and Aggizi cv. in the second and third seasons.

Number of Internodes/Shoot: Number of internodes/shoot cleared vast variability between the tested genotypes and parental cultivars (Table 4). Toffahi cultivar (as a mother) acquired the highest number of internodes/shoot and this was reflected to the coupled genotypes 68 and 69 (Toffahi x Kalamata), while the least values was reflected to genotype 15 during studied seasons.

Internodes Length (cm): Data presented in Table (4) exhibit significant mark variation in internodes length among olive genotypes under study. Genotypes 15 appeared the maximum values of internodes length in the three studied seasons respectively with partnership with genotype (53) in the second season. Otherwise, the minimum of internodes length was differed from season to another and attained by genotype (69) in both first and second seasons and genotype (14) in the third one. As comparing theses genotypes with their parental cultivars, the genotype (15) was surpassed than (Aggizi &

Arbequin) cultivars. On the other side, the shortest internodes length of Toffahi cv. (as a receptor) consequently reflected on genotypes 14 and 69 which exhibit the minimal one.

Leaves Characteristics

Average Number of Leaves/Meter: It is quite clear from the tabulated data of three studied seasons in Table (5) that, the genotype 77 and its parental cultivars (Arbequin & Aggizi) attained the highest values in the three studied seasons, shared with genotype 69 in the first season. The reverse was true with genotype 15 and 53 which recorded significantly the lowest value through the three studied seasons.

Leaf Length (cm): Regarding to the tabulated data in Table (5) that Kalamata cv. gave the highest leaf length in three studied seasons. On the contrary, the shortest leaf length was in concomitant to Manzanillo shared with Aggizi, Toffahi and Arbequin olive cultivars in three seasons.

Leaf Width (cm): Data in Table (5) shows obviously considerable variations in this respect, herein, the greatest values of leaf width was cleared in olive genotype 13 in the studied seasons, whereas the genotype 81, Toffahi and Manzanillo cultivars achieved the least width.

Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

	No. of lea			Leaf leng			Leaf wid				ice area (ci	'	Leaf shap	be index	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	111.51L	110.61J	119.63M	6.05DE	5.76G	6.40C	2.10A	2.16A	2.22A	6.12EF	6.50C	6.2H	2.88K	2.66G	2.88G
14	163.70C	167.20B	166.61B	5.86FG	5.96E	6.06EF	1.80C	1.85DE	1.90D	6.15EF	6.41D	6.43J	3.26IJ	3.22E	3.19F
15	95.60O	97.27L	104.90P	6.15CD	6.23CD	6.31CD	1.96B	1.98C	2.00C	6.23 DE	6.35E	6.84C	3.14J	3.15EF	3.16F
53	96.450	97.10L	101.62Q	6.05DE	5.78G	7.11A	1.79C	1.89D	1.99C	6.05F-H	6.12GH	6.72F	3.39HI	3.05F	3.57D
68	144.23G	153.64E	138.41I	6.65B	6.71B	6.77B	2.00B	2.07B	2.14B	6.52C	6.69B	6.97F	3.33HI	3.24E	3.17F
69	169.32A	168.13AB	162.41D	6.21C	6.23CD	6.25D	1.97B	1.98C	1.99C	6.23DE	6.50C	6.85C	3.15J	3.15EF	3.14F
75	165.75B	163.57C	163.82C	5.95EF	5.96E	5.97FG	1.60E	1.68G	1.76FG	6.06F-H	6.10H	6.82D	3.73CD	3.5D	3.39E
77	167.85A	169.87A	169.89A	5.80G	5.92EF	6.00F	1.30H	1.40K	1.50I	6.12EF	6.15G	6.55G	4.48A	4.24A	4.00A
79	155.32D	150.66F	150.38E	6.10CD	6.12D	6.14E	1.65DE	1.68G	1.71G	6.15EF	6.35E	6.75E	3.70D-F	3.64D	3.59D
81	130.28J	136.88H	125.05K	5.75GH	5.82FG	5.89GH	1.40G	1.46J	1.52HI	9.09A	6.12GH	6.32K	4.11B	3.99B	3.88AB
85	124.69K	138.49H	128.04J	5.65HI	5.68GH	5.71IJ	1.50F	1.54I	1.58H	5.97H	6.10H	6.47I	3.77CD	3.69D	3.61D
86	141.11H	156.49D	145.40G	6.15CD	6.26C	6.37C	1.70D	1.72G	1.74G	6.27D	6.38DE	6.81D	3.62EF	3.64D	3.66CD
Chemlali	129.04J	123.31I	122.76L	5.75GH	5.77G	5.79HI	1.50F	1.60H	1.70G	6.12EF	6.27F	6.29L	3.83CD	3.62D	3.41E
Aggizi	105.56N	96.20L	106.14O	5.55IJ	5.59HI	5.63JK	1.70D	1.78F	1.86DE	6.00GH	6.30F	6.10M	3.26IJ	3.14EF	3.03FG
Toffahi	153.44E	155.31DE	148.45F	5.45JK	5.53IJ	5.61JK	1.45FG	1.50IJ	1.55HI	6.10FG	6.11H	6.09M	3.76С-Е	3.69D	3.62D
Arbequin	147.99F	148.65FG	142.82H	5.55IJ	5.57HI	5.59JK	1.61E	1.71G	1.81EF	5.99GH	6.12GH	6.29L	3.45GH	3.26E	3.09F
Kalamata	134.22I	147.00G	142.11H	6.85A	6.95A	7.05A	1.76C	1.81EF	1.86DE	8.19B	8.29A	8.25A	3.89C	3.84C	3.79BC
Manzanillo	109.24M	107.93K	109.27N	5.35K	5.41J	5.57K	1.50F	1.52IJ	1.54HI	5.96H	6.03 I	6.09M	3.57FG	3.56D	3.62D

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level.

Leaf Surface Area: Leaf surface area calculated the highest values in Kalamata cultivar and genotype 81 in (2018 & 2019) and 2017 respectively. on the contrary, the lowest values of leaf surface area attained by Manzanillo cultivar during the three seasons, Aggizi and Toffahi cultivars in the third season (Table 5).

Leaf Shape Index: Concerning the leaf shape index of twelve genotypes and parental cultivars in Table (5) it could be noticed that, the greatest leaf shape index scored by genotype 77 and 81 during the three studied seasons and the third season respectively, followed by genotype 81 in second and third season. Whereas the least one attained by genotype 13. Since, in most cases the increase in leaf length was relatively higher than leaf width in different olive genotype under study and this could be logically explained on the unparalleled values in leaf shape index with different olive genotypes under study. According to the methodology for primary characterization of olive varieties cited by the International Olive Oil Council (IOC) [13] and by other morphological studies on olive cultivars, all of tested genotype and parental cultivars classificated elliptic (< 4), except genotype 77 was elliptic – lanceolate (4-6).

Differences in growth characteristics between olive genotypes and parental cultivars are in close conformity with the findings previously reported by Pritsa *et al.* [28]; Proietti *et al.* [29] and Mnastrie *et al.* [30].

Floral Characteristics and Fruit Set (%)

Flowering Duration: Data of start, end and duration period of flowering that presented in Table (6) and

Figs. (4, 5 and 6) during the three experimental seasons, cleared that, begging of flowering of tested genotypes and parental cultivars occurred during the period from February 28th to March 21st during three studied season. In this concern, the earliest date of flowering fulfilled by Toffahi cultivar in the 3rd season. Whereas, Arbequin cultivar was the latest one (1st season). Similarly, the end of flowering started at March 19th in Toffahi cultivar (3rd season) till April 11th, in Arbequin cultivar in 2017 season. As regard to the duration period of three studied seasons, it was ranged between (19-24) days. The aforementioned results agree with El-Badawy et al. [31] and Cesaraccio et al. [32] they noticed that, flowering habit differed according to cvs. and varied from one season to another. This may be due, to the differed in its thermal requirement and their physiological status. Moreover, the phonological behavior of olive tree is largely influenced by environmental factors such as temperature.

Flowering Density (No. of Inflorescence /M): There were variations among olive genotypes and parental cultivars during three seasons under study Table (7). The highest value was concomitant to genotype 14 in 2017 and 2019 seasons shared with genotype 81 in 2017 season and Aggizi cultivar in 2018 season, on the other side, the least value was attained by Manzanillo cultivar. This result goes generally with Bellini *et al.* [4]; El-Sayed [10] and Mikhail [9] reported that percentage of perfect flowers differed according to some factors such as cultivar, growing season, leaf to bud ratio, nutritional status and water stress during inflorescence development and vegetative vigor.

Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

Table 6: Begging of flowering, end of flowering and duration period of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

	Begging	of flowering	3	End of f	lowering		Duration p	eriod	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	8/3	7/3	6/3	31/3	29/3	28/3	23 days	21 days	22 days
14	11/3	10/3	9/3	31/3	30/3	29/3	20 days	20 days	20 days
15	6/3	5/3	4/3	26/3	25/3	24/3	20 days	20 days	20 days
53	13/3	12/3	11/3	3/4	2/4	1/4	21 days	21 days	21 days
68	8/3	7/3	6/3	30/3	29/3	28/3	22 days	22 days	23 days
69	10/3	9/3	8/3	29/3	28/3	27/3	19 days	19 days	20 days
75	12/3	11/3	10/3	31/3	30/3	29/3	19 days	20 days	20 days
77	11/3	10/3	9/3	3/4	2/4	1/4	23 days	23 days	24 days
79	13/3	12/3	11/3	5/4	4/4	3/4	23 days	23 days	23 days
81	10/3	9/3	8/3	3/4	2/4	1/4	24 days	24 days	24 days
85	18/3	17/3	16/3	9/4	8/4	7/4	21 days	22 days	22 days
86	16/3	15/3	14/3	7/4	6/4	5/4	23 days	22 days	22 days
Chemlali	13/3	12/3	11/3	2/4	1/4	31/3	20 days	20 days	21 days
Aggizi	6/3	5/3	4/3	28/3	27/3	26/3	22 days	22 days	22 days
Toffahi	2/3	1/3	28/2	21/3	20/3	19/3	19 days	19 days	19 days
Arbequin	21/3	20/3	19/3	11/4	10/4	9/4	21 days	21 days	22 days
Kalamata	16/3	15/3	14/3	8/4	7/4	6/4	23 days	23 days	23 days
Manzanillo	11/3	10/3	9/3	31/3	30/3	29/3	20 days	20 days	20 days

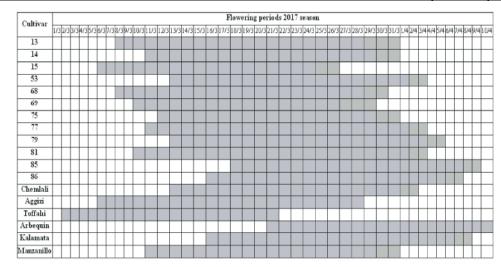


Fig. 4: Time of flowering (start, end and blooming) duration in 2017 season

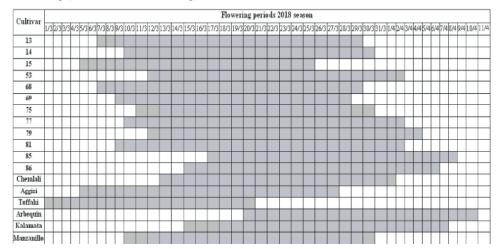


Fig. 5: Time of flowering (start, end and blooming) duration in 2018 season

Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

			_	_	_	_	_	_	_	_	_			_	_	_	_	_	_	F	low	erir	ıg p	erio	ds :	201	9 se	e as o	n		_	 		 _	_	_	_	_	_	_	_	_	_	_
Cultivar	28/2	1/3		3/3	4/3	3 5/	3	6/3	7/3	8/3	9/3	10/3	ıı/s			13/ 3		15 3			17/ 3		19/ 3	20/ 3		22 3		3/ 2/		257 : 3	26/ 3	28/ 3											9/4	10/4
13		Γ	Γ	Γ	Γ	Т	1												t								T	T	T							Γ		Γ			Π			
14					Γ	Τ	Τ												T								T	T	T							Γ					Π			
15		T	T	\square		T								T	1				T								T	T	Г							T		T	T		Π			
53		F	T	T	Г	Т	Т	_						T	+				t							T	t	t	T							F	T	T	T		Π	1		
68	\square	t	t	t	t	t	1							t	+			\square	t	1					\vdash	T	t	$^{+}$	t					Г	Г	t	t	t	t		Π	1		
69		F	t	T	T	t	Т	_		T				T	+				t						\square	T	t	t	T						T	t	T	T	T		Π			
75		t	t	t	t	t	+	_		Г	Г			t	+				t	1					F	t	t	t	+	-				F	t	t	t	t	t		H	+		
77		F	T	T	T	T	1							T	+				t							T	t	t	t							F	T	T	T		Π	1		
79		T	T	T	T	T	1												t								T	T	T					\square	T	t			T		Π	1		
81		t	t	t	t	t	t			t				t	+			\vdash	t	1					F	t	t	t	t	-			\vdash	F	t	Г	Г	t	t		H	+		
85		t	t	t	t	t	t			Г	Г	_		Г	Т				T	1					F	t	t	$^{+}$	†	-			F	F	t	t	t	t				+		
86		t	t	t	t	t	+	_			\square			t	+				t	1						t	t	$^{+}$	+	-				F	t	t	t	t	t		П	+		
Chemlali		t	t	\vdash	t	t	+				\square			t					t	1						\vdash	t	+	+					\vdash	Г	Г	Г	Г	Г		H	+		
Aggizi	\square	t	t	t	t	t	T							T	+			\square	t	1					\vdash	T	t	$^{+}$	t						T	t	t	t	T		Π	1		
Toffahi		t	t		F	t	t	_						t	+				t								Г	Т	Ť					\vdash	t	t	\uparrow	t	t		H	+	-	
Arbequin			Г	Г	Г	T	T							Г	T				T	1							t	t	t							t	t	t	T					
Kalamata		\square	T		t	t	t							T	+				t								T	T	t							T		F			Π	T		
Manzanillo		t	t	T	t	t	t								t				t	1							T	t	t							Γ	Г	Г			H	+		

Fig. 6: Time of flowering (start, end and blooming) duration in 2019 season

Table 7: Flowering density (No. of inflorescences/m), inflorescence length (cm), total number of flowers/inflorescence and number of perfect flowers/ inflorescence of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons.

		ensity (No. of inflo			cence length			ber of flowers/i			ect flowers / in	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	107.9I	111.0JK	101.10	3.12A	3.10A	3.10A	25.20B	23.90B	25.10B	19.85A	17.55B	17.69A
14	176.0A	158.8D	177.7A	3.01C	2.90E	2.99B	23.60D	23.20C	24.45C	20.06A	18.99A	17.53A
15	121.6H	131.6H	130.7K	2.590	2.63I	2.01J	19.40I	19.80H	20.12G	8.95E	10.20F	8.98E
53	102.8J	109.4K	103.6N	2.87H	2.75G	2.86D	22.30F	21.20F	23.75D	14.50B	12.30E	11.95D
68	144.9B	144.4F	137.8I	2.82J	2.87E	2.88D	23.20E	22.60D	20.95F	2.50I	5.70L	5.12I
69	162.9B	162.9C	160.0C	2.65N	2.70H	2.64H	14.60L	15.22K	16.71JK	5.20H	6.40K	5.90H
75	157.9C	159.8B	154.4D	2.95E	3.01BC	3.07A	15.30K	14.90L	14.75N	7.90F	8.10I	7.60FG
77	164.8B	167.8B	161.5B	2.97D	3.10A	2.99B	15.62K	15.90J	16.77J	8.01F	7.99I	7.15G
79	109.4I	113.2J	117.4M	3.03B	3.12A	2.70G	14.14M	14.85L	16.15KL	7.86F	7.23J	6.95G
81	175.7A	142.9F	146.5F	3.00C	3.09A	2.75F	16.16J	17.20I	17.72I	7.75F	8.20I	7.35G
85	148.6D	148.3E	143.7G	2.92F	2.99CD	3.00B	14.85L	14.15M	15.60LM	8.12F	7.32J	8.22F
86	138.3F	135.1G	149.5E	2.89G	2.97D	2.99B	14.72L	15.25K	15.35M	7.95F	6.99J	7.11G
Chemlali	130.4G	125.4I	125.6L	3.12A	3.10A	3.10A	25.70B	26.90A	27.10A	14.60B	15.50C	15.70B
Aggizi	140.4F	179.9A	131.9J	2.590	2.63I	2.10I	20.30H	21.50E	22.36E	8.92E	9.24G	9.24E
Toffahi	130.4G	125.4I	125.6L	2.78L	2.81F	2.86D	27.30A	23.30C	21.20F	9.70D	8.60H	7.40G
Arbequin	120.1H	126.0I	131.1JK	2.69M	3.03B	2.90C	24.22C	23.80B	23.70D	12.20C	12.70D	14.60C
Kalamata	138.4F	144.6F	141.1H	2.80K	2.90E	2.90C	21.60G	20.70G	18.30H	7.30G	8.60H	8.10F
Manzanillo	91.20K	91.33L	100.2P	2.85I	2.72GH	2.82E	22.50F	19.90H	24.60BC	12.20C	9.40G	12.50D

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level

Inflorescence Length (cm): Concerning the inflorescence length in Table (7), Chemlali cultivar (as a mother) acquired the maximum inflorescence length and this was reflected to the genotype 13 which achieved the highest inflorescence length, while genotype 15 that derived from Aggizi x Arbequin has the minimal inflorescence length as the effect of Aggizi cultivar. Moreover, all of genotypes and parental cultivars had medium inflorescence length according to Barranco *et al.* [20]; Laaribi *et al.* [2] and IOC [14].

Total Number of Flowers/Inflorescence: Data in Table (7) illustrated that, the greatest number of total flowers/ inflorescence were detected by Toffahi followed by Chemlali cultivar in the first season, while

Chemlali cultivar was superior in both of second and third season. Moreover, the increases of number of total flowers in genotype 13 and 14 due to genetic makeup of Chemlali cultivar. The reverse was true with group of genotypes 75, 85 and 86 that derived from (Arbequin x Aggizi) which recorded significantly least value.

Similarly according to the division of IOC (2015), the genotype 69 and all of genotypes that derived from (Arbequin x Aggizi) had low number of inflorescence per inflorescence (< 18 flowers), while other genotypes and parental cultivars had medium number of total number of flowers per inflorescence (18 - 25 flowers), except Chemlali cultivar which had high number of flowers per inflorescence (> 25).

Table 8: No of staminate flowers/inflorescence, perfect flowers (%), set fruit/m and yield (kg/tree) of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

	No of stamin	ate (male) flowers/	inflorescence	Perfect flo	owers (%)		Fruit Set /	m		Yield (kg/	tree)	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	4.35K	5.35K	7.46J	82.02B	73.43B	70.34B	33.33D	26.47H	28.18I	45.00C	26.50DE	32.00G
14	3.54L	4.21L	6.92J	85.00A	81.85A	71.70A	59.12A	39.79A	43.64A	48.00A	29.00A	43.00A
15	10.45F	9.60F	11.14DE	46.13H	51.52F	44.63H	29.25H	24.30K	27.69J	28.00J	17.00J	29.00H
53	7.80I	8.90G	11.80CD	65.02C	58.02C	50.32F	33.62D	31.02D	32.27E	44.00D	25.50F	35.00F
68	20.70A	16.20A	15.83A	10.78L	28.32N	24.44M	18.45L	16.59N	19.31L	25.50L	18.00I	29.50H
69	9.40G	9.52F	10.81EF	35.62J	37.45M	35.31L	25.85I	23.26L	28.18I	28.00J	16.00K	25.50L
75	7.40I	6.80J	7.15J	51.63F	54.36D	51.53EF	21.80K	18.84M	18.87M	26.00KL	14.00L	29.00H
77	7.611	7.91HI	9.62GH	51.28F	50.25G	42.64JK	33.07DE	31.02D	33.71D	26.50K	14.00L	26.50K
79	6.28J	7.62I	9.20H	55.59DE	48.69H	43.03IJ	38.33B	33.04B	36.04B	47.50AB	28.00B	43.00A
81	8.41H	9.00G	10.37FG	47.96G	47.67I	41.48K	25.85I	23.02L	28.27I	28.00J	18.00I	26.50K
85	6.73J	6.83J	7.38J	54.68E	51.73F	52.69E	35.39C	32.93B	34.09C	47.00B	27.50BC	42.00B
86	6.77J	8.26H	8.24I	54.01E	45.84J	46.32G	33.60D	28.89F	29.68G	44.50CD	27.00CD	43.00A
Chemlali	11.10E	11.40D	11.40C-E	56.81D	57.62C	57.93D	32.60EF	30.15E	33.65D	36.00F	20.50G	41.00C
Aggizi	11.38E	12.26C	13.12B	43.94I	42.98K	41.32G	31.47G	25.40I	32.57E	34.00G	19.00H	39.50D
Toffahi	17.60B	14.70B	13.80B	35.53J	36.91M	34.91L	31.30G	25.09J	29.04H	32.00H	17.00J	38.00E
Arbequin	12.02D	11.10D	9.10H	50.37F	53.36E	61.60C	32.28F	31.56C	34.00CD	43.00E	26.00EF	40.50C
Kalamata	14.30C	12.10C	10.20FG	33.80K	41.5L	44.26HI	23.09J	14.140	24.09K	28.00J	12.50M	24.00M
Manzanillo	10.30F	10.50E	12.10C	54.22E	47.264I	50.81F	31.06G	28.89F	30.77F	29.00I	19.00H	32.00G

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level.

Number of Perfect Flowers/Inflorescence: As shown in Table (7), genotypes 13 & 14 achieved the maximum records in total number of perfect flowers/inflorescence and superior on the parental cultivars. On the other side, the lowest value was attained by genotype 68. Similarly, other genotypes and parental cultivars were in between the aforesaid extremes.

Total Number of Staminate (Male) Flowers/Inflorescence: Tabulated data that presented in Table (9) demonstrated that the highest number of staminate flowers was detected by genotype 68 during three studied seasons, while the minimal one was achieved in genotype 14.

Perfect Flowerer Percent: Table (8) display obviously that, the highest percent of perfect flowers was statistically detected by genotype 14 comparing with other genotypes and parental cultivars during 2017, 2018 & 2019 experimental seasons. Anyhow, the least number of perfect flowers percent was significantly in concomitant to olive genotype 68 during the experimental seasons. Additionally, there was no apparent effect of parental cultivars on genotypes. In this concern, Osman [33] and Shereen [23] found that the percentage of perfect flowers in olive vary from year to year, tree to tree, shoot to shoot and inflorescence to inflorescence and the flowering density can be considered the major factor affected the percentage of perfect flowers.

Fruit Set (%): Data of the tested genotypes and parental cultivars in Table (8), showed a vast variability in the fruit set (%). Olive genotype 14 acquired the highest fruit set

(%) at the three studied seasons respectively and superiority on the other genotypes and parental cultivars. The least fruit set percent obtained by genotype 68. Similarly, the present result goes partially in the line with that pointed out by Cuevas and Rallo [34]; Ferri *et al.* [35] and El-Badawy *et al.* [31], who reported that differences between olive cvs. in fruit set (%) due to a varying degree of self fertility and cross pollination requirements and percentage of perfect flowers that effects on determining fruit set percentage.

Yield (Kg/tree): Data of the three years fruit yield presented in Table (8) demonstrated that, high differences were found between new olive genotypes and cultivars. Olive genotype 14 acquired the highest fruit yield at the three studied seasons respectively and superiority on the other genotypes and parental cultivars. Arbequin cultivar acquired the highest yield (Kg/tree) comparing with other cultivars and this was reflected to the genotypes 79, 85 and 86 (Arbequin x Aggizi) which achieved the maximum yield and superior on the other genotypes and cultivars. On the other side, the lowest vield was recorded in Kalamata as a table olive cultivar during 2017, 2018 & 2019 seasons. In general, the majority of genotypes that produced from (Arbequin x Aggizi) and (Chemlali x Toffahi) were characterized as a good yield. The aforementioned results goes partially in the line with that pointed out by Mikhail [9]; Yamen et al. [36] and Dridi et al. [37] who pointed that olive yield crop affected by several factors as biennial bearing phenomenon with different levels according to the cultivar genotypes, environmental factors and relatively independent of the number of flowers.

	Fruit weig			Seed weigh			Flesh wei	ght (g)	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	3.67H	3.71H	3.72J	0.57J	0.59J	0.56G-I	3.10H	3.12H	3.16I
14	3.85G	3.92G	3.91I	0.65G	0.63H	0.66EF	3.20G	3.29G	3.25H
15	6.14C	6.12BC	6.16C	0.77E	0.79E	0.80CD	5.37B	5.33B	5.36B
53	4.98D	4.96D	4.99E	0.82D	0.85D	0.86BC	4.16D	4.11E	4.13E
68	4.78E	4.74E	4.81G	0.74F	0.71F	0.74DE	4.04E	4.03E	4.07F
69	4.82E	4.78E	4.86F	0.65G	0.68G	0.69EF	4.17D	4.10E	4.17DE
75	2.54I	2.59I	2.61K	0.52K	0.54L	0.55HI	2.02I	2.05I	2.06J
77	2.22J	2.26J	2.29L	0.59IJ	0.61I	0.61F-H	1.63K	1.65K	1.68L
79	4.24F	4.28F	4.32H	0.63GH	0.59J	0.62F-H	3.61F	3.69F	3.70G
81	2.12K	2.07K	2.20M	0.61HI	0.64H	0.65E-G	1.51L	1.43L	1.55M
85	1.80L	1.87L	1.89N	0.31M	0.37N	0.35J	1.49L	1.50L	1.54M
86	2.25J	2.32J	2.30L	0.52K	0.56K	0.55HI	1.73J	1.76J	1.75K
Chemlali	0.98M	0.96N	0.94P	0.32M	0.350	0.37J	0.66N	0.61N	0.570
Aggizi	6.15C	6.09C	6.17C	0.91C	0.95B	0.94B	5.24C	5.14D	5.23C
Toffahi	8.00A	8.11A	7.95A	1.05A	1.12A	1.08A	6.95A	6.99A	6.87A
Arbequin	1.79L	1.74M	1.840	0.48L	0.52M	0.511	1.31M	1.22M	1.33N
Kalamata	6.28B	6.18B	6.27B	0.96B	0.95B	0.90B	5.32B	5.23C	5.37B
Manzanillo	5.03D	4.99D	5.07D	0.91C	0.87C	0.88BC	4.12D	4.12E	4.19D

Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

Table 9: Fruit weight (g), seed weight and flesh weight of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level.

Table 10: Fruit length (cm), fruit diameter (cm) and fruit shape index of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

	Fruit leng			Fruit diam	()		Fruit shap	e index	
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	2.35F	2.42EF	2.38I	1.78D	1.86G-I	1.78G	1.32E	1.38F	1.34G
14	2.47D	2.53DE	2.55F	1.77D	1.81FG	1.80G	1.40D	1.40E	1.42E
15	2.85A	2.79AB	2.86B	2.15B	2.12B	2.20B	1.33D	1.32G	1.30H
53	2.43DE	2.39F	2.44G	1.89C	1.86EF	1.90F	1.29EF	1.28HI	1.28I
68	2.78B	2.81AB	2.79C	1.92C	1.91C-E	1.93DE	1.45CD	1.47C	1.45D
69	2.65C	2.71BC	2.70E	1.89C	1.91C-E	1.93DE	1.40D	1.42D	1.40F
75	1.97G	1.95G	1.98JK	1.57E	1.51J	1.55I	1.25FG	1.29H	1.28I
77	1.99G	2.00G	1.99J	1.71D	1.74HI	1.72H	1.16HI	1.15L	1.16L
79	2.33F	2.30F	2.40H	1.90C	1.95C	1.96C	1.23G	1.18K	1.22K
81	1.95G	1.98G	1.98JK	1.76D	1.77GH	1.79G	1.11IJ	1.12M	1.11N
85	1.94G	1.96G	1.97K	1.38F	1.40K	1.41J	1.41D	1.40E	1.40F
86	1.98G	1.99G	1.98JK	1.69D	1.71I	1.73H	1.17H	1.16L	1.14M
Chemlali	1.01I	1.00I	0.96M	0.52H	0.51M	0.48L	1.94A	1.96A	2.00A
Aggizi	2.74B	2.59CD	2.71DE	2.17B	2.14B	2.19B	1.26FG	1.21J	1.24J
Toffahi	2.75B	2.83B	2.72L	2.54A	2.51A	2.55A	1.08J	1.09N	1.070
Arbequin	1.66H	1.74H	1.72L	0.89G	0.89L	0.88K	1.87B	1.96A	1.95B
Kalamata	2.87A	2.91A	2.95A	1.91C	1.92CD	1.95CD	1.50C	1.52B	1.51C
Manzanillo	2.41E	2.40EF	2.44G	1.91C	1.89DE	1.91EF	1.26FG	1.27I	1.28I

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level.

Table 11: Seed length (cm), seed diameter (cm) and seed shape index of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

	Seed length (cm)			Seed diameter (cm)			Seed shape index		
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	1.46FG	1.47G	1.48G	0.75HI	0.74F	0.74IJ	1.95D	1.99C	2.00B
14	1.44G	1.43H	1.42H	0.76H	0.75F	0.77GH	1.89F	1.91D	1.84E
15	1.74B	1.72C	1.75B	0.84E	0.82E	0.85E	2.07A	2.10A	2.06A
53	1.72BC	1.75B	1.76B	0.87D	0.85D	0.88D	1.98C	2.06AB	2.00B
68	1.79A	1.77A	1.79A	0.87D	0.88C	0.87DE	2.06A	2.01BC	2.06A
69	1.65D	1.66E	1.68D	0.82F	0.80E	0.82F	2.01B	2.08A	2.05A
75	0.71K	0.72L	0.71N	0.69J	0.71G	0.72JK	1.03N	1.01I	0.99J
77	0.85I	0.86J	0.85L	0.74I	0.75F	0.76HI	1.15L	1.15G	1.12H
79	1.47F	1.49F	1.51F	0.79G	0.76F	0.79G	1.86G	1.96CD	1.91CD
81	0.87I	0.85J	0.88K	0.76H	0.75F	0.77GH	1.14L	1.13G	1.14H
85	0.59L	0.61M	0.620	0.55M	0.57I	0.59M	1.07M	1.07H	1.05I
86	0.79J	0.80K	0.79M	0.67K	0.69G	0.70K	1.18K	1.16G	1.13H
Chemlali	0.78J	0.72L	0.680	0.41N	0.40J	0.37N	1.90F	1.80E	1.84E
Aggizi	1.74B	1.71C	1.72C	0.94B	0.89C	0.91BC	1.85G	1.92D	1.89D
Toffahi	1.80A	1.71C	1.63E	0.99A	0.97A	0.92B	1.82H	1.76E	1.77F
Arbequin	0.99H	0.94I	0.97J	0.65L	0.62H	0.64L	1.52J	1.52F	1.52G
Kalamata	1.55E	1.42H	1.02I	0.95B	0.93B	0.97A	1.63I	1.53F	1.05I
Manzanillo	1.71C	1.69D	1.71C	0.89C	0.88C	0.89CD	1.92E	1.92D	1.92C

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level.

Table 12: Flesh/seed ratio, flesh/fruit (%) and seed/fruit (%) of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

	Flesh/seed ratio			Flesh/fruit (%)			Seed / fruit (%)		
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	5.44F	5.29G	5.64D	84.47D	84.10DE	84.95D	15.53I	15.90IJ	15.05I
14	4.92H	5.22G	4.92F	83.12EF	83.93E	83.12E	16.88GH	16.07I	16.88H
15	6.97A	6.75A	6.70A	87.46A	87.09A	87.01A	12.54L	12.91M	12.99L
53	5.07G	4.84H	4.80G	83.53E	82.86F	82.77E	16.47H	17.14H	17.23H
68	5.46EF	5.68D	5.50E	84.52D	85.02C	84.62D	15.48I	14.98K	15.38I
69	6.42C	6.03C	6.04C	86.51B	85.77B	85.80C	13.49K	14.23L	14.20J
75	3.88K	3.80K	5.75I	79.53H	79.15H	78.93G	20.47E	20.85F	21.07F
77	2.76M	2.70M	2.75I	73.42J	73.01J	73.36I	26.58C	26.99D	26.64D
79	5.73D	6.25B	5.97C	85.14C	86.21B	85.65C	14.86J	13.79L	14.35J
81	2.48N	2.230	2.38M	71.23K	69.08L	70.45K	28.77B	30.92B	29.55B
85	4.81I	4.05J	4.40H	82.78F	80.21G	81.48F	17.22G	19.79G	18.52G
86	3.33L	3.14L	3.18J	76.89I	75.86I	76.09H	23.11D	24.14E	23.91E
Chemlali	2.060	1.74P	1.4N	67.35L	63.54M	60.64L	32.65A	36.46A	39.36A
Aggizi	5.76D	5.41F	5.56DE	85.20C	84.40DE	84.76D	14.80J	15.60IJ	15.24I
Toffahi	6.62B	6.24B	6.36B	86.88B	86.19B	86.42B	13.13K	13.81L	13.58K
Arbequin	2.73M	2.35N	2.61L	73.18J	70.11K	72.28J	26.82C	29.89C	27.72C
Kalamata	5.54E	5.51E	5.97C	84.71CD	84.63CD	85.65C	15.29IJ	15.37JK	14.35J
Manzanillo	4.53J	4.74I	4.76G	81.91G	82.57F	82.64E	18.09F	17.43H	17.36H

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level

Fruit Characteristics

Fruit, Seed and Flesh Weight (g): Fruit characters measurements of the new obtained olive genotypes and their corresponding genitors are given in Tables (9, 10, 11 and 12). Significant differences were observed according to fruit, stone and flesh weight. The highest fruit, seed

and flesh weight value was obtained from Toffahi cultivar as a table olive cultivar, while the Chemlali cultivar as oil cultivar acquired the lowest fruit, seed and flesh weight during three studied seasons. Moreover, the (G85) gave the minimal seed weight in the partnership of Chemlali cultivar in the first and third season.

Fruit weight and flesh weight or percent are considered the most important parameters that used as descriptive fruit characters and thus are required for the new cultivar registration procedure for olive cultivar candidates in breeding studies according the description of fruit weight by Barranco et al. [20], the genotype 85, Chemlali and Arbequin had low fruit weight, the genotypes 13, 14, 75, 77 & 81 had medium fruit weight, the genotypes 53, 68, 69, 79 and manzanillo cultivar had high fruit weight and the genotype 15, Aggizi, Toffahi and Kalamata cultivar had very high fruit weight. On the other hand, Bellini et al. [4] classificated fruit as their weight to: less than (2.5 g) are usually classified as oil and the bigger than (2.5 g) are classified as table olive cultivars. Similarly, flesh percentage, is an important criteria for the classification of olive cultivars. Cultivars with more than 80% flesh are considered as table olive, while cultivars with less than (80%) flesh are classified as oil olive [5].

Fruit Length, Width (cm) and Fruit Shape Index (L/W): As regard to the presented data in Table (10), it could be noticed that, Kalamata olive cultivar significantly appeared the greatest length of fruit (2.87, 2.91, 2.95 cm) followed by genotype 15 (2.85, 2.79 and 2.86 cm) in the three season respectively, Toffahi olive cultivar achieved the maximum fruit width whereas, the least fruit length and width was in concomitant to Chemlali olive cultivar. Meantime, the ratio between the length and the width was calculated to determine the differences among the tested genotypes and parental cultivars in shape, it is appearing the superiority of Chemlali cultivar, otherwise Toffahi cultivar was the minimal one. According the description of IOC [14], the genotypes 77, 79, 81 & 86, Aggizi and Toffahi cultivar had spherical shape; the genotypes 13, 14, 15, 53, 69, 75, 77, 85 and Manzanillo cultivar had ovoid fruit shape and the genotype 68, Chemlali, Arbequin and Kalamata had elongated fruit shape. This result was the parallel with those found by Ozdemir et al. [5] and Yamen et al. [36] who reported that Length, diameter and length/ diameter ratio influence the olive shape which is important in characterization during breeding programs.

Seed Length, Width (cm) and Seed Shape Index (L/W): Data in Table (11) displays obviously that, the genotype 68 was the superior in seed length of the three studied seasons, also Toffahi cultivar had the superiority in the first season. Whereas, the least fruit length was concomitant to genotype 85. Moreover, the highest seed width acquired by Toffahi followed by Kalamata cultivars. However, the least records were attained by Chemlali cultivar. Referring to length/width ratio for seeds conformed to the seed shape index, it was Spherical in genotypes 75, 77, 81 and 86; Ovoid in Toffahi, Arbequin and Kalamata cultivars and Elliptic in genotypes 13, 14, 15, 53, 68, 75 and 79 as well as in Chemlali, Aggizi and Manzanillo cultivars.

Flesh/Seed Ratio, Flesh/Fruit (%) and Seed/Fruit (%): As regard to the presented data in Table (12), the highest value of flesh/fruit weight and flesh/seed ratio was obtained in genotype 15, while Chemlali cultivar achieved the least one. Otherwise, the highest seed/fruit (%) attained by Chemlali cultivar, whereas, the genotype 15 was the minimal one in three studied seasons respectively. In general, many studies on genotypes, revealed that, a high percentage of pulps means a better commercial value for both table and oil production [38, 2, 5].

Moisture Content in the Fruit: Regarding to the moisture content that presented in Table (13), it was demonstrated the superiority of genotype 69 in the three studied season as well as, Toffahi cultivar in the first season in moisture content, but decreased to the minimum in genotype 75 in the first season; Arpequin and Kalamata in the second season and genotype 81 and Chemlali cultivar in the third season. Moreover, the finding of many studies observed that, the moisture content is an important parameter for determined the quantity of oil, a high moisture content indicates at lower oil content [38, 36, 37].

Percentage of Oil Content in the Fruit (Fresh and Dry Weight Basis): Table (13) showed that the highest oil content as fresh weight and dry weight was attained by Chemlali and Arbequin, whereas, the lowest content attained by genotype 14 and genotype 79 for dry weight in the three seasons of study and Chemlali cv. in the 3rd season. While, Toffahi cultivar was the minimal one in fruits fresh and dry weights oil content in all studied seasons. Del Rio and Caballero [39] and Tous and Romero [40] divided olives into three groups (based on oil percentage on dry weight basis) as high (> 46%), moderate (38-46%) and low (< 38%). In the current research, according to the oil percentage on the basis of dry matter, all of genotypes and parental cultivars had the high oil percentage group whereas, genotypes 18, 68 and 69 had moderate oil percentage group and Aggizi and Toffahi cultivars had 'low' oil percentage group. Oil content in olive fruits is affected by several factors, especially genetic competence (Genotype) of the cultivar,

Table 13: Moisture content, oil content (dry weight basis); oil content (fresh weight basis): and rooting ability % of twelve olive genotypes and parental cultivars during 2017, 2018 and 2019 experimental seasons

	Moisture (%)			Oil % (dry weight basis)			Oil % (fresh weight basis)			Rooting ability (%)		
Genotypes and parental cultivars	2017	2018	2019	2017	2018	2019	2017	2018	2019	2017	2018	2019
13	66.95C	64.48H	66.75F-H	53.15B	47.78F	51.47E	18.19G	16.41EF	17.11E	21.50FG	23.00E	23.50F
14	66.75C	64.77GH	66.95FG	54.00A	54.45A	56.91A	19.11A	19.25A	19.59A	19.00K	24.50D	25.50E
15	70.15B	69.15C	73.01B	47.78F	46.29G	51.7E	17.83I	11.78H	11.30G	19.50JK	22.50E	22.50GI
53	64.48DE	67.10D	66.95FG	51.47D	53.15B	51.95E	18.70D	17.69C	18.16C	19.00K	20.00G	21.00JK
68	70.45B	69.15C	68.1D	40.22H	42.59H	47.78F	12.68L	13.14G	12.50F	17.50L	16.00J	15.500
69	72.15A	70.65A	74.35A	40.22H	37.07K	40.65H	11.20N	10.88I	10.43H	16.50M	17.00I	17.50N
75	62.99F	65.45FG	67.10EF	52.02C	47.50F	52.31DE	12.04M	18.22B	17.83CD	19.50JK	21.50F	19.50M
77	64.65DE	66.35DE	68.09DE	53.15B	47.95F	53.95BC	18.29F	17.06D	16.97E	21.50FG	20.00G	20.50KI
79	66.32C	65.95EF	66.75F-H	53.95A	52.92B	56.91A	18.94B	18.70B	19.11B	20.50HI	18.50H	21.50IJ
81	64.05D-F	65.99EF	64.48JK	52.92B	49.86E	53.95BC	17.11J	16.97D	17.83CD	21.00GH	19.00H	22.00HI
85	64.37DE	64.48H	65.75HI	53.17B	52.25C	56.19A	18.94B	18.22B	18.20C	22.00EF	20.00G	20.00LN
86	64.85DE	56.89J	65.37IJ	51.90C	51.95C	54.45B	18.81C	18.81C	18.16C	20.00IJ	20.50G	21.50IJ
Chemlali	65.11D	64.65GH	64.05HI	50.11E	51.18D	56.15A	18.06H	17.19D	17.69D	22.50E	24.00D	23.00FG
Aggizi	70.15B	69.62BC	70.35C	19.85I	22.95L	26.03J	5.930	6.97J	7.72I	53.00C	55.00C	57.00B
Toffahi	71.68A	70.15AB	69.85C	18.08J	19.85M	18.25K	5.12P	5.93K	5.50J	52.00D	55.00C	52.00D
Arbequin	64.37DE	63.16I	65.12IJ	50.03E	49.33E	53.14CD	18.50E	17.01D	18.11C	60.00A	58.00A	58.00A
Kalamata	63.75EF	63.45I	68.09DE	40.35H	38.20J	39.28I	16.97K	16.76DE	17.19E	10.05N	9.92K	10.25P
Manzanillo	63.95D-F	66.66DE	65.95G-I	42.92G	39.36I	41.87G	18.52E	16.97D	17.11E	55.00B	56.00B	53.00C

Means within a column having the same letters are not significantly different according to Duncan's Multiple Range Test at 5% level.

Table 14: List of some morphological descriptors and oil content of the twelve evaluated genotypes and parental cultivars according to (IOC) under Egypt condition

Genotypes and parental cultivars	Leaf shape index	Fruit shape index	Fruit weight	Flesh/ seed	Oil content in dry weight
13	Eleptic- lanceolate	Ovoid	Medium	Medium	High
14	Eleptic- lanceolate	Ovoid	Medium	Medium	High
15	Eleptic- lanceolate	Ovoid	Very high	Medium	Medium
53	Eleptic- lanceolate	Spherical	High	Medium	High
68	Eleptic- lanceolate	Elongated	High	Medium	Medium
69	Eleptic- lanceolate	Ovoid	High	High	Medium
75	Eleptic- lanceolate	Ovoid	Medium	Low	High
77	Lanceolate	Spherical	Medium	Low	Medium
79	Eleptic- lanceolate	Spherical	High	Medium	High
81	Eleptic- lanceolate	Spherical	Medium	Low	High
85	Eleptic- lanceolate	Ovoid	Low	Low	High
86	Eleptic- lanceolate	Spherical	Medium	Low	Medium
Chemlali	Eleptic- lanceolate	elongated	Low	Low	High
Aggizi	Eleptic- lanceolate	Spherical	Very high	Medium	Very low
Toffahi	Eleptic- lanceolate	spherical	Very high	Medium	Very low
Arbequin	Eleptic- lanceolate	elongated	Low	Low	High
Kalamata	Eleptic- lanceolate	elongated	Very high	Medium	High
Manzanillo	Eleptic- lanceolate	ovoid	High	Low	High

climatic conditions and soil type, agricultural practices and harvesting date [41]. In general, the total oil contents of olive cultivars in our study were closely matched the results of the previous studies on international imported cultivars and the result on local cultivars [37, 10, 31].

Rooting Ability %: Rooting ability percentage of the semi-hardwood cuttings taken from tested genotypes and parental cultivars cleared in Table (13) illustrated that the rooting percent varied from 10.05, 9.92 and 10.25 % in Kalamata cv., to reach 60.00, 58.00 and 58.00 % in Arbequin cv. during the three studied seasons. Accordingly, genotypes 68 & 69 and Kalamata cultivar classified as very low, whereas each of Aggizi, Toffahi,

Arbequin and Manzanillo cultivars were classified as medium. As the previous study, the classification of rooting ability percent helps to divide the types of cuttings into easy and hard to root [42, 23].

General Evaluation of the Twelve Evaluated Genotypes and Parental Cultivars under Egypt Condition: Data in Table (15) that obtained from this investigation was selected to a system of numerical evaluation of tested genotype. The final evaluation was calculated on basis of 100 units, which were shared between No. of inflorescences/m, perfect flowers (%), fruit set /m, fruit weight (g), flesh/fruit weight, yield and fruit oil (dry weight basis) (%) characteristics which were specified as

Table 15: General evaluation of the twelve evaluated genotypes and parental cultivars during average three seasons (2017, 2018 and 2019) under Egypt condition Perfect flowers (%) Fruit oil (%) dry weight basis Total units No. of inflorescences/m Fruit set /m Fruit weight (g) Flesh/fruit weight Yield (Kg/tree) % Units 5 % Units 5 % Units 15 % Units 10 % Units 5 Kg/tree Units 30 % Units 30 100 13 106.3 3.12 75.26 4.73 29.33 9.26 3.70 4.61 84.50 4.74 34.50 25.88 50.80 27.65 79 99 14 170.5 5.00 79 52 5.00 47 52 15.00 3 89 4 85 83 39 4 68 40.00 30.00 55 12 30.00 94 53 15 128.0 3.75 47.43 2.98 27.08 8.55 6.14 7.66 87.19 4.89 24.67 18.50 48.59 26.45 72.78 53 110.5 3 24 57 79 3 63 32 30 10 20 4 98 6 21 84 16 4 72 34.83 26.12 52 19 28 41 82 53 68 142.4 4.18 21.18 1.33 18.12 5.72 4.78 5.96 84.72 4.75 24.33 18.25 43.53 23.69 63.88 69 167.9 4.92 36.13 2.27 25.76 8.13 4.82 6.01 89.09 5.00 23.17 17.38 39.31 21.40 65.11 75 1573 4 61 52 51 3 30 19.84 6.26 2.58 3 22 79.20 4 4 4 23.00 17 25 50.61 27 55 66 63 77 169.2 4.96 48.06 3.02 32.60 10.29 2.26 2.81 73.11 4.10 22.17 16.63 51.68 28.13 69.94 79 29.63 29.71 87.2 113.3 3.32 49.10 3.09 35.80 11.30 4.28 5.34 4.81 39.50 54.59 85.67 81 155.0 4.55 45.70 2.87 25 71 2.13 2.66 70.27 3.94 24.00 18.00 52 24 28.43 68.57 8.12 34.14 10.78 85 146.9 4.31 53.03 3.33 1.85 2.31 81.47 4.57 38.83 29.12 53.87 29.32 83.74 86 141.0 413 48 72 3.06 30.72 9 70 2 29 2.86 76.27 4 28 38 17 28.63 52 77 28.72 81 38 Chemlali 127.1 3.73 57.45 3.61 32.13 10.14 0.96 1.20 63.89 3.59 32.50 24.38 52.48 28.56 75.21 84.79 22.94 12.49 136.7 4.01 42.75 2.69 29.81 9.41 6.14 4.76 30.67 23.00 64.01 Aggizi 7.65 Toffahi 127.1 3.73 35.78 2.25 28 48 8 99 8 02 10.00 86.49 4 85 29.00 21.75 18.73 10.19 61.76 32.61 Arbequin 125.7 3.69 55.11 3.47 10.29 1.79 2.23 71.88 4.03 36.50 27.38 50.83 27.67 78.76 Kalamata 1414 4.15 39.85 2 51 20.44 6.45 624 7.78 85.00 477 21.50 16.13 39 28 21.38 63.17 Manzanillo 94.5 2.77 50.76 3.19 30.24 9.55 5.03 6.27 82.37 4.62 26.67 20.00 41.38 22.52 66.92

Am-Euras. J. Agric. & Environ. Sci., 21 (1): 22-38, 2021

shown in Table (14) to: (5) units for No. of inflorescences/m, perfect flowers (%) and flesh fruit weight; (10) units for fruit weight (g); (15) units for fruit set, (30) units for the total yield/tree and fruit oil (%) (dry weight basis). Within each criterion, the genotypes that gave the highest value received the "full mark" for it, i.e. all the units specified for this criterion and the other tested genotypes received lower units calculated. From the tabulated data the genotypes (G14, G79, G85, G53, G13) achieved the highest units, while the genotypes (G68&G69) achieved the least units [25].

REFERAENCES

- Kitsaki, C.K., E. Andreadis and D.L. Bouranis, 2010. Developmental events in differentiating floral buds of four olive (*Olea europaea*, L.) cultivars during late winter to early spring. Flora, 205: 599-607.
- Laaribi, I., A.M. Mezghani and M. Messaoud, 2014. Phenotypic diversity of some olive tree progenies issued from a Tunisian breeding program. European Sci. Jour., 10: 1857-7881.
- Leon, L., R. De La Rosa, D. Barranco and L. Rallo, 2011. Agronomic characterization of 15 selections of the olive cross breeding program of Cordoba, Spain. proceedings of the Second International Seminar olivebioteq. "Biotechnology and quality of olive tree produces around the Mediterranean basin". Eur. J. Sci. Technol., 1: 87-93.
- Bellini, E., E. Giordani, M.V. Parlati and S. Pandolfi, 2002. Olive genetic improvement; thirty years of research Acta Hort., 586: 105-108

- Ozdemir, Y., A. Ozturk, E. Guven, M.A. Nebioglu, N.A. Tangu, M.E. Akcay and S. Ercisli, 2016. Fruit and oil characteristics of olive candidate cultivars from Turkey. Notulae Bot. Hortic. Agrobot. Cluj-Napoca., 44: 147-154.
- Lavee, S., N. Avidan, A. Haskal and A. Ogrodovich, 1996. Juvenility period reduction in olive seedlings a tool for enhancement of breeding. Olivae, 60: 33-41.
- Santos Antunes, A.F., A. Mohedo, I. Trujillo and L. Rallo, 1999. Influence of the genitors on the flowering of olive seedlings under forced growth. Acta Horticulturae, 474: 103-105.
- Rallo, L. 2014. Breeding oil and table olives for mechanical harvesting in Spain. Hort Technology, 24: 295-300.
- Mikhail, E.G., 2015. Behaviour of some olive accessions resulting from an olive improvement program. Annals of Agric. Sci., Moshtohor, 53: 1-16.
- El-Sayed, S.M., 2014. Bio-morphological characterization of some local olive oil clones compared with world cultivars. M. Sc. Thesis Department of Pomology Faculty of Agriculture Cairo University Egypt.
- Jackson, M.L., 1973. Soil Chemical Analysis, Constable and Co. Ltd. Prentice Hall of India Pvt. Ltd. New Delhi, pp: 10-114.
- Ahmed, F.F. and M.H. Morsy, 1999. A new method for measuring leaf area in different fruit species. Minia J. of Agric. & Develop., 19: 97-105.
- IOC, 1997. Méthodologie pour la caractérisation primaire des variétés d'olivier. Projet RESGEN, 97., pp: 10 pages.

- IOC, 2015. Norme commerciale applicable aux huiles d'olive et aux huiles de grignons d'olive. COI/T.15/ NC n°3/Rév. 8.
- Hegazi, E.S. and G.R. Stino, 1982. Dormancy, flowering and sex expression in 20 olive cvs. *Olea europaea* L. under Giza conditions. Acta Agrobotanica, 35: 79-86.
- Rallo and Fernández-Escobar, 1985. Influence of cultivar and flower thinning within the inflorescence on competition among olive fruit .J. Amer. Soc. Hort. Sci., 110: 303-308.
- Hegazi, A.A., 2001. Studies on shot berries formation in olives. Ph.D. Thesis, Fac. Agric., Cairo Univ., Egypt.
- Hegazi, E.S. and A.A. Hegazi, 2005. Floral biology and fruiting. Proceeding of the Sixth Arabian Conference for Horticulture, March 20-22, Suez Canal University Ismailia, Egypt, pp: 48-57.
- Hegazi, A.A., 2007. A comparative study for identification between seven olive cultivars. a-Morphological identification. Egypt. J. Appl. Sci., 22: 164-171.
- Barranco, D., A. Cimato, P. Fiorino, L. Rallo, A. Touzani, C. Castaneda, F. Serafini and I. Trujillo, 2000. Methodology descriptor files. World Catalogue of Olive Varieties., pp: 15-21 publisher International olive council.
- A.O.A.C., 2000. Association of Official Agricultural Chemists. Official Methods of Analysis. 17th ed. Association of Official Analytical Chemists. Published by Washington, D. C., USA, pp: 234.
- Kurd, A.A., I.A. Hussain, S. Awan and I. Ali, 2010. Effect of indole butyric acid (IBA) on rooting of olive stem cuttings. Pakistan Journal of Agricultural Research, 23: 193-195
- Shereen, A.S., 2019. Rooting Ability of Some Olive Genotypes by Sub-Terminal Cuttings. Journal of Horticultural Science & Ornamental Plants, 11: 27-37.
- Hartman, H.T., D.E. Kester, F.T. Davies and R.L. Geneve, 2007. Plant Propagation Principles and Practices. Prentice-Hall, New Jersey, pp: 656.
- Elhusseiny, A.M., 2012. Evaluation of Some Olive Strains. Ph.D. Thesis, Fac. Of Agric. Benha Univ. Egypt.
- Snedecor, G.W. and W.G. Cochran, 1980. Statistical Methods (7th ed) Iowa State Univ. Press, Ames, Lowa U.S.A, pp: 507.
- 27. Duncan, D.B., 1955. Multiple range and multiple F test. Biometrcs, 11: 1-24.

- Pritsa, T.S., D.G. Voyiatzis, C.J. Voyiatzis and M.S. Sotiriou, 2003. Evaluation of vegetative growth traits and their relation to time to first flowering of olive seedlings. Australian Journal of Agriculture Research, 54: 371-376.
- Proietti, P., L. Nasini, L. Reale, T. Caruso and F. Ferranti, 2015. Productive and vegetative behavior of olive cultivars in super high-density olive grove Sci. Agric., 72: 20-27.
- Mnastrie, S.R., O.D. Saddoud, S. Rouz, M. Ben Saleh and A. Ferchichi, 2017. Morphological analysis of the autochthon olive varieties cultivated in the North West of Tunisia, 37: 2286-5314.
- El-Badawy, H.E.M, S.F. El-Gioushy, I. Saadeldin and R. Abo El-Ata, 2019. Evaluation of Some Morphological and Flowering Traits in New Six Olive Genotypes Grown under Egypt Conditions. Asian Journal of Agricultural and Horticultural Research, 3: 1-16.
- Cesaraccio, C., A. Canu; G. Pellizzaro and C. Sirca, 2006. A detailed description of flowering stages in olive tree in relation to side tree crown exposure. 17 conference on Biometeorology and Aerobiology ams.confex.com/ams/pdfpapers/110988.pdf.
- Osman, I.M.S., 2015. Agronomic and qualitative evaluation of some olive selections derived from a breeding program. Annals of Agric. Sci., Moshtohor, pp: 17-32.
- Cuevas, J. and L. Rallo, 1990. Response to cross pollination in olive trees with different level of flowering. Proc. International Symposium on Olive Growing. Acta Hort., 286: 179-182.
- 35. Ferri, A., G. Padula, F. Giordani and F. Billini, 2006. First observations on floral biology of advanced selections of olive obtained by crossing. Proceedings of the Second International Seminar Olive bioteq 2006. "Biotechnology and quality of olive tree produces around the Mediterranean Basin", 1: 127-130
- 36. Yamen, M., S. Mohamed and W. Chourname, 2017. Evaluation of some Productive and technological traits in local and introduced olive cultivars. International Journal of Agriculture& Environmental Science, 11: 2394-2568.
- Dridi, J., M., Fendri, Breton, C.M. and M. Msallem, 2019. Characterization of olive progenies derived from a Tunisian breeding program by morphological traits and SSR markers. Scientia Horticulturae, 236: 127-136.

- Medina, E., A. Morales-Sillero, E.M. Ramirez, Rallo, P.M. Brenes and C. Romero, 2012. New genotypes of table olives: profile of bioactive compounds. International Journal of Food Sci. & Tech., 47(11): 2334-2341.
- Del Rio, C. and J.M. Caballero, 1994. Preliminary agronomical characterization of 131 cultivars introduced in olive germplasm bank of Cordoba in March 1987. Acta Horticulturae 356: 110-115.
- 40. Tous, J. and A. Romero, 1994. Cultivar and location effects on olive oil quality in Catalonia (Spain). Acta Horticulturae, 356: 323-326.
- Abdul-Sadeg, S.M., 2014. Morphological and Molecular Characterization of Libyan Olive Olea europaea L. (42 local and 16 wild type) in comparison to 41 Introduced world cultivars, "Department of Horticulture and Landscape Architecture, Doctorate Thesis. Colorado State University, 119p. n Olive Growing. Acta Hort., 286: 179-182.
- Cirillo, C., R. Russo, M. Famiani and C. Divaino, 2017. Investigation on rooting ability of twenty olive cultivars from southern Italy. Adv. Hort. Sci., 31: 311-317.